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(54) Synthetic plant genes and method for preparation

Synthetische Pflanzengene und Verfahren zu ihrer Herstellung

Gènes synthétiques de plantes et méthode pour leur préparation

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(56) References cited:

EP-A-0 142 924 EP-A-0 223 452  
EP-A-0 275 957 EP-A-0 359 472

- UCLA SYMP. MOL. CELL. BIOL., NEW. SER. vol. 48, 1987, MOLECULAR STRATEGIES FOR CROP PROTECTION, pages 345-353, Alan R. Lila, Inc.; M.J. ADANG et al.: "Expression of a *Bacillus thuringiensis* insecticidal crystal protein gene in tobacco plants"
- PLANT PHYSIOLOGY, vol. 85, 1987, pages 1103-1109; K.A. BARTON et al.: "Bacillus thuringiensis delta-endotoxin expressed in transgenic *Nicotiana tabacum* provides resistance to lepidopteran insects"
- BIOLOGICAL ABSTRACTS/RRM BR35: 107674, & 154TH NATIONAL AMERICAN ASSOCIATION FOR THE ADVANCEMENT OF SCIENCE ANNUAL MEETING, Boston, Massachusetts, US, 11th-15th February 1988; M. ADANG et al.: "Engineering crop plants for insect resistance", & AM. ASSOC. ADV. SCI. ABSTR. PAP. NATL. MEET. O (154). 1988.
- NUCLEIC ACIDS RESEARCH, vol. 17, no. 2, 1989, pages 477-498, IRL Press, Oxford, NL; E.E. MURRAY et al.: "Codon usage in plant genes"

**Description****BACKGROUND OF THE INVENTION**

5 [0001] The present invention relates to genetic engineering and more particularly to plant transformation in which a plant is transformed to express a heterologous gene.

[0002] Although great progress has been made in recent years with respect to transgenic plants which express foreign proteins such as herbicide resistant enzymes and viral coat proteins, very little is known about the major factors affecting expression of foreign genes in plants. Several potential factors could be responsible in varying degrees for the level of protein expression from a particular coding sequence. The level of a particular mRNA in the cell is certainly a critical factor.

[0003] The potential causes of low steady state levels of mRNA due to the nature of the coding sequence are many. First, full length RNA synthesis might not occur at a high frequency. This could, for example, be caused by the premature termination of RNA during transcription or due to unexpected mRNA processing during transcription. Second, full length RNA could be produced but then processed (splicing, polyA addition) in the nucleus in a fashion that creates a non-functional mRNA. If the RNA is properly synthesized, terminated and polyadenylated, it then can move to the cytoplasm for translation. In the cytoplasm, mRNAs have distinct half lives that are determined by their sequences and by the cell type in which they are expressed. Some RNAs are very short-lived and some are much more long-lived. In addition, there is an effect, whose magnitude is uncertain, of translational efficiency on mRNA half-life. In addition, every RNA molecule folds into a particular structure, or perhaps family of structures, which is determined by its sequence. The particular structure of any RNA might lead to greater or lesser stability in the cytoplasm. Structure per se is probably also a determinant of mRNA processing in the nucleus. Unfortunately, it is impossible to predict, and nearly impossible to determine, the structure of any RNA (except for tRNA) in vitro or in vivo. However, it is likely that dramatically changing the sequence of an RNA will have a large effect on its folded structure. It is likely that structure per se or particular structural features also have a role in determining RNA stability.

[0004] Some particular sequences and signals have been identified in RNAs that have the potential for having a specific effect on RNA stability. This section summarizes what is known about these sequences and signals. These identified sequences often are A+T rich, and thus are more likely to occur in an A+T rich coding sequence such as a *B.t.* gene. The sequence motif ATTAA (or AUUUA as it appears in RNA) has been implicated as a destabilizing sequence in mammalian cell mRNA (Shaw and Kamen, 1986). No analysis of the function of this sequence in plants has been done. Many short lived mRNAs have A+T rich 3' untranslated regions, and these regions often have the ATTAA sequence, sometimes present in multiple copies or as multimers (e.g., ATTATATTAA...). Shaw and Kamen showed that the transfer of the 3' end of an unstable mRNA to a stable RNA (globin or VA1) decreased the stable RNA's half life dramatically. They further showed that a pentamer of ATTAA had a profound destabilizing effect on a stable message, and that this signal could exert its effect whether it was located at the 3' end or within the coding sequence. However, the number of ATTAA sequences and/or the sequence context in which they occur also appear to be important in determining whether they function as destabilizing sequences. Shaw and Kamen showed that a trimer of ATTAA had much less effect than a pentamer on mRNA stability and a dimer or a monomer had no effect on stability (Shaw and Kamen, 1987). Note that multimers of ATTAA such as a pentamer automatically create an A+T rich region. This was shown to be a cytoplasmic effect, not nuclear. In other unstable mRNAs, the ATTAA sequence may be present in only a single copy, but it is often contained in an A+T rich region. From the animal cell data collected to date, it appears that ATTAA at least in some contexts is important in stability, but it is not yet possible to predict which occurrences of ATTAA are destabilizing elements or whether any of these effects are likely to be seen in plants.

[0005] Some studies on mRNA degradation in animal cells also indicate that RNA degradation may begin in some cases with nucleolytic attack in A+T rich regions. It is not clear if these cleavages occur at ATTAA sequences. There are also examples of mRNAs that have differential stability depending on the cell type in which they are expressed or on the stage within the cell cycle at which they are expressed. For example, histone mRNAs are stable during DNA synthesis but unstable if DNA synthesis is disrupted. The 3' end of some histone mRNAs seems to be responsible for this effect (Pandey and Marzluff, 1987). It does not appear to be mediated by ATTAA, nor is it clear what controls the differential stability of this mRNA. Another example is the differential stability of IgG mRNA in B lymphocytes during B cell maturation (Genovese and Milicarek, 1988). A final example is the instability of a mutant beta-thalassemic globin mRNA. In bone marrow cells, where this gene is normally expressed, the mutant mRNA is unstable, while the wild-type mRNA is stable. When the mutant gene is expressed in HeLa or L cells in vitro, the mutant mRNA shows no instability (Liu et al., 1988). These examples all provide evidence that mRNA stability can be mediated by cell type or cell cycle specific factors. Furthermore this type of instability is not yet associated with specific sequences. Given these uncertainties, it is not possible to predict which RNAs are likely to be unstable in a given cell. In addition, even the ATTAA motif may act differentially depending on the nature of the cell in which the RNA is present. Shaw and Kamen (1987) have reported that activation of protein kinase C can block degradation mediated by ATTAA.

5 [0006] The addition of a polyadenylate string to the 3' end is common to most eucaryotic mRNAs, both plant and animal. The currently accepted view of polyA addition is that the nascent transcript extends beyond the mature 3' terminus. Contained within this transcript are signals for polyadenylation and proper 3' end formation. This processing at the 3' end involves cleavage of the mRNA and addition of polyA to the mature 3' end. By searching for consensus sequences near the polyA tract in both plant and animal mRNAs, it has been possible to identify consensus sequences that apparently are involved in polyA addition and 3' end cleavage. The same consensus sequences seem to be important to both of these processes. These signals are typically a variation on the sequence AATAAA. In animal cells, some variants of this sequence that are functional have been identified; in plant cells there seems to be an extended range of functional sequences (Wickens and Stephenson, 1984; Dean et al., 1986). Because all of these consensus 10 sequences are variations on AATAAA, they all are A+T rich sequences. This sequence is typically found 15 to 20 bp before the polyA tract in a mature mRNA. Experiments in animal cells indicate that this sequence is involved in both polyA addition and 3' maturation. Site directed mutations in this sequence can disrupt these functions (Conway and Wickens, 1988; Wickens et al., 1987). However, it has also been observed that sequences up to 50 to 100 bp 3' to the putative polyA signal are also required; i.e., a gene that has a normal AATAAA but has been replaced or disrupted 15 downstream does not get properly polyadenylated (Gill and Proudfoot, 1984; Sadofsky and Alwine, 1984; McDevitt et al., 1984). That is, the polyA signal itself is not sufficient for complete and proper processing. It is not yet known what specific downstream sequences are required in addition to the polyA signal, or if there is a specific sequence that has this function. Therefore, sequence analysis can only identify potential polyA signals.

20 [0007] In naturally occurring mRNAs that are normally polyadenylated, it has been observed that disruption of this process, either by altering the polyA signal or other sequences in the mRNA, profound effects can be obtained in the level of functional mRNA. This has been observed in several naturally occurring mRNAs, with results that are gene specific so far. There are no general rules that can be derived yet from the study of mutants of these natural genes, and no rules that can be applied to heterologous genes. Below are four examples:

25 1. In a globin gene, absence of a proper polyA site leads to improper termination of transcription. It is likely, but not proven, that the improperly terminated RNA is nonfunctional and unstable (Proudfoot et al., 1987).

2. In a globin gene, absence of a functional polyA signal can lead to a 100-fold decrease in the level of mRNA accumulation (Proudfoot et al., 1987).

30 3. A globin gene polyA site was placed into the 3' ends of two different histone genes. The histone genes contain a secondary structure (stem-loop) near their 3' ends. The amount of properly polyadenylated histone mRNA produced from these chimeras decreased as the distance between the stem-loop and the polyA site increased. Also, the two histone genes produced greatly different levels of properly polyadenylated mRNA. This suggests an interaction between the polyA site and other sequences on the mRNA that can modulate mRNA accumulation (Pandy and Marzluff, 1987).

35 4. The soybean leghemoglobin gene has been cloned into HeLa cells, and it has been determined that this plant gene contains a "cryptic" polyadenylation signal that is active in animal cells, but is not utilized in plant cells. This leads to the production of a new polyadenylated mRNA that is nonfunctional. This again shows that analysis of a gene in one cell type cannot predict its behavior in alternative cell types (Wiebauer et al., 1988).

40 [0008] From these examples, it is clear that in natural mRNAs proper polyadenylation is important in mRNA accumulation, and that disruption of this process can effect mRNA levels significantly. However, insufficient knowledge exists to predict the effect of changes in a normal gene. In a heterologous gene, where we do not know if the putative polyA sites (consensus sequences) are functional, it is even harder to predict the consequences. However, it is possible that the putative sites identified are dysfunctional. That is, these sites may not act as proper polyA sites, but instead 45 function as aberrant sites that give rise to unstable mRNAs.

45 [0009] In animal cell systems, AATAAA is by far the most common signal identified in mRNAs upstream of the polyA, but at least four variants have also been found (Wickens and Stephenson, 1984). In plants, not nearly so much analysis has been done, but it is clear that multiple sequences similar to AATAAA can be used. The plant sites below called major or minor refer only to the study of Dean et al. (1986) which analyzed only three types of plant gene. The designation of polyadenylation sites as major or minor refer only to the frequency of their occurrence as functional sites in naturally occurring genes that have been analyzed. In the case of plants this is a very limited database. It is hard to predict with any certainty that a site designated major or minor is more or less likely to function partially or completely when found in a heterologous gene such as *B.t.*

PA	AATAAA	Major consensus site
P1A	AATAAT	Major plant site
P2A	ACCAAA	Minor plant site
P3A	ATATAA	"
P4A	AATCAA	"
P5A	ATACTA	"
P6A	ATAAAA	"
P7A	ATGAAA	"
P8A	AAGCAT	"
P9A	ATTAAT	"
P10A	ATACAT	"
P11A	AAAATA	"
P12A	ATAAAA	Minor animal site
P13A	AATTAA	"
P14A	AATACA	"
P15A	CATAAA	"

[0010] Another type of RNA processing that occurs in the nucleus is Intron splicing. Nearly all of the work on Intron processing has been done in animal cells, but some data is emerging from plants. Intron processing depends on proper 5' and 3' splice junction sequences. Consensus sequences for these junctions have been derived for both animal and plant mRNAs, but only a few nucleotides are known to be invariant. Therefore, it is hard to predict with any certainty whether a putative splice junction is functional or partially functional based solely on sequence analysis. In particular, the only invariant nucleotides are GT at the 5' end of the intron and AG at the 3' end of the intron. In plants, at every nearby position, either within the intron or in the exon flanking the intron, all four nucleotides can be found, although some positions show some nucleotide preference (Brown, 1986; Hanley and Schuler, 1988).

[0011] A plant Intron has been moved from a patatin gene into a GUS gene. To do this, site directed mutagenesis was performed to introduce new restriction sites, and this mutagenesis changed several nucleotides in the Intron and exon sequences flanking the GT and AG. This Intron still functioned properly, indicating the importance of the GT and AG and the flexibility at other nucleotide positions. There are of course many occurrences of GT and AG in all genes that do not function as intron splice junctions, so there must be some other sequence or structural features that identify splice junctions. In plants, one such feature appears to be base composition per se. Weibauer et al. (1988) and Goodall et al. (1988) have analyzed plant Introns and exons and found that exons have ~50% A+T while Introns have ~70% A+T. Goodall et al. (1988) also created an artificial plant Intron that has consensus 5' and 3' splice junctions and a random A+T rich internal sequence. This Intron was spliced correctly in plants. When the internal segment was replaced by a G+C rich sequence, splicing efficiency was drastically reduced. These two examples demonstrate that Intron recognition in plants may depend on very general features - splice junctions that have a great deal of sequence diversity and A+T richness of the Intron itself. This, of course, makes it difficult to predict from sequence alone whether any particular sequence is likely to function as an active or partially active Intron for RNA processing.

[0012] *B.t.* genes being A+T rich contain numerous stretches of various lengths that have 70% or greater A+T. The number of such stretches identified by sequence analysis depends on the length of sequence scanned.

[0013] As for polyadenylation described above, there are complications in predicting what sequences might be utilized as splice sites in any given gene. First, many naturally occurring genes have alternative splicing pathways that create alternative combinations of exons in the final mRNA (Gallega and Nadal-Ginard, 1988; Helfman and Ricci, 1988; Tsurushita and Korn, 1989). That is, some splice junctions are apparently recognized under some circumstances or

In certain cell types, but not in others. The rules governing this are not understood. In addition, there can be an interaction between processing paths such that utilization of a particular polyadenylation site can interfere with splicing at a nearby splice site and vice versa (Adam and Nevins, 1988; Brady and Wold, 1988; Marzluff and Pandey, 1988). Again no predictive rules are available. Also, sequence changes in a gene can drastically alter the utilization of particular splice junctions. For example, in a bovine growth hormone gene, small deletions in an exon a few hundred bases downstream of an intron cause the splicing efficiency of the intron to drop from greater than 95% to less than 2% (essentially nonfunctional). Other deletions however have essentially no effect (Hampson and Rottman, 1988). Finally, a variety of *in vitro* and *in vivo* experiments indicate that mutations that disrupt normal splicing lead to rapid degradation of the RNA in the nucleus. Splicing is a multistep process in the nucleus and mutations in normal splicing can lead to blockades in the process at a variety of steps. Any of these blockades can then lead to an abnormal and unstable RNA. Studies of mutants of normally processed (polyadenylation and splicing) genes are relevant to the study of heterologous genes such as *B.t.* *B.t.* genes might contain functional signals that lead to the production of aberrant nonfunctional mRNAs, and these mRNAs are likely to be unstable. But the *B.t.* genes are perhaps even more likely to contain signals that are analogous to mutant signals in a natural gene. As shown above these mutant signals are very likely to cause defects in the processing pathways whose consequence is to produce unstable mRNAs.

[0014] It is not known with any certainty what signals RNA transcription termination in plant or animal cells. Some studies on animal genes that indicate that stretches of sequence rich in T cause termination by calf thymus RNA polymerase II *in vitro*. These studies have shown that the 3' ends of *in vitro* terminated transcripts often lie within runs of T such as T5, T6 or T7. Other identified sites have not been composed solely of T, but have had one or more other nucleotides as well. Termination has been found to occur within the sequences TATTTTT, ATTCTC, TTCTT (Derrick et al., 1987; Reines et al., 1987). In the case of these latter two, the context in which the sequence is found has been C+T rich as well. It is not known if this is essential. Other studies have implicated stretches of A as potential transcriptional terminators. An interesting example from SV40 illustrates the uncertainty in defining terminators based on sequence alone. One potential terminator in SV40 was identified as being A rich and having a region of dyad symmetry (potential stem-loop) 5' to the A rich stretch. However, a second terminator identified experimentally downstream in the same gene was not A rich and included no potential secondary structure (Kessler et al., 1988). Of course, due to the A+T content of *B.t.* genes, they are rich in runs of A or T that could act as terminators. The importance of termination to stability of the mRNA is shown by the globin gene example described above. Absence of a normal polyA site leads to a failure in proper termination with a consequent decrease in mRNA.

[0015] There is also an effect on mRNA stability due to the translation of the mRNA. Premature translational termination in human triose phosphate isomerase leads to instability of the mRNA (Daar et al., 1988). Another example is the beta-thalassemic globin mRNA described above that is specifically unstable in bone marrow cells (Lim et al., 1988). The defect in this mutant gene is a single base pair deletion at codon 44 that leads to translational termination (a nonsense codon) at codon 60. Compared to properly translated normal globin mRNA, this mutant RNA is very unstable. These results indicate that an improperly translated mRNA is unstable. Other work in yeast indicates that proper but poor translation can have an effect on mRNA levels. A heterologous gene was modified to convert certain codons to more yeast preferred codons. An overall 10-fold increase in protein production was achieved, but there was also about a 3-fold increase in mRNA (Hoekema et al., 1987). This indicates that more efficient translation can lead to greater mRNA stability, and that the effect of codon usage can be at the RNA level as well as the translational level. It is not clear from codon usage studies which codons lead to poor translation, or how this is coupled to mRNA stability.

[0016] EP-A-0 359 472 discloses modifying *B.t.* sequences to render them more plant-like. The sequence is modified so that the codon usage in the sequence is approximately the same as the codon usage in a plant. In contrast, the claimed invention is related to a specific methodology for increasing the expression of the gene in a plant by removing the occurrence of particular DNA sequences.

[0017] Therefore, it is an object of the present invention to provide a method for preparing synthetic plant genes which express their respective proteins at relatively high levels when compared to wild-type genes. It is yet another object of the present invention to provide synthetic plant genes which express the crystal protein toxin of *Bacillus thuringiensis* at relatively high levels.

#### 50 BRIEF DESCRIPTION OF THE DRAWINGS

[0018]

Figure 1 illustrates the steps employed in modifying a wild-type gene to increase expression efficiency in plants. Figure 2 illustrates a comparison of the changes in the modified *B.t.k.* HD-1 sequence of Example 1 (lower line) versus the wild-type sequence of *B.t.k.* HD-1 which encodes the crystal protein toxin (upper line). Figure 3 illustrates a comparison of the changes in the synthetic *B.t.k.* HD-1 sequence of Example 2 (lower line) versus the wild-type sequence of *B.t.k.* HD-1 which encodes the crystal protein toxin (upper line).

Figure 4 illustrates a comparison of the changes in the synthetic *B.t.k* HD-73 sequence of Example 3 (lower line) versus the wild-type sequence of *B.t.k* HD-73 (upper line).  
 Figure 5 represents a plasmid map of intermediate plant transformation vector cassette pMON893.  
 Figure 6 represents a plasmid map of intermediate plant transformation vector cassette pMON900.  
 Figure 7 represents a map for the disarmed T-DNA of *A. tumefaciens* ACO.  
 Figure 8 illustrates a comparison of the changes in the synthetic truncated *B.t.k* HD-73 gene (Amino acids 29-615 with an N-terminal Met-Ala) of Example 3 (lower line) versus the wild-type sequence of *B.t.k* HD-73 (upper line).  
 Figure 9 illustrates a comparison of the changes in the synthetic/wild-type full length *B.t.k* HD-73 sequence of Example 3 (lower line) versus the wild-type full-length sequence of *B.t.k* HD-73 (upper line).  
 Figure 10 illustrates a comparison of the changes in the synthetic/modified full length *B.t.k* HD-73 sequence of Example 3 (lower line) versus the wild-type full-length sequence of *B.t.k* HD-73 (upper line).  
 Figure 11 illustrates a comparison of the changes in the fully synthetic full-length *B.t.k* HD-73 sequence of Example 3 (lower line) versus the wild-type full-length sequence of *B.t.k* HD-73 (upper line).  
 Figure 12 illustrates a comparison of the changes in the synthetic *B.t.t* sequence of Example 5 (lower line) versus the wild-type sequence of *B.t.t* which encodes the crystal protein toxin (upper line).  
 Figure 13 illustrates a comparison of the changes in the synthetic *B.t*. P2 sequence of Example 6 (lower line) versus the wild-type sequence of *B.t*. *entomocidus* which encodes the Btnt protein toxin (upper line).  
 Figure 14 illustrates a comparison of the changes in the synthetic *B.t*. *entomocidus* sequence of Example 7 (lower line) versus the wild-type sequence of *B.t*. *entomocidus* which encodes the Btnt protein toxin (upper line).  
 Figure 15 illustrates a plasmid map for plant expression cassette vector pMON744.  
 Figure 16 illustrates a comparison of the changes in the synthetic potato leaf roll virus (PLRV) coat protein sequence of Example 9 (lower line) versus the wild-type coat protein sequence of PLRV (upper line).

#### STATEMENT OF THE INVENTION

[0019] The present invention provides a method for modifying a wild-type structural gene sequence which encodes an insecticidal protein of *Bacillus thuringiensis* to enhance the expression of said protein in plants which comprises:

- identifying regions within said sequence with greater than four consecutive adenine or thymine nucleotides;
- modifying the regions of step (a) which have two or more polyadenylation signals within a ten base sequence to remove said signals while maintaining a gene sequence which encodes said protein; and
- modifying the 15-30 base regions surrounding the regions of step (a) to remove major plant polyadenylation signals, consecutive sequences containing more than one minor polyadenylation signal and consecutive sequences containing more than one ATTAA sequence while maintaining a gene sequence which encodes said protein.

[0020] The invention further provides a method for modifying a wild-type structural gene sequence which encodes an insecticidal protein of *Bacillus thuringiensis* to enhance the expression of said protein in plants which comprises:

- removing polyadenylation signals contained in said wild-type gene while retaining a sequence which encodes said protein; and
- removing ATTAA sequences contained in said wild-type gene while retaining a sequence which encodes said protein.

[0021] According to a further embodiment a method for improving the expression of a heterologous gene in plants is provided, wherein said gene comprises a modified chimeric gene containing a promoter which functions in plant cells operably linked to a structural coding sequence and a 3' non-translated region containing a polyadenylation signal which functions in plants to cause the addition of polyadenylate nucleotides to the 3' end of the RNA, and wherein said structural coding sequence encodes an insecticidal protein, at least a portion of which was derived from a *Bacillus thuringiensis* protein, wherein said method comprises modifying said structural coding sequence so that said sequence has a DNA sequence which differs from the naturally occurring DNA sequence encoding said *Bacillus thuringiensis* protein and said structural coding sequence does not contain more than 5 consecutive nucleotides consisting of either adenine or thymine residues.

[0022] As a further embodiment, a method for improving the expression of a heterologous gene in plants is provided, wherein said gene comprises a modified chimeric gene containing a promoter which functions in plant cells operably linked to a structural coding sequence and a 3' non-translated region containing a polyadenylation signal which functions in plants to cause the addition of polyadenylate nucleotides to the 3' end of the RNA, wherein said structural

coding sequence encodes an insecticidal protein at least a portion of which was derived from a *Bacillus thuringiensis* protein, wherein said method comprises modifying said structural coding sequence so that said sequence has a DNA sequence which differs from the naturally occurring DNA sequence encoding said *Bacillus thuringiensis* protein and has the following characteristics:

5 said structural coding sequence has a region which is complementary to the following sequence:

10 
$$\begin{matrix} \text{GGCTTGATTCTAGCGA} & \text{ACTCTTCGATTC} & \text{CTCTGGTTGATGAGCTGTT} \\ 1 & 5 & 10 & 15 & 20 & 25 & 30 & 35 & 40 & 45 \end{matrix}$$

said region in said coding sequence having eliminated 2 ACCAA and 1 ATTAA sequence:

15 [0023] The present invention provides a method for preparing synthetic plant genes which encode the crystal protein toxin of *Bacillus thuringiensis* (*B.t.*). Suitable *B.t.* subspecies include, but are not limited to, *B.t. kurstaki* HD-73, *B.t. satoi*, *B.t. berliner*, *B.t. thuringiensis*, *B.t. tolworthi*, *B.t. dendrolimus*, *B.t. alesti*, *B.t. galleriae*, *B.t. aizawai*, *B.t. subtilis*, *B.t. entomocidus*, *B.t. tenebrionis* and *B.t. san diego*.

20 [0024] The expression of *B.t.* genes in plants is problematic. Although the expression of *B.t.* genes in plants at insecticidal levels has been reported, this accomplishment has not been straightforward. In particular, the expression of a full-length lepidopteran specific *B.t.* gene (comprising DNA from a *B.t.k.* isolate) has been reported to be unsuccessful in yielding insecticidal levels of expression in some plant species (Vaeck et al., 1987 and Barton et al., 1987).

25 [0025] It has been reported that expression of the full-length gene from *B.t.k.* HD-1 was detectable in tomato plants but that truncated genes led to a higher frequency of insecticidal plants with an overall higher level of expression. Truncated genes of *B.t. berlinei* also led to a higher frequency of insecticidal plants in tobacco (Vaeck et al., 1987). On the other hand, insecticidal plants were provided from lettuce transformants using a full-length gene.

30 [0026] It has also been reported that the full length gene from *B.t.k.* HD-73 gave some insecticidal effect in tobacco (Adang et al., 1987). However, the *B.t.* mRNA detected in these plants was only 1.7 kb compared to the expected 3.7 kb indicating improper expression of the gene. It was suggested that this truncated mRNA was too short to encode a functional truncated toxin, but there must have been a low level of longer mRNA in some plants or no insecticidal activity would have been observed. Others have reported in a publication that they observed a large amount of shorter than expected mRNA from a truncated *B.t.k.* gene, but some mRNA of the expected size was also observed. In fact, it was suggested that expression of the full length gene is toxic to tobacco callus (Barton et al., 1987). The above illustrates that lepidopteran type *B.t.* genes are poorly expressed in plants compared to other chimeric genes previously expressed from the same promoter cassettes.

35 [0027] The expression of *B.t.* in tomato and potato is at levels similar to that of *B.t.k.* (i.e., poor). *B.t.t.* and *B.t.k.* genes share only limited sequence homology, but they share many common features in terms of base composition and the presence of particular A-T rich elements.

40 [0028] All reports in the field have noted the lower than expected expression of *B.t.* genes in plants. In general, insecticidal efficacy has been measured using insects very sensitive to *B.t.* toxin such as tobacco hornworm. Although it has been possible to obtain plants totally protected against tobacco hornworm, it is important to note that hornworm is up to 500 fold more sensitive to *B.t.* toxin than some agriculturally important insect pests such as beet armyworm. It is therefore of interest to obtain transgenic plants that are protected against all important lepidopteran pests (or against Colorado potato beetle in the case of *B.t. tenebrionis*), and in addition to have a level of *B.t.* expression that provides an additional safety margin over and above the efficacious protection level. It is also important to devise plant genes which function reproducibly from species to species, so that insect resistant plants can be obtained in a predictable fashion.

45 [0029] In order to achieve these goals, it is important to understand the nature of the poorer than expected expression of *B.t.* genes in plants. The level of stable *B.t.* mRNA in plants is much lower than expected. That is, compared to other coding sequences driven by the same promoter, the level of *B.t.* mRNA measured by Northern analysis or nucleic acid protection experiments is much lower. For example, tomato plant 337 (Fischhoff et al., 1987) was selected as the best expressing plant with pMON9711 which contains the *B.t.k.* HD-1 KpnI fragment driven by the CaMV 35S promoter and contains the NOS-NPTII-NOS selectable marker gene. In this plant the level of *B.t.* mRNA is between 100 to 1000 fold lower than the level of NPTII mRNA, even though the 35S promoter is approximately 50-fold stronger than the NOS promoter (Sanders et al., 1987).

50 [0030] The level of *B.t.* toxin protein detected in plants is consistent with the low level of *B.t.* mRNA. Moreover, the insecticidal efficacy of the transgenic plants correlates with the *B.t.* protein level indicating that the toxin protein produced in plants is biologically active. Therefore, the low level of *B.t.* toxin expression may be the result of the low levels

of *B.t.* mRNA.

[0031] Messenger RNA levels are determined by the rate of synthesis and rate of degradation. It is the balance between these two that determines the steady state level of mRNA. The rate of synthesis has been maximized by the use of the CaMV 35S promoter, a strong constitutive plant expressible promoter. The use of other plant promoters such as nopaline synthase (NOS), mannopine synthase (MAS) and ribulose bisphosphatecarboxylase small subunit (RUBISCO) have not led to dramatic changes in the levels of *B.t.* toxin protein expression indicating that the effects determining *B.t.* toxin protein levels are promoter independent. These data imply that the coding sequences of DNA genes encoding *B.t.* toxin proteins are somehow responsible for the poor expression level, and that this effect is manifested by a low level of accumulated stable mRNA.

[0032] Lower than expected levels of mRNA have been observed with four different lepidopteran specific genes (two from *B.t.k.* HD-1; *B.t. berliner* and *B.t.k.* HD-73) as well as the gene from the coleopteran specific *B.t. tenebrionis*. It appears that for lepidopteran type *B.t.* genes these effects are manifest more strongly in the full length coding sequences than in the truncated coding sequences. These effects are seen across plant species although their magnitude seems greater in some plant species such as tobacco.

[0033] The nature of the coding sequences of *B.t.* genes distinguishes them from plant genes as well as many other heterologous genes expressed in plants. In particular, *B.t.* genes are very rich (~62%) in adenine (A) and thymine (T) while plant genes and most bacterial genes which have been expressed in plants are on the order of 45-55% A+T. The A+T content of the genomes (and thus the genes) of any organism are features of that organism and reflect its evolutionary history. While within any one organism genes have similar A+T content, the A+T content can vary tremendously from organism to organism. For example, some *Bacillus* species have among the most A+T rich genomes while some *Streptomyces* species are among the least A+T rich genomes (~30 to 35% A+T).

[0034] Due to the degeneracy of the genetic code and the limited number of codon choices for any amino acid, most of the "excess" A+T of the structural coding sequences of some *Bacillus* species are found in the third position of the codons. That is, genes of some *Bacillus* species have A or T as the third nucleotide in many codons. Thus A+T content

25 In part can determine codon usage bias. In addition, it is clear that genes evolve for maximum function in the organism in which they evolve. This means that particular nucleotide sequences found in a gene from one organism, where they may play no role except to code for a particular stretch of amino acids, have the potential to be recognized as gene control elements in another organism (such as transcriptional promoters or terminators, polyA addition sites, Intron splice sites, or specific mRNA degradation signals). It is perhaps surprising that such misread signals are not a more common feature of heterologous gene expression, but this can be explained in part by the relatively homogeneous A+T content (~50%) of many organisms. This A+T content plus the nature of the genetic code put clear constraints on the likelihood of occurrence of any particular oligonucleotide sequence. Thus, a gene from *E. coli* with a 50% A+T content is much less likely to contain any particular A+T rich segment than a gene from *B. thuringiensis*.

[0035] As described above, the expression of *B.t.* toxin protein in plants has been problematic. Although the observations made in other systems described above offer the hope of a means to elevate the expression level of *B.t.* toxin proteins in plants, the success obtained by the present method is quite unexpected. Indeed, inasmuch as it has been recently reported that expression of the full-length *B.t.k.* toxin protein in tobacco makes callus tissue necrotic (Barton et al., 1987); one would reasonably expect that high level expression of *B.t.* toxin protein to be unattainable due to the reported toxicity effects.

[0036] In its most rigorous application, the method of the present invention involves the modification of an existing structural coding sequence ("structural gene") which codes for a particular protein by removal of ATTAA sequences and putative polyadenylation signals by site directed mutagenesis of the DNA comprising the structural gene. It is most preferred that substantially all the polyadenylation signals and ATTAA sequences are removed although enhanced expression levels are observed with only partial removal of either of the above identified sequences. Alternately if a synthetic gene is prepared which codes for the expression of the subject protein, codons are selected to avoid the ATTAA sequence and putative polyadenylation signals. For purposes of the present invention putative polyadenylation signals include, but are not necessarily limited to, AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA and CATAAA. In replacing the ATTAA sequences and polyadenylation signals, codons are preferably utilized which avoid the codons which are rarely found in plant genomes.

[0037] Another embodiment of the present invention, represented in the flow diagram of Figure 1, employs a method for the modification of an existing structural gene or alternately the *de novo* synthesis of a structural gene which method is somewhat less rigorous than the method first described above. Referring to Figure 1, the selected DNA sequence is scanned to identify regions with greater than four consecutive adenine (A) or thymine (T) nucleotides. The A+T regions are scanned for potential plant polyadenylation signals. Although the absence of five or more consecutive A or T nucleotides eliminates most plant polyadenylation signals, if there are more than one of the minor polyadenylation signals identified within ten nucleotides of each other, then the nucleotide sequence of this region is preferably altered to remove these signals while maintaining the original encoded amino acid sequence.

[0038] The second step is to consider the 15 to 30 nucleotide regions surrounding the A+T rich region identified in step one. If the A+T content of the surrounding region is less than 80%, the region should be examined for polyadenylation signals. Alteration of the region based on polyadenylation signals is dependent upon (1) the number of polyadenylation signals present and (2) presence of a major plant polyadenylation signal.

[0039] The extended region is examined for the presence of plant polyadenylation signals. The polyadenylation signals are removed by site-directed mutagenesis of the DNA sequence. The extended region is also examined for multiple copies of the ATTTA sequence which are also removed by mutagenesis.

[0040] It is also preferred that regions comprising many consecutive A+T bases or G+C bases are disrupted since these regions are predicted to have a higher likelihood to form hairpin structure due to self-complementarity. Therefore, insertion of heterogeneous base pairs would reduce the likelihood of self-complementary secondary structure formation which are known to inhibit transcription and/or translation in some organisms. In most cases, the adverse effects may be minimized by using sequences which do not contain more than five consecutive A+T or G+C.

#### SYNTHETIC OLIGONUCLEOTIDES FOR MUTAGENESIS

[0041] The oligonucleotides used in the mutagenesis are designed to maintain the proper amino acid sequence and reading frame and preferably to not introduce common restriction sites such as BgIII, HindIII, SacI, KpnI, EcoRI, NcoI, PstI and SalI into the modified gene. These restriction sites are found in multilinker insertion sites of cloning vectors such as plasmids pUC118 and pMON7258. Of course, the introduction of new polyadenylation signals, ATTTA sequences or consecutive stretches of more than five A+T or G+C, should also be avoided. The preferred size for the oligonucleotides is around 40-50 bases, but fragments ranging from 18 to 100 bases have been utilized. In most cases, a minimum of 5 to 8 base pairs of homology to the template DNA on both ends of the synthesized fragment are maintained to insure proper hybridization of the primer to the template. The oligonucleotides should avoid sequences longer than five base pairs A+T or G+C. Codons used in the replacement of wild-type codons should preferably avoid the TA or CG doublet wherever possible. Codons are selected from a plant preferred codon table (such as Table I below) so as to avoid codons which are rarely found in plant genomes, and efforts should be made to select codons to preferably adjust the G+C content to about 50%.

Table I

Preferred Codon Usage in Plants		
Amino Acid	Codon	Percent Usage in Plants
ARG	CGA	7
	CGC	11
	CGG	5
	CGU	25
	AGA	29
	AGG	23
LEU	CUA	8
	CUC	20
	CUG	10
	CUU	28
	UUA	5
	UUG	30
SER	UCA	14
	UCC	26
	UCG	3
	UCU	21
	AGC	21
	AGU	15
THR	ACA	21
	ACC	41

Table I (continued)

Preferred Codon Usage In Plants		
Amino Acid	Codon	Percent Usage in Plants
5 PRO	ACG	7
	ACU	31
	CCA	45
	CCC	19
	CCG	9
	CCU	26
10 ALA	GCA	23
	GCC	32
	GCG	3
	GCU	41
15 GLY	GGA	32
	GGC	20
	GGG	11
	GGU	37
20 ILE	AUA	12
	AUC	45
	AUU	43
25 VAL	GUA	9
	GUC	20
	GUG	28
	GUU	43
30 LYS	AAA	36
	AAG	64
35 ASN	AAC	72
	AAU	28
40 GLN	CAA	64
	CAG	36
45 HIS	CAC	65
	CAU	35
50 GLU	GAA	48
	GAG	52
55 ASP	GAC	48
	GAU	52
TYR	UAC	68
	UAU	32
CYS	UGC	78

Table I (continued)

Preferred Codon Usage In Plants		
Amino Acid	Codon	Percent Usage In Plants
PHE	UGU	22
	UUC	56
	UUU	44
MET	AUG	100
	UGG	100

[0042] Regions with many consecutive A+T bases or G+C bases are predicted to have a higher likelihood to form hairpin structures due to self-complementarity. Disruption of these regions by the insertion of heterogeneous base pairs is preferred and should reduce the likelihood of the formation of self-complementary secondary structures such as hairpins which are known in some organisms to inhibit transcription (transcriptional terminators) and translation (attenuators). However, it is difficult to predict the biological effect of a potential hairpin forming region.

[0043] It is evident to those skilled in the art that while the above description is directed toward the modification of the DNA sequences of wild-type genes, the present method can be used to construct a completely synthetic gene for a given amino acid sequence. Regions with five or more consecutive A+T or G+C nucleotides should be avoided. Codons should be selected avoiding the TA and CG doublets in codons whenever possible. Codon usage can be normalized against a plant preferred codon usage table (such as Table I) and the G+C content preferably adjusted to about 50%. The resulting sequence should be examined to ensure that there are minimal putative plant polyadenylation signals and ATTAA sequences. Restriction sites found in commonly used cloning vectors are also preferably avoided. However, placement of several unique restriction sites throughout the gene is useful for analysis of gene expression or construction of gene variants.

#### Plant Gene Construction

[0044] The expression of a plant gene which exists in double-stranded DNA form involves transcription of messenger RNA (mRNA) from one strand of the DNA by RNA polymerase enzyme, and the subsequent processing of the mRNA primary transcript inside the nucleus. This processing involves a 3' non-translated region which adds polyadenylate nucleotides to the 3' end of the RNA. Transcription of DNA into mRNA is regulated by a region of DNA usually referred to as the "promoter." The promoter region contains a sequence of bases that signals RNA polymerase to associate with the DNA and to initiate the transcription of mRNA using one of the DNA strands as a template to make a corresponding strand of RNA.

[0045] A number of promoters which are active in plant cells have been described in the literature. These include the napinase synthase (NOS) and octopine synthase (OCS) promoters (which are carried on tumor-inducing plasmids of *Agrobacterium tumefaciens*), the Cauliflower Mosaic Virus (CaMV) 19S and 35S promoters, the light-inducible promoter from the small subunit of ribulose bis-phosphate carboxylase (ssRUBISCO, a very abundant plant polypeptide) and the mannopine synthase (MAS) promoter (Velten et al. 1984 and Velten & Schell, 1985). All of these promoters have been used to create various types of DNA constructs which have been expressed in plants (see e.g., PCT publication WO84/02913 (Rogers et al., Monsanto).

[0046] Promoters which are known or are found to cause transcription of RNA in plant cells can be used in the present invention. Such promoters may be obtained from plants or plant viruses and include, but are not limited to, the CaMV35S promoter and promoters isolated from plant genes such as ssRUBISCO genes. As described below, it is preferred that the particular promoter selected should be capable of causing sufficient expression to result in the production of an effective amount of protein.

[0047] The promoters used in the DNA constructs (i.e. chimeric plant genes) of the present invention may be modified, if desired, to affect their control characteristics. For example, the CaMV35S promoter may be ligated to the portion of the ssRUBISCO gene that represses the expression of ssRUBISCO in the absence of light, to create a promoter which is active in leaves but not in roots. The resulting chimeric promoter may be used as described herein. For purposes of this description, the phrase "CaMV35S" promoter thus includes variations of CaMV35S promoter, e.g., promoters derived by means of ligation with operator regions, random or controlled mutagenesis, etc. Furthermore, the promoters may be altered to contain multiple "enhancer sequences" to assist in elevating gene expression.

[0048] The RNA produced by a DNA construct of the present invention also contains a 5' non-translated leader

sequence. This sequence can be derived from the promoter selected to express the gene, and can be specifically modified so as to increase translation of the mRNA. The 5' non-translated regions can also be obtained from viral RNA's, from suitable eukaryotic genes, or from a synthetic gene sequence. The present Invention is not limited to constructs as presented in the following examples. Rather, the non-translated leader sequence can be part of the 5' end of the non-translated region of the coding sequence for the virus coat protein, or part of the promoter sequence, or can be derived from an unrelated promoter or coding sequence. In any case, it is preferred that the sequence flanking the initiation site conform to the translational consensus sequence rules for enhanced translation initiation reported by Kozak (1984).

[0049] The DNA construct of the present invention also contains a modified or fully-synthetic structural coding sequence encoding the crystal toxin protein of *Bacillus thuringiensis* which has been changed to enhance the performance of the gene in plants. The structural genes of the present invention may optionally encode a fusion protein comprising an amino-terminal chloroplast transit peptide or secretory signal sequence (see for instance, Examples 10 and 11).

[0050] The DNA construct also contains a 3' non-translated region. The 3' non-translated region contains a polyadenylation signal which functions in plants to cause the addition of polyadenylated nucleotides to the 3' end of the viral RNA. Examples of suitable 3' regions are (1) the 3' transcribed, non-translated regions containing the polyadenylation signal of *Agrobacterium* tumor-inducing (Ti) plasmid genes, such as the nopaline synthase (NOS) gene, and (2) plant genes like the soybean storage protein (7S) genes and the small subunit of the RuBP carboxylase (E9) gene. An example of a preferred 3' region is that from the 7S gene, described in greater detail in the examples below.

## 20 Plant Transformation

[0051] A chimeric plant gene containing a structural coding sequence of the present invention can be inserted into the genome of a plant by any suitable method. Suitable plants for use in the practice of the present invention include, but are not limited to, soybean, cotton, alfalfa, oilseed rape, flax, tomato, sugarbeet, sunflower, potato, tobacco, maize, 25 rice and wheat. Suitable plant transformation vectors include those derived from a Ti plasmid of *Agrobacterium tumefaciens*, as well as those disclosed, e.g., by Herrera-Estrella (1983), Bevan (1983), Klee (1985) and EPO publication 120,516 (Schilperoort et al.). In addition to plant transformation vectors derived from the Ti or root-inducing (Ri) plasmids of *Agrobacterium*, alternative methods can be used to insert the DNA constructs of this invention into plant cells. Such methods may involve, for example, the use of liposomes, electroporation, chemicals that increase free DNA uptake, 30 free DNA delivery via microprojectile bombardment, and transformation using viruses or pollen.

[0052] A particularly useful Ti plasmid cassette vector for transformation of dicotyledonous plants is shown in Figure 5. Referring to Figure 5, the expression cassette pMON893 consists of the enhanced CaMV35S promoter (EN 35S) and the 3' end including polyadenylation signals from a soybean gene encoding the alpha-prime subunit of beta-conglycinin. Between these two elements is a multilinker containing multiple restriction sites for the insertion of genes.

[0053] The enhanced CaMV35S promoter was constructed as follows. A fragment of the CaMV35S promoter extending between position -343 and +9 was previously constructed in pUC13 by Odell et al. (1985). This segment contains a region identified by Odell et al. (1985) as being necessary for maximal expression of the CaMV35S promoter. It was excised as a ClaI-HindIII fragment, made blunt ended with DNA polymerase I (Klenow fragment) and inserted into the HindIII site of pUC18. This upstream region of the 35S promoter was excised from this plasmid as a HindIII-EcoRV fragment (extending from -343 to -90) and inserted into the same plasmid between the HindIII and PstI sites. The enhanced CaMV35S promoter thus contains a duplication of sequences between -343 and -90 (Kay et al., 1987).

[0054] The 3' end of the 7S gene is derived from the 7S gene contained on the clone designated 17.1 (Schuler et al., 1982). This 3' end fragment, which includes the polyadenylation signals, extends from an Avall site located about 30 bp upstream of the termination codon for the beta-conglycinin gene in clone 17.1 to an EcoRI site located about 450 bp downstream of this termination codon.

[0055] The remainder of pMON893 contains a segment of pBFR322 which provides an origin of replication in *E. coli* and a region for homologous recombination with the disarmed T-DNA in *Agrobacterium* strain ACO (described below); the oriV region from the broad host range plasmid RK1; the streptomycin/spectinomycin resistance gene from Tn7; and a chimeric NPTII gene, containing the CaMV35S promoter and the nopaline synthase (NOS) 3' end, which provides 50 kanamycin resistance in transformed plant cells.

[0056] Referring to Figure 6, transformation vector plasmid pMON900 is a derivative of pMON893. The enhanced CaMV35S promoter of pMON893 has been replaced with the 1.5kb mannopine synthase (MAS) promoter (Velten et al. 1984). The other segments are the same as plasmid pMON893. After incorporation of a DNA construct into plasmid vector pMON893 or pMON900, the intermediate vector is introduced into *A. tumefaciens* strain ACO which contains 55 a disarmed Ti plasmid. Cointegrate Ti plasmid vectors are selected and used to transform dicotyledonous plants.

[0057] Referring to Figure 7, *A. tumefaciens* ACO is a disarmed strain similar to pTiB6SE described by Fraley et al. (1985). For construction of ACO the starting *Agrobacterium* strain was the strain A208 which contains a nopaline-type Ti plasmid. The Ti plasmid was disarmed in a manner similar to that described by Fraley et al. (1985) so that essentially

all of the native T-DNA was removed except for the left border and a few hundred base pairs of T-DNA inside the left border. The remainder of the T-DNA extending to a point just beyond the right border was replaced with a novel piece of DNA including (from left to right) a segment of pBR322, the oriV region from plasmid RK2, and the kanamycin resistance gene from Tn601. The pBR322 and oriV segments are similar to the segments in pMON893 and provide a region of homology for co-integrate formation.

[0058] The following examples are provided to better elucidate the practice of the present invention and should not be interpreted in any way to limit the scope of the present invention. Those skilled in the art will recognize that various modifications, truncations etc. can be made to the methods and genes described herein while not departing from the spirit and scope of the present invention.

**Example 1 -- Modified *B.t.k* HD-1 Gene**

[0059] Referring to Figure 2, the wild-type *B.t.k* HD-1 gene is known to be expressed poorly in plants as a full length gene or as a truncated gene. The G+C content of the *B.t.k* gene is low (37%) containing many A+T rich regions, potential polyadenylation sites (18 sites; see Table II for the list of sequences) and numerous ATTTA sequences.

Table II

List of Sequences of the Potential  
Polyadenylation Signals

AATAAA*	AAGCAT
AATAAT*	ATTAAT
AACCAA	ATACAT
ATATAA	AAAATA
AATCAA	ATTTAA**
ATACTA	AATTAA**
ATAAAA	AATACA**
ATGAAA	CATAAA**

\* indicates a potential major plant polyadenylation site.

\*\* indicates a potential minor animal polyadenylation site.

All others are potential minor plant polyadenylation sites.

[0060] Table III lists the synthetic oligonucleotides designed and synthesized for the site-directed mutagenesis of the *B.t.k* HD-1 gene.

Table III

Mutagenesis Primers for B.t.k. HD-1 Gene

Primer	Length (bp)	Sequence
BTK185	18	TCCCCAGATA ATATCAAC
BTK240	48	GGCTTGATTC CTAGCGAACT CTTCGATTCT CTGGTTGATG AGCTGTTC
BTK462	54	CAAAACTGAG AGGTGGAGGT TGGCAGCTTG AACGTACACG GAGAGGGAGAGGAAC
BTK669	48	AGTTAGTGTA AGCTCTCTTC TGAACCTGGTT GTACCTGATC CAATCTCT
BTK930	39	AGCCATGATC TGGTGAACCGG ACCACTAGTA TTCTCCCTCT
BTK1110	32	AGTTGTTGGT TGTTGATCCC GATGTTAAAA GG

Table III - continued

Mutagenesis Primers for B.t.k. HD-1 Gene

Primer	Length (bp)	Sequence
BTK1380A	37	GTGATGAAGG GATGATGTTG TTGAACTCAG CACTACG
BTK1380T	100	CAGAAGTTCC AGAGCCAAGA TTAGTAGACT TGGTGAGTGG GATTTGGGTG ATTTGTGATG AAGGGATGAT GTTGTGAAAC TCAGCACTAC GATGTATCCA
BTK1600	27	TGATGTGTGG AACTGAAGGT TTGTGGT

[0061] The *B.t.k* HD-1 gene (BglII fragment from pMON9921 encoding amino acids 29-607 with a Met-Ala at the N-terminus) was cloned into pMON7258 (pUC18 derivative which contains a BglII site in the multicloning region) at the BglII site resulting in pMON5342. The orientation of the *B.t.k.* gene was chosen so that the opposite strand (negative strand) was synthesized in filamentous phage particles for the mutagenesis. The procedure of Kunkle (1985) was used for the mutagenesis using plasmid pMON5342 as starting material.

[0062] The regions for mutagenesis were selected in the following manner. All regions of the DNA sequence of the *B.t.k.* gene were identified which contained five or more consecutive base pairs which were A or T. These were ranked in terms of length and highest percentage of A+T in the surrounding sequence over a 20-30 base pair region. The DNA was then analysed for regions which might contain polyadenylation sites (see Table II above) or ATTAA sequences. Oligonucleotides were designed which maximized the elimination of A+T consecutive regions which contained one or more polyadenylation sites or ATTAA sequences. Two potential plant polyadenylation sites were rated more critical (see Table II) based on published reports. Codons were selected which increased G+C content, did not generate restriction sites for enzymes useful for cloning and assembly of the modified gene (BamHI, BglII, SacI, NcoI, EcoRV) and did not contain the doublets TA or GC which have been reported to be infrequently found in codons in plants. The oligonucleotides were at least 18 bp long ranging up to 100 base pairs and contained at least 5-8 base pairs of direct homology to native sequences at the ends of the fragments for efficient hybridization and priming in site-directed mutagenesis reactions. Figure 2 compares the wild-type *B.t.k.* HD-1 gene sequence with the sequence which resulted from the modifications by site-directed mutagenesis.

[0063] The end result of these changes was to increase the G+C content of *B.t.k.* gene from 37% to 41% while also decreasing the potential plant polyadenylation sites from 18 to 7 and decreasing the ATTAA regions from 13 to 7. Specifically, the mutagenesis changes from amino (5') terminus to the carboxy (3') terminus are as follows:

[0064] BTK185 is an 18-mer used to eliminate a plant polyadenylation site in the midst of a nine base pair region of A+T.

[0065] BTK240 is a 48-mer. Seven base pairs were changed by this oligonucleotide to eliminate three potential polyadenylation sites (2 AACCAA, 1 ATTAA). Another region close to the region altered by BTK240, starting at bp 312, had a high A+T content (13 of 15 base pairs) and an ATTAA region. However, it did not contain a potential polyadenylation site and its longest string of uninterrupted A+T was seven base pairs.

[0066] BTK462 is a 54-mer introducing 13 base pair changes. The first six changes were to reduce the A+T richness of the gene by replacing wild-type codons with codons containing G and C while avoiding the CG doublet. The next

seven changes made by BTK462 were used to eliminate an A+T rich region (13 of 14 base pairs were A or T) containing two ATTTA regions.

[0067] BTK669 is a 48-mer making nine individual base pair changes eliminating three possible polyadenylation sites (ATATAA, AATCAA, and AATTAA) and a single ATTTA site.

[0068] BTK933 is a 39-mer designed to increase the G+C content and to eliminate a potential polyadenylation site (AATAAT - a major site). This region did contain a nine base pair region of consecutive A+T sequence. One of the base pair changes was a G to A because a G at this position would have created a G+C rich region (CCGG(G)C). Since sequencing reactions indicate that there can be difficulties generating sequence through G+C consecutive bases, it was thought to be prudent to avoid generating potentially problematic regions even if they were problematic only *in vitro*.  
 [0069] BTK1110 is a 32-mer designed to introduce five changes in the wild-type gene. One potential site (AATAAT - a major site) was eliminated in the midst of an A+T rich region (19 of 22 base pairs).

[0070] BTK1380A and BTK1380T are responsible for 14 individual base pair changes. The first region (1380A) has 17 consecutive A+T base pairs. In this region is an ATTTA and a potential polyadenylation site (AATAAT). The 100-mer (1380T) contains all the changes dictated by 1380A. The large size of this primer was in part an experiment to determine if it was feasible to utilize large oligonucleotides for mutagenesis (over 60 bases in length). A second consideration was that the 100-mer was used to mutagenize a template which had previously been mutagenized by 1380A. The original primer ordered to mutagenize the region downstream and adjacent to 1380A did not anneal efficiently to the desired site as indicated by an inability to obtain clean sequence utilizing the primer. The large region of homology of 1380T did assure proper annealing. The extended size of 1380T was more of a convenience rather than a necessity.

[0071] BTK1600 is a 27-mer responsible for five individual base pair changes. An ATTTA region and a plant polyadenylation site were identified and the appropriate changes engineered.

[0072] A total of 62 bases were changed by site-directed mutagenesis. The G+C content increased by 55 base pairs, the potential polyadenylation sites were reduced from 18 to seven and the ATTTA sequences decreased from 13 to seven. The changes in the DNA sequence resulted in changes in 55 of the 579 codons in the truncated *B.t.k* gene in pMON5342 (approximately 9.5%).

[0073] Referring to Table IV modified *B.t.k* HD-1 genes were constructed that contained all of the above modifications (pMON5370) or various subsets of individual modifications. These genes were inserted into pMON893 for plant transformation and tobacco plants containing these genes were analyzed. The analysis of tobacco plants with the individual modifications was undertaken for several reasons. Expression of the wild type truncated gene in tobacco is very poor, resulting in infrequent identification of plants toxic to THW. Toxicity is defined by leaf feeding assays as at least 60% mortality of tobacco hornworm neonate larvae with a damage rating of 1 or less (scale is 0 to 4; 0 is equivalent to total protection, 4 total damage). The modified HD-1 gene (pMON5370) shows a large increase in expression (estimated to be approximately 100-fold; see Table VIII) in tobacco. Therefore, increases in expression of the wild-type gene due to individual modifications would be apparently a large increase in the frequency of toxic tobacco plants and the presence of detectable *B.t.k* protein. Results are shown in the following table:

Table IV.

Relative effects of Regional Modifications within the <i>B.t.k</i> Gene				
Construct	Position Modified	# of Plants	# of Toxic Plants	
pMON5370	185, 240, 669, 930, 1110, 1380a+b, 1600	38	22	
pMON10707	185, 240, 462, 669	48	19	
pMON10706	930, 1110, 1380a+b, 1600	43	1	
pMON10539	185	55	2	
pMON10537	240	57	17	
pMON10540	185, 240	88	23	
pMON10705	462	47	1	

[0074] The effects of each individual oligonucleotides' changes on expression did reveal some overall trends. Six

different constructs were generated which were designed to identify the key regions. The nine different oligonucleotides were divided in half by their position on the gene. Changes in the N-terminal half were incorporated into pMON10707 (185,240,462,669). C-terminal half changes were incorporated into pMON10706 (930,1110,1380a+b,1600). The results of analysis of plants with these two constructs indicate that pMON10707 produces a substantial number of toxic plants (19 of 48). Protein from these plants is detectable by ELISA analysis. pMON10706 plants were rarely identified as insecticidal (1 of 43) and the levels of *B.t.k* were barely detectable by immunological analysis. Investigation of the N-terminal changes in greater detail was done with 4 pMON constructs; 10539 (185 alone), 10537 (240 alone), 10540 (185 and 240) and 10705 (462 alone). The results indicate that the presence of the changes in 240 were required to generate a substantial number of toxic plants (pMON10540; 23 of 88, pMON10537; 17 of 57). The absence of the 240 changes resulted in a low frequency of toxic plants with low *B.t.k* protein levels, identical to results with the wild type gene. These results indicate that the changes in 240 are responsible for a substantial increase in *B.t.k* expression levels over an analogous wild-type construct in tobacco. Changes in additional regions (185,462,669) in conjunction with 240 may result in increases in *B.t.k* expression (>2 fold). However, changes at the 240 region of the N-terminal portion of the gene do result in dramatic increases in expression.

[0075] Despite the importance of the alteration of the 240 region in expression of modified genes, increased expression can be achieved by alteration of other regions. Hybrid genes, part wild-type, part synthetic, were generated to determine the effects of synthetic gene segments on the levels of *B.t.k* expression. A hybrid gene was generated with a synthetic N-terminal third (base pair 1 to 590 of Figure 2; to the XbaI site) with the C-terminal wild type *B.t.k* HD-1 (pMON5378). Plants transformed with this vector were as toxic as plants transformed with the modified HD-1 gene (pMON5370). This is consistent with the alteration of the 240 region. However, pMON10538, a hybrid with a wild-type N-terminal third (wild type gene for the first 600 base pairs, to the second XbaI site) and a synthetic C-terminal last two-thirds (base pair 590 to 1845 of Figure 3 was used to transform tobacco and resulted in a dramatic increase in expression. The levels of expression do not appear to be as high as those seen with the synthetic gene, but are comparable to the modified gene levels. These results indicate that modification of the 240 segment is not essential to increased expression since pMON10538 has an intact 240 region. A fully synthetic gene is, in most cases, superior for expression levels of *B.t.k*. (See Example 2.)

#### Example 2 -- Fully Synthetic *B.t.k* HD-1 Gene

[0076] A synthetic *B.t.k* HD-1 gene was designed using the preferred plant codons listed in Table V below. Table V lists the codons and frequency of use in plant genes of dicotyledonous plants compared to the frequency of their use in the wild type *B.t.k* HD-1 gene (amino acids 1-615) and the synthetic gene of this example. The total number of each amino acid in this segment of the gene is listed in the parenthesis under the amino acid designated.

Table V

Codon In Usage Synthetic <i>B.t.k</i> HD-1 Gene				
Amino Acid	Codon	Percent Usage In Plants	Wt <i>B.t.k</i> /Syn	
ARG (43)	CGA	7	11	2
	CGC	11	5	5
	CGG	5	2	0
	CGU	25	14	27
	AGA	29	55	41
	AGG	23	14	25
LE (49)	CUA	8	16	4
	CUC	20	0	20
	CUG	10	2	6
	CUU	28	22	24
	UUA	5	50	0
	UUG	30	10	45

Table V (continued)

Codon in Usage Synthetic B.t.k. HD-1 Gene					
	Amino Acid	Codon	Percent Usage in Plants/Wt B.t.k./Syn		
5	SER (64)	UCA	14	27	
		UCC	26	9	
		UCG	3	8	
		UCU	21	19	
		AGC	21	6	
		AGU	15	31	
10	THR (42)	ACA	21	14	
		ACC	41	19	
		ACG	7	14	
		ACU	31	36	
15	PRO (34)	CCA	45	35	
		CCC	19	6	
		CCG	9	21	
		CCU	26	38	
20	ALA (31)	GCA	23	38	
		GCC	32	9	
		GCG	3	3	
		GCU	41	50	
25	GLY (46)	GGA	32	52	
		GGC	20	17	
		GGG	11	15	
		GGU	37	15	
30	ILE (48)	AUA	12	39	
		AUC	45	11	
		AUU	43	50	
35	VAL (38)	GUA	9	45	
		GUC	20	5	
		GUG	28	11	
		GUU	43	39	
40	LYS (3)	AAA	36	100	
		AAG	64	0	
45	ASN (44)	AAC	72	27	
		AAU	28	73	
50				20	

Table V (continued)

Codon in Usage Synthetic <i>B.t.k</i> HD-1 Gene				
Amino Acid	Codon	Percent Usage in Plants/Wt <i>B.t.k</i> /Syn		
GLN (31)	CAA	64	77	61
	CAG	36	23	39
HIS (10)	CAC	65	0	80
	CAU	35	100	20
GLU (30)	GAA	48	87	50
	GAG	52	13	50
ASP (23)	GAC	48	17	65
	GAU	52	83	35
TYR (25)	UAC	68	20	72
	UAU	32	80	28
CYS (2)	UGC	78	50	100
	UGU	22	50	0
PHE (36)	UUC	56	17	83
	UUU	44	83	17
MET (9)	AUG	100	100	100
TRP (9)	UGG	100	100	100

[0077] The resulting synthetic gene lacks ATTAA sequences, contains only one potential polyadenylation site and has a G+C content of 48.5%. Figure 3 is a comparison of the wild-type HD-1 sequence to the synthetic gene sequence for amino acids 1-615. There is approximately 77% DNA homology between the synthetic gene and the wild-type gene and 356 of the 615 codons have been changed (approximately 60%).

#### Example 3 – Synthetic *B.t.k* HD-73 Gene

[0078] The crystal protein toxin from *B.t.k* HD-73 exhibits a higher unit activity against some important agricultural pests. The toxin protein of HD-1 and HD-73 exhibit substantial homology (~90%) in the N-terminal 450 amino acids, but differ substantially in the amino acid region 451-615. Fusion proteins comprising amino acids 1-450 of HD-1 and 451-615 of HD-73 exhibit the insecticidal properties of the wild-type HD-73. The strategy employed was to use the 5'-two thirds of the synthetic HD-1 gene (first 1350 bases, up to the SacI site) and to dramatically modify the final 590 bases (through amino acid 645) of the HD-73 in a manner consistent with the algorithm used to design the synthetic HD-1 gene. Table VI below lists the oligonucleotides used to modify the HD-73 gene in the order used in the gene from 5' to 3' end. Nine oligonucleotides were used in a 590 base pair region, each nucleotide ranging in size from 33 to 60 bases. The only regions left unchanged were areas where there were no long consecutive strings of A or T bases (longer than six). All polyadenylation sites and ATTAA sites were eliminated.

Table VI

Mutagenesis Primers for B.t.k. HD-73

Primer	Length (bp)	Sequence
73K1363	51	AATACTATCG GATGCGATGA TGTTGTTGAA CTCAGCACTA CGGTGTATCC A
73K1437	33	TCCTGAAATG ACAGAACCGT TGAAGAGAAA GTT
73K1471	48	ATTTCCACTG CTGTTGAGTC TAACGAGGTC TCCACCAGTG AATCCTGG
73K1561	60	GTGAATAGGG GTCACAGAAAG CATACCTCAC ACGAACTCTA TATCTGGTAG ATGTTGGATGG
73K1642	33	TCTAGCTGGA ACTGTATTGG AGAAGAGATGGA TGA
73K1675	48	TTCAAAGTAA CCGAAATCGC TGGATTGGAG ATTATCCAAG CAGGTAGC
73K1741	39	ACTAAAGTTT CTAACACCCA CGATGTTACC GAGTGAAGA

Table VI - continued

## Mutagenesis Primers for B.t.k. HD-73

Primer	Length (bp)	Sequence
73K1797	36	AACTGGAATG AACTCGAAC TGTGATAAT CACTCC
73KTERM	54	GGACACTAGA TCTTAGTGAT AATCGGTAC ATTTGCTTG AGTCCAAGCT GGTT

[0079] The resulting gene has two potential polyadenylation sites (compared to 18 in the WT) and no ATTTA sequence (12 in the WT). The G+C content has increased from 37% to 48%. A total of 59 individual base pair changes were made using the primers in Table VI. Overall, there is 90% DNA homology between the region of the HD-73 gene modified by site directed mutagenesis and the wild-type sequence of the analogous region of HD-73. The synthetic HD-73 is a hybrid of the first 1360 bases from the synthetic HD-1 and the next 590 bases or so modified HD-73 sequence. Figure 4 is a comparison of the above-described synthetic B.t.k. HD-73 and the wild-type B.t.k. HD-73 encoding amino acids 1-645. In the modified region of the HD-73 gene 44 of the 170 codons (25%) were changed as a result of the site-directed mutagenesis changes resulting from the oligonucleotides found in Table VI. Overall, approximately 50% of the codons in the synthetic B.t.k. HD-73 differ from the analogous segment of the wild-type and HD-73 gene. [0080] A one base pair deletion in the synthetic HD-73 gene was detected in the course of sequencing the 3' end at base pair 1890. This results in a frame-shift mutation at amino acid 625 with a premature stop codon at amino acid 640 (pMON5379). Table VII below compares the codon usage of the wild-type gene of B.t.k. HD-73 versus the synthetic gene of this example for amino acids 451-645 and codon usage of naturally occurring genes of dicotyledonous plants. The total number of each amino acid encoded in this segment of the gene is found in the parentheses under the amino acid designation.

Table VII

Codon Usage in Synthetic B.t.k. HD-73 Gene				
Amino Acid	Codon	Percent Usage in Plants/Wt HD-73/Syn		
ARG (10)	CGA	7	10	0
	CGC	11	0	8
	CGG	5	10	0
	CGU	25	20	23
	AGA	29	60	62
	AGG	23	0	8

Table VII (continued)

Codon Usage In Synthetic B.t.k. HD-73 Gene				
	Amino Acid	Codon	Percent Usage In Plants/Wt HD-73/Syn	
5	LEU (12)	CUA	8	25
		CUC	20	17
		CUG	10	17
		CUU	28	8
		UUA	5	33
		UUG	30	0
10	SER (21)	UCA	14	24
		UCC	28	10
		UCG	3	10
		UCU	21	24
		AGC	21	0
		AGU	15	33
15	THR (15)	ACA	21	47
		ACC	41	13
		ACG	7	13
		ACU	31	27
		CCA	45	71
		CCC	19	0
20	PRO (7)	CCG	9	14
		CCU	26	14
		ALA (14)	23	29
		GCA	32	31
		GCC	7	8
		GCG	41	43
25	GLY (15)	GCU	3	21
		GGA	41	46
		GGC	32	33
		GGG	20	0
		GGU	11	27
		AUA	37	40
30	ILE (15)	AUC	12	43
		AUU	45	60
		AUA	43	53
		AUC	17	40
		AUU	8	7
		AUA	0	0

Table VII (continued)

Codon Usage In Synthetic B.t.k. HD-73 Gene				
Amino Acid	Codon	Percent Usage In Plants/Wt HD-73/Syn		
VAL (15)	GUA	9	40	7
	GUC	20	0	7
	GUG	28	20	36
	GUU	43	40	50
LYS (3)	AAA	36	67	100
	AAG	64	33	0
ASN (20)	AAC	72	20	53
	AAU	28	80	47
GLN (5)	CAA	64	60	67
	CAG	36	40	33
HIS (3)	CAC	65	67	100
	CAU	35	33	0
GLU (7)	GAA	48	88	57
	GAG	52	14	43
ASP (5)	GAC	48	40	50
	GAU	52	60	50
TYR (5)	UAC	68	0	20
	UAU	32	100	80
CYS (0)	UGC	78	0	0
	UGU	22	0	0
PHE (13)	UUC	56	8	67
	UUU	44	92	33
MET (2)	AUG	100	100	100
TRP (2)	UGG	100	100	100

[0081] Another truncated synthetic HD-73 gene was constructed. The sequence of this synthetic HD-73 gene is identical to that of the above synthetic HD-73 gene in the region in which they overlap (amino acids 29-615), and it also encodes Met-Ala at the N-terminus. Figure 8 shows a comparison of this truncated synthetic HD-73 gene with the N-terminal Met-Ala versus the wild-type HD-73 gene.

[0082] While the previous examples have been directed at the preparation of synthetic and modified genes encoding truncated B.t.k. proteins, synthetic or modified genes can also be prepared which encode full length toxin proteins.

[0083] One full length B.t.k. gene consists of the synthetic HD-73 sequence of Figure 4 from nucleotide 1-1845 plus wild-type HD-73 sequence encoding amino acids 616 to the C-terminus of the native protein. Figure 9 shows a com-

parison of this synthetic/wild-type full length HD-73 gene versus the wild-type full length HD-73 gene.

[0084] Another full length *B.t.k.* gene consists of the synthetic HD-73 sequence of Figure 4 from nucleotide 1-1845 plus a modified HD-73 sequence ending amino acids 616 to the C-terminus of the native protein. The C-terminal portion has been modified by site-directed mutagenesis to remove putative polyadenylation signals and ATTAA sequences according to the algorithm of Figure 1. Figure 10 shows a comparison of this synthetic/modified full length HD-73 gene versus the wild-type full length HD-73 gene.

[0085] Another full length *B.t.k.* gene consists of a fully synthetic HD-73 sequence which incorporates the synthetic HD-73 sequence of Figure 4 from nucleotide 1-1845 plus a synthetic sequence encoding amino acids 616 to the C-terminus of the native protein. The C-terminal synthetic portion has been designed to eliminate putative polyadenylation signals and ATTAA sequences and to include plant preferred codons. Figure 11 shows a comparison of this fully synthetic full length HD-73 gene versus the wild-type full length HD-73 gene.

[0086] Alternatively, another full length *B.t.k.* gene consists of a fully synthetic sequence comprising base pairs 1-1830 of *B.t.k.* HD-1 (Figure 3) and base pairs 1834-3534 of *B.t.k.* HD-73 (Figure 11).

15 **Example 4 -- Expression of Modified and Synthetic *B.t.k.* HD-1 and Synthetic HD-73**

[0087] A number of plant transformation vectors for the expression of *B.t.k.* genes were constructed by incorporating the structural coding sequences of the previously described genes into plant transformation cassette vector pMON893.

20 The respective intermediate transformation vector is inserted into a suitable disarmed *Agrobacterium* vector such as *A. tumefaciens* ACO, supra. Tissue explants are cocultured with the disarmed *Agrobacterium* vector and plants regenerated under selection for kanamycin resistance using known protocols: tobacco (Horsch et al., 1985); tomato (McCormick et al., 1986) and cotton (Trolinder et al., 1987).

a) **Tobacco.**

25 [0088] The level of *B.t.k.* HD-1 protein in transgenic tobacco plants containing pMON9921 (wild type truncated), pMON5370 (modified HD-1, Example 1, Figure 2) and pMON5377 (synthetic HD-1, Example 2, Figure 3) were analyzed by Western analysis. Leaf tissue was frozen in liquid nitrogen, ground to a fine powder and then ground in a 12 (wt:volume) of SDS-PAGE sample buffer. Samples were frozen on dry ice, then incubated for 10 minutes in a boiling water bath and microfuged for 10 minutes. The protein concentration of the supernatant was determined by the method of Bradford (Anal. Biochem. 72:248-254). Fifty ug of protein was run per lane on 9% SDS-PAGE gels, the protein transferred to nitrocellulose and the *B.t.k.* HD-1 protein visualized using antibodies produced against *B.t.k.* HD-1 protein as the primary antibody and alkaline phosphatase conjugated second antibody as described by the manufacturer (Promega, Madison, WI). Purified HD-1 tryptic fragment was used as the control. Whereas the *B.t.k.* protein from tobacco plants containing pMON9921 was below the level of detection, the *B.t.k.* protein from plants containing the modified (pMON5370) and synthetic (pMON5377) genes was easily detected. The *B.t.k.* protein from plants containing pMON9921 remained undetectable, even with 10 fold longer incubation times. The relative levels of *B.t.k.* HD-1 protein in these plants is estimated in Table VIII. Because the protein from plants containing pMON9921 was not observed, the level of protein in these plants was estimated from the relative mRNA levels (see below). Plants containing the modified gene (pMON5370) expressed approximately 100 fold more *B.t.k.* protein than plants containing the wild-type gene (pMON9921). Plants containing the fully synthetic *B.t.k.* HD-1 gene (pMON5377) expressed approximately five fold more protein than plants containing the modified gene. The modified gene contributes the majority of the increase in *B.t.k.* expression observed. The plants used to generate the above data are the best representatives from each construct based either on a tobacco hornworm bioassay or on data derived from previous Western analysis.

45 Table VIII

Expression of <i>B.t.k.</i> HD-1 Protein in Transgenic Tobacco			
Gene Description	Vector	<i>B.t.k.</i> Protein* Concentration	Fold Increase in <i>B.t.k.</i> Expression
Wild type	pMON9921	10	1
Modified	pMON5370	1000	100
Synthetic	pMON5377	5000	500

55 \* *B.t.k.* protein concentrations are expressed in ng/mg of total soluble protein. The level of *B.t.k.* protein for plants containing the wild type gene are estimated from mRNA levels.

[0089] Plants containing these genes were tested for bioactivity to determine whether the increased quantities of

protein observed by Western analysis result in a corresponding increase in bioactivity. Leaves from the same plants used for the Western data in Table I were tested for bioactivity against two insects. A detached leaf bioassay was first done using tobacco hornworm, an extremely sensitive lepidopteran insect. Leaves from all three transgenic tobacco plants were totally protected and 100% mortality of tobacco hornworm observed (see Table IX below). A much less sensitive insect, beet armyworm, was then used in another detached leaf bioassay. Beet armyworm is approximately 500 fold less sensitive to *B.t.k.* HD-1 protein than tobacco hornworm. The difference in sensitivity of these two insects was determined using purified HD-1 protein in a diet incorporation assay (see below). Plants containing the wild-type gene (pMON9921) showed only minimal protection against beet armyworm, whereas plants containing the modified gene showed almost complete protection and plants containing the fully synthetic gene were totally protected against beet armyworm damage. The results of these bioassays confirm the levels of *B.t.k.* HD-1 expression observed in the Western analysis and demonstrates that the increased levels of *B.t.k.* HD-1 protein correlates with increased insecticidal activity.

Table IX

Protection of Tobacco Plants from Tobacco Hornworm and Beet Armyworm			
Gene Description	Vector	Tobacco Hornworm Damage*	Beet Armyworm Damage*
None	None	NL	NL
Wild type	pMON9921	0	3
Modified	pMON5370	0	1
Synthetic	pMON5377	0	0

\* Extent of insect damage was rated: 0, no damage; 1, slight; 2, moderate; 3, severe; or NL, no leaf left.

[0090] The bioactivity of the *B.t.k.* HD-1 protein produced by these transgenic plants was further investigated to more accurately quantitate the relative activities. Leaf tissue from tobacco plants containing the wild-type, modified and synthetic genes were ground in 100 mM sodium carbonate buffer, pH 10 at a 1:2 (wt/vol) ratio. Particulate material was removed by centrifugation. The supernatant was incorporated into a synthetic diet similar to that described by Marrone et al. (1985). The diet medium was prepared the day of the test with the plant extract solutions incorporated in place of the 20% water component. One ml of the diet was aliquoted into 96 well plates.

[0091] After the diet dried, one neonate tobacco budworm larva was added to each well. Sixteen insects were tested with each plant sample. The plants were incubated at 27°C. After seven days, the larvae from each treatment were combined and weighed on an analytical balance. The average weight per insect was calculated and compared to a standard curve relating *B.t.k.* protein concentrations to average larval weight. Insect weight was inversely proportional (in a logarithmic manner) to the relative increase in *B.t.k.* protein concentration. The amount of *B.t.k.* HD-1 protein, based on the extent of larval growth inhibition was determined for two different plants containing each of the three genes. The specific activity (ng of *B.t.k.* HD-1 per mg of plant protein) was determined for each plant. Plants containing the modified HD-1 gene (pMON5370) averaged approximately 1400 ng (1200 and 1600 ng) of *B.t.k.* HD-1 per mg of plant extract protein. This value compares closely with the 1000 ng of *B.t.k.* HD-1 protein per mg of plant extract protein as determined by Western analysis (Table I). *B.t.k.* HD-1 concentrations for the plants containing the synthetic HD-1 gene averaged approximately 8200 ng (7200 and 9200 ng) of *B.t.k.* HD-1 protein per mg of plant extract protein. This number compares well to the 5000 ng of HD-1 protein per mg of plant extract protein estimated by Western analysis. Likewise, plants containing the synthetic gene showed approximately a six-fold higher specific activity than the corresponding plants containing the modified gene for these bioassays. In the Western analysis the ratio was approximately 10 fold, again both are in good agreement. The level of *B.t.k.* protein in plants containing the wild-type HD-1 gene (pMON9921) was too low to give a significant decrease in larval weight and hence was below a level that could be quantitated in this assay. In conclusion, the levels of *B.t.k.* HD-1 protein determined by both the bioassays and the Western analysis for these plants containing the modified and synthetic genes agree, which demonstrates that the *B.t.k.* HD-1 protein produced by these plants is biologically active.

[0092] The levels of mRNA were determined in the plants containing the wild-type *B.t.k.* HD-1 gene (pMON9921) and the modified gene (pMON5370) to establish whether the increased levels of protein production result from increased transcription or translation. mRNA from plants containing the synthetic gene could not be analyzed directly with the same DNA probe as used for the wild-type and modified genes because of the numerous changes made in the coding sequence. mRNA was isolated and hybridized with a single-stranded DNA probe homologous to approximately the 5' 90 bp of the wild-type or modified gene coding sequences. The hybrids were digested with S1 nuclease and the protected probe fragments analyzed by gel electrophoresis. Because the procedure used a large excess of probe and long hybridization time, the amount of protected probe is proportional to the amount of *B.t.k.* mRNA present in the sample. Two plants expressing the modified gene (pMON5370) were found to produce up to ten-fold more RNA

than a plant expressing the wild-type gene (pMON9921).

[0093] The increased mRNA level from the modified gene is consistent with the result expected from the modifications introduced into this gene. However, this 10 fold increase in mRNA with the modified gene compared to the wild-type gene is in contrast to the 100 fold increase in *B.t.k.* protein from these genes in tobacco plants. If the two mRNAs were equally well translated then a 10 fold increase in stable mRNA would be expected to yield a 10 fold increase in protein. The higher increase in protein indicates that the modified gene mRNA is translated at about a 10 fold higher efficiency than wild-type. Thus, about half of the total effect on gene expression can be explained by changes in mRNA levels and about half to changes in translational efficiency. This increase in translational efficiency is striking in that only about 9.5% of the codons have been changed in the modified gene; that is, this effect is clearly not due to wholesale codon usage changes. The increased translational efficiency could be due to changes in mRNA secondary structure that affect translation or to the removal of specific translational blockades due to specific codons that were changed.

[0094] The increased expression seen with the synthetic HD-1 gene was also seen with a synthetic HD-73 gene in tobacco. *B.t.k.* HD-73 was undetected in extracts of tobacco plants containing the wild-type truncated HD-73 gene (pMON5367), whereas *B.t.k.* HD-73 protein was easily detected in extracts from tobacco plants containing the synthetic HD-73 gene of Figure 4 (pMON5383). Approximately 1000 ng of *B.t.k.* HD-73 protein was detected per mg of total soluble plant protein.

[0095] As described in Example 3 above, the *B.t.k.* HD-73 protein encoded in pMON5383 contains a small C-terminal extension of amino acids not encoded in the wild-type HD-73 protein. These extra amino acids had no effect on insect toxicity or on increased plant expression. A second synthetic HD-73 gene was constructed as described in Example 20 3 (Figure 8) and used to transform tobacco (pMON5390). Analysis of plants containing pMON5390 showed that this gene was expressed at levels comparable to that of pMON5383 and that these plants had similar insecticidal efficacy.

[0096] In tobacco plants the synthetic HD-1 gene was expressed at approximately a 5-fold higher level than the synthetic HD-73 gene. However, this synthetic HD-73 gene still was expressed at least 100-fold better than the wild-type HD-73 gene. The HD-73 protein is approximately 5-fold more toxic to many insect pests than the HD-1 protein, so both synthetic HD-1 and HD-73 genes provide approximately comparable insecticidal efficacy in tobacco.

[0097] The full length *B.t.k.* HD-73 genes described in Example 3 were also incorporated into the plant transformation vector pMON893 so that they were expressed from the En 35S promoter. The synthetic/wild-type full length HD-73 gene of Figure 9 was incorporated into pMON893 to create pMON10505. The synthetic/modified full length HD-73 gene of Figure 10 was incorporated into pMON893 to create pMON10526. The fully synthetic HD-73 gene of Figure 11 was incorporated into pMON893 to create pMON10518. These vectors were used to obtain transformed tobacco plants, and the plants were analyzed for insecticidal efficacy and for *B.t.k.* HD-73 protein levels by Western blot or ELISA Immunoassay.

[0098] Tobacco plants containing all three of these full length *B.t.k.* genes produced detectable *B.t.k.* protein and showed 100% mortality of tobacco hornworm. This result is surprising in light of previous reported attempts to express the full length *B.t.k.* genes in transgenic plants. Vaect et al. (1987) reported that a full length *B.t.k. berolin* gene similar to our HD-1 gene could not be detectably expressed in tobacco. Barton et al. (1987) reported a similar result for another full length gene from *B.t.k.* HD-1 (the so called 4.5 kb gene), and further indicated that tobacco callus containing this gene became necrotic, indicating that the full length gene product was toxic to plant cells. Fischhoff et al. (1987) reported that the full length *B.t.k.* HD-1 gene in tomato was poorly expressed compared to a truncated gene, and no plants that were fully toxic to tobacco hornworm could be recovered. All three of the above reports indicated much higher expression levels and recovery of toxic plants if the respective *B.t.k.* genes were truncated. Adang et al. reported that the full length HD-73 gene yielded a few tobacco plants with some biological activity (none were highly toxic) against hornworm and barely detectable *B.t.k.* protein. It was also noted by them that the major *B.t.k.* mRNA in these plants was a truncated 1.7 kb species that would not encode a functional toxin. This indicated improper expression of the gene in tobacco. In contrast to all of these reports, the three full length *B.t.k.* HD-73 genes described above all lead to relatively high levels of protein and high levels of insect toxicity.

[0099] *B.t.k.* protein and mRNA levels in tobacco plants are shown in Table X for these three vectors. As can be seen from the table, the synthetic/wild-type gene (pMON10506) produces *B.t.k.* protein as about 0.01% of total soluble protein; the synthetic/modified gene produces *B.t.k.* as about 0.02% of total soluble protein; and the fully synthetic gene produces *B.t.k.* as about 0.2% of total soluble protein. *B.t.k.* mRNA was analyzed in these plants by Northern blot analysis using the common 5' synthetic half of the genes as a probe. As shown in Table X, the increased protein levels can largely be attributed to increased mRNA levels. Compared to the truncated modified and synthetic genes, this could indicate that the major contributors to increased translational efficiency are in the 5' half of the gene while the 3' half of the gene contains mostly determinants of mRNA stability. The increased protein levels also indicate that increasing the amount of the full length gene that is synthetic or modified increases *B.t.k.* protein levels. Compared to the truncated synthetic *B.t.k.* HD-73 genes (pMON5383 or pMON5390), the fully synthetic gene (pMON10518) produces as much or slightly more *B.t.k.* protein demonstrating that the full length genes are capable of being expressed at high levels in plants. These tobacco plants with high levels of full length HD-73 protein show no evidence of abnor-

5 mally and are fully fertile. The *B.t.k.* protein levels in these plants also produce the expected levels of insect toxicity based on feeding studies with beet armyworm or diet incorporation assays of plant extracts with tobacco budworm. The *B.t.k.* protein detected by Western blot analysis in these tobacco plants often contains a varying amount of protein of about 80 kDa which is apparently a proteolytic fragment of the full length protein. The C-terminal half of the full length protein is known to be proteolytically sensitive, and similar proteolytic fragments are seen from the full length gene in *E. coli* and *B.t.* itself. These fragments are fully insecticidal. The Northern analysis indicated that essentially all of the mRNA from these full length genes was of the expected full length size. There is no evidence of truncated mRNAs that could give rise to the 80 kDa protein fragment. In addition, it is possible that the fragment is not present in intact plant cells and is merely due to proteolysis during extraction for immunoassay.

10

Table X

Full Length <i>B.t.k.</i> HD-73 Protein and mRNA Levels In Transgenic Tobacco Plants			
Gene description	Vector	B.t.k. protein concentration	Relative B.t.k. mRNA level
Synthetic/wild type	pMON10508	>100	0.5
Synthetic/modified	pMON10528	400	1
Fully synthetic	pMON10518	>2000	40

20 [0100] Thus, there is no serious impediment to producing high levels of *B.t.k.* HD-73 protein in plants from synthetic genes, and this is expected to be true of other full length lepidopteran active genes such as *B.t.k.* HD-1 or *B.t. entomocidus*. The fully synthetic *B.t.k.* HD-1 gene of Example 3 has been assembled in plant transformation vectors such as pMON893.

25 [0101] The fully synthetic gene in pMON10518 was also utilized in another plant vector and analyzed in tobacco plants. Although the CaMV35S promoter is generally a high level constitutive promoter in most plant tissues, the expression level of genes driven by the CaMV35S promoter is low in floral tissue relative to the levels seen in leaf tissue. Because the economically important targets damaged by some insects are the floral parts or derived from floral parts (e.g., cotton squares and bolls, tobacco buds, tomato buds and fruit), it may be advantageous to increase the expression of *B.t.* protein in these tissues over that obtained with the CaMV35S promoter.

30 [0102] The 35S promoter of Figwort Mosaic Virus (FMV) is analogous to the CaMV35S promoter. This promoter has been isolated and engineered into a plant transformation vector analogous to pMON893. Relative to the CaMV promoter, the FMV 35S promoter is highly expressed in the floral tissue, while still providing similar high levels of gene expression in other tissues such as leaf. A plant transformation vector, pMON10517, was constructed in which the full length synthetic *B.t.k.* HD-73 gene of Figure 11 was driven by the FMV 35S promoter. This vector is identical to pMON10518 of Example 3 except that the FMV promoter is substituted for the CaMV promoter. Tobacco plants transformed with pMON10517 and pMON10518 were obtained and compared for expression of the *B.t.k.* protein by Western blot or ELISA immunoassay in leaf and floral tissue. This analysis showed that pMON10517 containing the FMV promoter expressed the full length HD-73 protein at higher levels in floral tissue than pMON10518 containing the CaMV promoter. Expression of the full length *B.t.k.* HD-73 protein from pMON10517 in leaf tissue is comparable to that seen with the most highly expressing plants containing pMON10518. However, when floral tissue was analyzed, tobacco plants containing pMON10518 that had high levels of *B.t.k.* protein in leaf tissue did not have detectable *B.t.k.* protein in the flowers. On the other hand, flowers of tobacco plants containing pMON10517 had levels of *B.t.k.* protein nearly as high as the levels in leaves at approximately 0.05% of total soluble protein. This analysis showed that the FMV promoter could be used to produce relatively high levels of *B.t.k.* protein in floral tissue compared to the CaMV promoter.

45 b) Tomato.

50 [0103] The wild-type, modified and synthetic *B.t.k.* HD-1 genes tested in tobacco were introduced into other plants to demonstrate the broad utility of this invention. Transgenic tomatoes were produced which contain these three genes. Data show that the increased expression observed with the modified and synthetic gene in tobacco also extends to tomato. Whereas the *B.t.k.* HD-1 protein is only barely detectable in plants containing the wild type HD-1 gene (pMON9921), *B.t.k.* HD-1 was readily detected and the levels determined for plants containing the modified (pMON5370) or synthetic (pMON5377) genes. Expression levels for the plants containing the wild-type, modified and synthetic HD-1 genes were approximately 10, 100 and 500 ng per mg of total plant extract (see Table XI below). The increase in *B.t.k.* HD-1 protein for the modified gene accounted for the majority of increase observed; 10 fold higher than the plants containing the wild-type gene, compared to only an additional five-fold increase for plants containing the synthetic gene. Again the site-directed changes made in the modified gene are the major contributors to the increased expression of *B.t.k.* HD-1.

Table XI

B.t.k. HD-1 Expression In Transgenic Tomato Plants			
Gene Description	Vector	B.t.k. Protein* Concentration	Fold Increase in B.t.k. Expression
Wild type	pMON9921	10	1
Modified	pMON5370	100	10
Synthetic	pMON5377	500	50

\* B.t.k. HD-1 protein concentrations are expressed in ng/mg of total soluble plant protein. Data for plants containing the wild-type gene are estimates from mRNA levels and protein levels determined by ELISA.

[0104] These differences in B.t.k. HD-1 expression were confirmed with bioassays against tobacco hornworm and beet armyworm. Leaves from tomato plants containing each of these genes controlled tobacco hornworm damage and produced 100% mortality. With beet armyworm, leaves from plants containing the wild-type HD-1 gene (pMON9921) showed significant damage, leaves from plants containing the modified gene (pMON5370) showed less damage and leaves from plants containing the synthetic gene (pMON5377) were completely protected (see Table XII below).

Table XII

Protection of Tomato Plants from Tobacco Hornworm and Beet Armyworm			
Gene Description	Vector	Tobacco Hornworm Damage*	Beet Armyworm Damage*
None	None	NL	NL
Wild type	pMON9921	0	3
Modified	pMON5370	0	1
Synthetic	pMON5377	0	0

\* Damage was rated as shown in Table IX.

[0105] The generality of the synthetic gene approach was extended in tomato with a synthetic B.t.k. HD-73 gene.

[0106] In tomato, extracts from plants containing the wild-type truncated HD-73 gene (pMON5367) showed no detectable HD-73 protein. Extracts from plants containing the synthetic HD-73 gene (pMON5383) showed high levels of B.t.k. HD-73 protein, approximately 2000 ng per mg of plant extract protein. These data clearly demonstrate that the changes made in the synthetic HD-73 gene lead to dramatic increases in the expression of the HD-73 protein in tomato as well as in tobacco.

[0107] In contrast to tobacco, the synthetic HD-73 gene in tomato is expressed at approximately 4-fold to 5-fold higher levels than the synthetic HD-1 gene. Because the HD-73 protein is about 5-fold more active than the HD-1 protein against many insect pests including *Heliothis* species, the increased expression of synthetic HD-73 compared to synthetic HD-1 corresponds to about a 25-fold increased insecticidal efficacy in tomato.

[0108] In order to determine the mechanisms involved in the increased expression of modified and synthetic B.t.k. HD-1 genes in tomato, S1 nuclease analysis of mRNA levels from transformed tomato plants was performed. As indicated above, a similar analysis had been performed with tobacco plants, and this analysis showed that the modified gene produced up to 10-fold more mRNA than the wild-type gene. The analysis in tomato utilized a different DNA probe that allowed the analysis of wild-type (pMON9921), modified (pMON5370) and synthetic (pMON5377) HD-1 genes with the same probe. This probe was derived from the 5' untranslated region of the CaMV35S promoter in pMON893 that was common to all three of these vectors (pMON9921, pMON5370 and pMON5377). This S1 analysis indicated that B.t.k. mRNA levels from the modified gene were 3 to 5 fold higher than for the wild-type gene, and that mRNA levels for the synthetic gene were about 2 to 3 fold higher than for the modified gene. Three independent transformants were analyzed for each gene. Compared to the fold increases in B.t.k. HD-1 protein from these genes in tomato shown

in Table XI, these mRNA increases can explain about half of the total protein increase as was seen in tobacco for the wild-type and modified genes. For tomato the total mRNA increase from wild-type to synthetic is about 6 to 15 fold compared to a protein increase of about 50 fold. This result is similar to that seen for tobacco in comparing the wild-type and modified genes, and it extends to the synthetic gene as well. That is, about half of the total fold increase in B.t.k. protein from wild-type to modified genes can be explained by mRNA increases and about half to enhanced translational efficiency. The same is also true in comparing the modified gene to the synthetic gene. Although there is an additional increase in RNA levels, this mRNA increase can explain only about half of the total protein increase.

[0109] The full length B.t.k. genes described above were also used to transform tomato plants and these plants were

analyzed for *B.t.k* protein and insecticidal efficacy. The results of this analysis are shown in Table XIII. Plants containing the synthetic/wild-type gene (pMON10506) produce the *B.t.k* HD-73 protein at levels of about 0.01% of their total soluble protein. Plants containing the synthetic/modified gene (pMON10526) produce about 0.04% *B.t.k* protein, and plants containing the fully synthetic gene (pMON10518) produce about 0.2% *B.t.k* protein. These results are very similar to the tobacco plant results for the same genes. mRNA levels estimated by Northern blot analysis in tomato also increase in parallel with the protein level increase. As for tobacco with these three genes, most of the protein increase can be attributed to increased mRNA with a small component of translational efficiency increase indicated for the fully synthetic gene. The highest levels of full length *B.t.k* protein (from pMON10518) are comparable to or just slightly lower than the highest levels observed for the truncated HD-73 genes (pMON5383 and pMON5390). Tomato plants expressing these full length genes have the insecticidal activity expected for the observed protein levels as determined by feeding assays with beet armyworm or by diet incorporation of plant extracts with tobacco hornworm.

Table XIII

Full Length <i>B.t.k</i> HD-73 Protein and mRNA Levels In Transgenic Tomato Plants			
Gene description	Vector	<i>B.t.k</i> protein concentration	Relative <i>B.t.k</i> mRNA level
Synthetic/wild type	pMON10506	100	1
Synthetic/modified	pMON10526	400	2.4
Fully synthetic	pMON10518	2000	10

c) Cotton.

[0110] The generality of the increased expression of *B.t.k* HD-1 and *B.t.k* HD-73 by use of the modified and synthetic genes was extended to cotton. Transgenic calli were produced which contain the wild type (pMON9921) and the synthetic HD-1 (pMON5377) genes. Here again the *B.t.k* HD-1 protein produced from calli containing the wild-type gene was not detected, whereas calli containing the synthetic HD-1 gene expressed the HD-1 protein at easily detectable levels. The HD-1 protein was produced at approximately 1000 ng/mg of plant calli extract protein. Again, to ensure that the protein produced by the transgenic cotton calli was biologically active and that the increased expression observed with the synthetic gene translated to increased biological activity, extracts of cotton calli were made in similar manner as described for tobacco plants, except that the calli was first dried between Whatman filter paper to remove as much of the water as possible. The dried calli were then ground in liquid nitrogen and ground in 100 mM sodium carbonate buffer, pH 10. Approximately 0.5 ml aliquotes of this material was applied to tomato leaves with a paint brush. After the leaf dried, five tobacco hornworm larvae were applied to each of two leaf samples. Leaves painted with extract from control calli were completely destroyed. Leaves painted with extract from calli containing the wild-type HD-1 gene (pMON9921) showed severe damage. Leaves painted with extract from calli containing the synthetic HD-1 gene (pMON5377) showed no damage (see Table XIV below).

Table XIV

Protection against Tobacco Hornworm by Tomato Leaves Painted with Extracts Prepared from Cotton Calli Containing a Control, the Wild-Type <i>B.t.k</i> HD-1 Gene, Synthetic HD-1 Gene or Synthetic HD-73 Gene		
Gene Description	Vector	Tobacco Hornworm Damage*
Control	Control	NL
Wild type HD-1	pMON9921	3
Synthetic HD-1	pMON5377	0
Synthetic HD-73	pMON5383	0

\* Damage was rated as shown in Table IX.

[0111] Cotton calli were also produced containing another synthetic gene, a gene encoding *B.t.k* HD-73. The preparation of this gene is described in Example 3. Calli containing the synthetic HD-73 gene produced the corresponding HD-73 protein at even higher levels than the calli which contained the synthetic HD-1 gene. Extracts made from calli containing the HD-73 synthetic gene (pMON5383) showed complete control of tobacco hornworm when painted onto tomato leaves as described above for extracts containing the HD-1 protein. (See Table XV).

[0112] Transgenic cotton plants containing the synthetic *B.t.k* HD-1 gene (pMON5377) or the synthetic *B.t.k* HD-73 gene (pMON5383) have also been examined. These plants produce the HD-1 or HD-73 proteins at levels comparable to that seen in cotton callus with the same genes and comparable to tomato and tobacco plants with these genes.

For either synthetic truncated HD-1 or HD-73 genes, cotton plants expressing *B.t.k.* protein at 1000 to 2000 ng/mg total protein (0.1% to 0.2%) were recovered at a high frequency. Insect feeding assays were performed with leaves from cotton plants expressing the synthetic HD-1 or HD-73 genes. These leaves showed no damage (rating of 0) when challenged with larvae of cabbage looper (*Trichoplusia ni*), and only slight damage when challenged with larvae of 5 beet armyworm (*Spodoptera exigua*). Damage ratings are as defined in Table IX above. This demonstrated that cotton plants as well as calli expressed the synthetic HD-1 or HD-73 genes at high levels and that those plants were protected from damage by Lepidopteran insect larvae.

[0113] Transgenic cotton plants containing either the synthetic truncated HD-1 gene (pMON5377) or the synthetic truncated HD-73 gene (pMON5383) were also assessed for protection against cotton bollworm at the whole plant level 10 in the greenhouse. This is a more realistic test of the ability of these plants to produce an agriculturally acceptable level of control. The cotton bollworm (*Heliothis zea*) is a major pest of cotton that produces economic damage by destroying terminals, squares and bolls, and protection of these fruiting bodies as well as the leaf tissue will be important for effective insect control and adequate crop protection. To test the protection afforded to whole plants, *B.t.k.* progeny of cotton plants expressing high levels of either *B.t.k.* HD-1 (pMON5377) or *B.t.k.* HD-73 (pMON5383) were assayed by 15 applying 10-15 eggs of cotton bollworm per boll or square to the 20 uppermost squares or bolls on each plant. At least 12 plants were analyzed per treatment. The hatch rate of the eggs was approximately 70%. This corresponds to very high insect pressure compared to numbers of larvae per plant seen under typical field conditions. Under these conditions 100% of the bolls on control cotton plants were destroyed by insect damage. For the transgenics, significant boll protection was observed. Plants containing pMON5377 (HD-1) had 70-75% of the bolls survive the intense pressure 20 of this assay. Plants containing pMON5383 (HD-73) had 80% to 90% boll protection. This is likely to be a consequence of the higher activity of HD-73 protein against cotton bollworm compared to HD-1 protein. In cases where the transgenic plants were damaged by the insects, the surviving larvae were delayed in their development by at least one instar.

[0114] Therefore, the increased expression obtained with the modified and synthetic genes is not limited to any one 25 crop; tobacco, tomato and cotton calli and cotton plants all showed drastic increases in *B.t.k.* expression when the plants/callus were produced containing the modified or synthetic genes. Likewise, the utility of changes made to produce the modified and synthetic *B.t.k.* HD-1 gene is not limited to the HD-1 gene. The synthetic HD-73 gene in all three species also showed drastic increases in expression.

[0115] In summary, it has been demonstrated that: (1) the genetic changes made in the HD-1 modified gene lead to very significant increases in *B.t.k.* HD-1 expression; (2) production of a totally synthetic gene lead to a further five-fold 30 increase in *B.t.k.* HD-1 expression; (3) the changes incorporated into the modified HD-1 gene accounted for the majority of the increased *B.t.k.* expression observed with the synthetic gene; (4) the increased expression was demonstrated in three different plants -- tobacco plants, tomato plants and cotton calli and cotton plants; (5) the increased expression as observed by Western analysis also correlated with similar increases in bioactivity, showing that the *B.t.k.* HD-1 proteins produced were comparably active; (6) when the method of the present invention used to design the synthetic 35 HD-1 gene was employed to design a synthetic HD-73 gene it also was expressed at much higher levels in tobacco, tomato and cotton than the wild-type equivalent gene with consequent increases in bioactivity; (7) a fully synthetic full length *B.t.k.* gene was expressed at levels comparable to synthetic truncated genes.

#### Example 5 -- Synthetic *B.t. tenebrionis* Gene in Tobacco, Tomato and Potato

[0116] Referring to Figure 12, a synthetic gene encoding a Coleopteran active toxin is prepared by making the indicated changes in the wild-type gene of *B.t. tenebrionis* or de novo synthesis of the synthetic structural gene. The synthetic gene is inserted into an intermediate plant transformation vector such as pMON893. Plasmid pMON893 containing the synthetic *B.t.t.* gene is then inserted into a suitable disarmed *Agrobacterium* strain such as *A. tumefaciens* 45 ACO.

#### Transformation and Regeneration of Potato

[0117] Sterile shoot cultures of Russet Burbank are maintained in vials containing 10 ml of PM medium (Murashige and Skoog (MS) Inorganic salts, 30 g/l sucrose, 0.17 g/l  $\text{NaH}_2\text{PO}_4\text{H}_2\text{O}$ , 0.4 mg/l thiamine-HCl, and 100 mg/l myoinositol, solidified with 1 g/l Gelrite at pH 6.0). When shoots reached approximately 5 cm in length, stem internode segments of 7-10 mm are excised and smeared at the cut ends with a disarmed *Agrobacterium tumefaciens* vector containing the synthetic *B.t.t.* gene from a four day old plate culture. The stem explants are co-cultured for three days at 23°C on a sterile filter paper placed over 1.5 ml of a tobacco cell feeder layer overlaid on 1/10 P medium (1/10 strength MS Inorganic salts and organic addenda without casein as in Jarret et al. (1980), 30 g/l sucrose and 8.0 g/l agar). Following co-culture the explants are transferred to full strength P-1 medium for callus induction, composed of MS Inorganic salts, organic additions as in Jarret et al. (1980) with the exception of casein, 3.0 mg/l benzyladenine (BA), and 0.01 mg/l naphthaleneacetic acid (NAA) (Jarret, et al., 1980). Carbenicillin (500 mg/l) is included to inhibit 50

bacterial growth, and 100 mg/l kanamycin is added to select for transformed cells. After four weeks the explants are transferred to medium of the same composition but with 0.3 mg/l gibberellic acid (GA3) replacing the BA and NAA (Jarret et al., 1981) to promote shoot formation. Shoots begin to develop approximately two weeks after transfer to shoot induction medium; these are excised and transferred to vials of PM medium for rooting. Shoots are tested for kanamycin resistance conferred by the enzyme neomycin phosphotransferase II, by placing a section of the stem onto callus induction medium containing MS organic and inorganic salts, 30 g/l sucrose, 2.25 mg/l BA, 0.186 mg/l NAA, 10 mg/l GA3 (Webb, et al., 1983) and 200 mg/l kanamycin to select for transformed cells.

[0118] The synthetic *B.t.t* gene described in figure 12, was placed into a plant expression vector as described in example 5. The plasmid has the following characteristics; a synthetic BglII fragment having approximately 1800 base pairs was inserted into pMON893 in such a manner that the enhanced 35S promoter would express the *B.t.t* gene. This construct, pMON1982, was used to transform both tobacco and tomato. Tobacco plants, selected as kanamycin resistant plants were screened with rabbit anti-*B.t.t* antibody. Cross-reactive material was detected at levels predicted to be suitable to cause mortality to CPB. These target insects will not feed on tobacco, but the transgenic tobacco plants do demonstrate that the synthetic gene does improve expression of this protein to detectable levels.

[0119] Tomato plants with the pMON1982 construct were determined to produce *B.t.t* protein at levels insecticidal to CPB. In initial studies, the leaves of four plants (5190, 5225, 5328 and 5133) showed little or no damage when exposed to CPB larvae (damage rating of 0-1 on a scale of 0 to 4 with 4 as no leaf remaining). Under these conditions the control leaves were completely eaten. Immunological analysis of these plants confirmed the presence of material cross-reactive with anti-*B.t.t* antibody. Levels of protein expression in these plants were estimated at approximately 1 to 5 ng of *B.t.t* protein in 50 ug of total extractable protein. A total of 17 tomato plants (17 of 65 tested) have been identified which demonstrate protection of leaf tissue from CPB (rating of 0 or 1) and show good insect mortality.

[0120] Results similar to those seen in tobacco and tomato with pMON1982 were seen with pMON1984 in the same plant species. pMON1984 is identical to pMON1982 except that the synthetic protease inhibitor (CMT) is fused upstream of the native proteolytic cleavage site. Levels of expression in tobacco were estimated to be similar to pMON1982, between 10-15 ng per 50ug of total soluble protein.

[0121] Tomato plants expressing pMON1984 have been identified which protect the leaves from ingestion by CPB. The damage rating was 0 with 100% insect mortality.

[0122] Potato was transformed as described in example 5 with a vector similar to pMON1982 containing the enhanced CaMV35S/synthetic *B.t.t* gene. Leaves of potato plants transformed with this vector, were screened by CPB insect bioassay. Of the 35 plants tested, leaves from 4 plants, 16a, 13c, 13d, and 23a were totally protected when challenged. Insect bioassays with leaves from three other plants, 13e, 1a, and 13b, recorded damage levels of 1 on a scale of 0 to 4 with 4 being total devastation of the leaf material. Immunological analysis confirmed the presence of *B.t.t* cross-reactive material in the leaf tissue. The level of *B.t.t* protein in leaf tissue of plant 16a (damage rating of 0) was estimated at 20-50 ng of *B.t.t* protein/50 ug of total soluble protein. The levels of *B.t.t* protein seen in 16a tissue was consistent with its biological activity. Immunological analysis of 13e and 13b (tissue which scored 1 in damage rating) reveal less protein (5-10 ng/50 ug of total soluble protein) than in plant 16a. Cuttings of plant 16a were challenged with 50 to 200 eggs of CPB in a whole plant assay. Under these conditions 16a showed no damage and 100% mortality of insects while control potato plants were heavily damaged.

#### 40 Example 6 -- Synthetic *B.t.k*, P2 Protein Gene

[0123] The P2 protein is a distinct insecticidal protein produced by some strains of *B.t*. Including *B.t.k* HD-1. It is characterized by its activity against both lepidopteran and dipteran insects (Yamamoto and Iizuka, 1983). Genes encoding the P2 protein have been isolated and characterized (Donovan et al., 1988). The P2 proteins encoded by these genes are approximately 600 amino acids in length. These proteins share only limited homology with the lepidopteran specific P1 type proteins, such as the *B.t.k* HD-1 and HD-73 proteins described in previous examples.

[0124] The P2 proteins have substantial activity against a variety of lepidopteran larvae including cabbage looper, tobacco hornworm and tobacco budworm. Because they are active against agronomically important insect pests, the P2 proteins are a desirable candidate in the production of insect tolerant transgenic plants either alone or in combination with the other *B.t* toxins described in the above examples. In some plants, expression of the P2 protein alone might be sufficient to provide protection against damaging insects. In addition, the P2 proteins might provide protection against agronomically important dipteran pests. In other cases, expression of P2 together with the *B.t.k* HD-1 or HD-73 protein might be preferred. The P2 proteins should provide at least an additive level of insecticidal activity when combined with the crystal protein toxin of *B.t.k* HD-1 or HD-73, and the combination may even provide a synergistic activity.

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Although the mode of action of the P2 protein is unknown, its distinct amino acid sequence suggests that it functions differently from the *B.t.k* HD-1 and HD-73 type of proteins. Production of two insect tolerance proteins with different modes of action in the same plant would minimize the potential for development of insect resistance to *B.t* proteins in plants. The lack of substantial DNA homology between P2 genes and the HD-1 and HD-73 genes minimizes the po-

tential for recombination between multiple insect tolerance genes in the plant chromosome.

[0125] The genes encoding the P2 protein although distinct in sequence from the *B.t.k.* HD-1 and HD-73 genes share many common features with these genes. In particular, the P2 protein genes have a high A+T content (65%), multiple potential polyadenylation signal sequences (26) and numerous ATTAA sequences (10). Because of its overall similarity to the poorly expressed wild-type *B.t.k.* HD-1 and HD-73 genes, the same problems are expected in expression of the wild-type P2 gene as were encountered with the previous examples. Based on the above-described method for designing the synthetic *B.t.* genes, a synthetic P2 gene has been designed which gene should be expressed at adequate levels for protection in plants. A comparison of the wild-type and synthetic P2 genes is shown in Figure 13.

10 Example 7 -- Synthetic *B.t.* *Entomocidus* Gene

[0126] The *B.t. entomocidus* ("Btent") protein is a distinct insecticidal protein produced by some strains of *B.t.* bacteria. It is characterized by its high level of activity against some lepidopterans that are relatively insensitive to *B.t.k.* HD-1 and HD-73 such as *Spodoptera* species including beet armyworm (Visser et al., 1988). Genes encoding the Btent protein have been isolated and characterized (Honee et al., 1988). The Btent proteins encoded by these genes are approximately the same length as *B.t.k.* HD-1 and HD-73. These proteins share only 68% amino acid homology with the *B.t.k.* HD-1 and HD-73 proteins. It is likely that only the N-terminal half of the Btent protein is required for insecticidal activity as is the case for HD-1 and HD-73. Over the first 625 amino acids, Btent shares only 38% amino acid homology with HD-1 and HD-73.

[0127] Because of their higher activity against *Spodoptera* species that are relatively insensitive to HD-1 and HD-73, the Btent proteins are a desirable candidate for the production of insect tolerant transgenic plants either alone or in combination with the other *B.t.* toxins described in the above examples. In some plants production of Btent alone might be sufficient to control the agronomically important pests. In other plants, the production of two distinct insect tolerance proteins would provide protection against a wider array of insects. Against those insects where both proteins are active, the combination of the *B.t.k.* HD-1 or HD-73 type protein plus the Btent protein should provide at least additive insecticidal efficacy, and may even provide a synergistic activity. In addition, because of its distinct amino acid sequence, the Btent protein may have a different mode of action than HD-1 or HD-73. Production of two insecticidal proteins in the same plant with different modes of action would minimize the potential for development of insect resistance to *B.t.* proteins in plants. The relative lack of DNA sequence homology with the *B.t.k.* type genes minimizes the potential for recombination between multiple insect tolerance genes in the plant chromosome.

[0128] The genes encoding the Btent protein although distinct in sequence from the *B.t.k.* HD-1 and HD-73 genes share many common features with these genes. In particular, the Btent protein genes have a high A+T content (62%), multiple potential polyadenylation signal sequences (39 in the full length coding sequence and 27 in the first 1875 nucleotides that is likely to encode the active toxic fragment) and numerous ATTAA sequences (16 in the full length coding sequence and 12 in the first 1875 nucleotides). Because of its overall similarity to the poorly expressed wild-type *B.t.k.* HD-1 and HD-73 genes, the wild-type Btent genes are expected to exhibit similar problems in expression as were encountered with the wild-type HD-1 and HD-73 genes. Based on the above-described method used for designing the other synthetic *B.t.* genes, a synthetic Btent gene has been designed which gene should be expressed at adequate levels for protection in plants. A comparison of the wild type and synthetic Btent genes is shown in Figure 14.

40 Example 8 -- Synthetic *B.t.k.* Genes for Expression in Corn

[0129] High level expression of heterologous genes in corn cells has been shown to be enhanced by the presence of a corn gene intron (Callis et al., 1987). Typically these introns have been located in the 5' untranslated region of the chimeric gene. It has been shown that the CaMV35S promoter and the NOS 3' end function efficiently in the expression of heterologous genes in corn cells (Fromm et al., 1986).

[0130] Referring to Figure 15, a plant expression cassette vector (pMON744) was constructed that contains these sequences. Specifically the expression cassette contains the enhanced CaMV 35S promoter followed by intron 1 of the corn Adhl gene (Callis et al., 1987). This is followed by a multilinker cloning site for insertion of coding sequences; this multilinker contains a BgIII site among others. Following the multilinker is the NOS 3' end. pMON744 also contains the selectable marker gene 3SS/NPTII/NOS 3' for kanamycin selection of transgenic corn cells. In addition, pMON744 has an *E. coli* origin of replication and an ampicillin resistance gene for selection of the plasmid in *E. coli*.

[0131] Five *B.t.k.* coding sequences described in the previous examples were inserted into the BgIII site of pMON744 for corn cell expression of *B.t.k.* The coding sequences inserted and resulting vectors were:

1. Wild type *B.t.k.* HD-1 from pMON9921 to make pMON8652.
2. Modified *B.t.k.* HD-1 from pMON5370 to make pMON8642.

3. Synthetic *B.t.k* HD-1 from pMON5377 to make pMON8643.
4. Synthetic *B.t.k* HD-73 from pMON5390 to make pMON8644.
5. Synthetic full length *B.t.k* HD-73 from pMON10518 to make pMON10902.

5 [0132] pMON8652 (wild-type *B.t.k* HD-1) was used to transform corn cell protoplasts and stably transformed kanamycin resistant callus was isolated. *B.t.k* mRNA in the corn cells was analyzed by nuclease S1 protection and found to be present at a level comparable to that seen with the same wild-type coding sequence (pMON9921) in transgenic tomato plants.

10 [0133] pMON8652 and pMON8642 (modified HD-1) were used to transform corn cell protoplasts in a transient expression system. The level of *B.t.k* mRNA was analyzed by nuclease S1 protection. The modified HD-1 gave rise to a several fold increase in *B.t.k* mRNA compared to the wild-type coding sequence in the transiently transformed corn cells. This indicated that the modifications introduced into the *B.t.k* HD-1 gene are capable of enhancing *B.t.k* expression in monocot cells as was demonstrated for dicot plants and cells.

15 [0134] pMON8642 (modified HD-1) and pMON8643 (synthetic HD-1) were used to transform Black Mexican Sweet (BMS) corn cell protoplasts by PEG-mediated DNA uptake, and stably transformed corn callus was selected by growth on kanamycin containing plant growth medium. Individual callus colonies that were derived from single transformed cells were isolated and propagated separately on kanamycin containing medium.

20 [0135] To assess the expression of the *B.t.k* genes in these cells, callus samples were tested for insect toxicity by bioassay against tobacco hornworm larvae. For each vector, 96 callus lines were tested by bioassay. Portions of each callus were placed on sterile water agar plates, and five neonate tobacco hornworm larvae were added and allowed to feed for 4 days. For pMON8643, 100% of the larvae died after feeding on 15 of the 96 calli and these calli showed little feeding damage. For pMON8642, only 1 of the 96 calli was toxic to the larvae. This showed that the *B.t.k* gene was being expressed in these samples at insecticidal levels. The observation that significantly more calli containing pMON8643 were toxic than for pMON8642 showed that significantly higher levels of expression were obtained when the synthetic HD-1 coding sequence was contained in corn cells than when the modified HD-1 coding sequence was used, similar to the previous examples with dicot plants. A semiquantitative immunoassay showed that the pMON8643 toxic samples had significantly higher *B.t.k* protein levels than the pMON8642 toxic sample.

25 [0136] The 16 callus samples that were toxic to tobacco hornworm were also tested for activity against European corn borer. European corn borer is approximately 40-fold less sensitive to the HD-1 gene product than is tobacco hornworm. Larvae of European corn borer were applied to the callus samples and allowed to feed for 4 days. Two of the 16 calli tested, both of which contained pMON8643 (synthetic HD-1), were toxic to European corn borer larvae.

30 [0137] To assess the expression of the *B.t.k* genes in differentiated corn tissue, another method of DNA delivery was used. Young leaves were excised from corn plants, and DNA samples were delivered into the leaf tissue by microprojectile bombardment. In this system, the DNA on the microprojectiles is transiently expressed in the leaf cells after bombardment. Three DNA samples were used, and each DNA was tested in triplicate.

1. pMON744, the corn expression vector with no *B.t.k* gene.
2. pMON8643 (synthetic HD-1).
3. pMON752, a corn expression vector for the GUS gene, no *B.t.k* gene.

40 [0138] The leaves were incubated at room temperature for 24 hours. The pMON752 samples were stained with a substrate that allows visual detection of the GUS gene product. This analysis showed that over one hundred spots in each sample were expressing the GUS product and the triplicate samples showed very similar levels of GUS expression. For the pMON744 and pMON8643 samples 5 larvae of tobacco hornworm were added to each leaf and allowed to feed for 48 hours. All three samples bombarded with pMON744 showed extensive feeding damage and no larval mortality. All three samples bombarded with pMON8643 showed no evidence of feeding damage and 100% larval mortality. The samples were also assayed for the presence of *B.t.k* protein by a qualitative immunoassay. All of the pMON8643 samples had detectable *B.t.k* protein. These results demonstrated that the synthetic *B.t.k* gene was expressed in differentiated corn plant tissue at insecticidal levels.

50 Example 9 -- Expression of Synthetic *B.t* Genes with RUBISCO Small Subunit Promoters and Chloroplast Transit Peptides

55 [0139] The genes in plants encoding the small subunit of RUBISCO (SSU) are often highly expressed, light regulated and sometimes show tissue specificity. These expression properties are largely due to the promoter sequences of these genes. It has been possible to use SSU promoters to express heterologous genes in transformed plants. Typically a plant will contain multiple SSU genes, and the expression levels and tissue specificity of different SSU genes will be different. The SSU proteins are encoded in the nucleus and synthesized in the cytoplasm as precursors that contain

an N-terminal extension known as the chloroplast transit peptide (CTP). The CTP directs the precursor to the chloroplast and promotes the uptake of the SSU protein into the chloroplast. In this process, the CTP is cleaved from the SSU protein. These CTP sequences have been used to direct heterologous proteins into chloroplasts of transformed plants.

[0140] The SSU promoters might have several advantages for expression of *B.t.k* genes in plants. Some SSU promoters are very highly expressed and could give rise to expression levels as high or higher than those observed with the CaMV35S promoter. The tissue distribution of expression from SSU promoters is different from that of the CaMV35S promoter, so for control of some insect pests, it may be advantageous to direct the expression of *B.t.k* to those cells in which SSU is most highly expressed. For example, although relatively constitutive, in the leaf the CaMV35S promoter is more highly expressed in vascular tissue than in some other parts of the leaf, while most SSU promoters are most highly expressed in the mesophyll cells of the leaf. Some SSU promoters also are more highly tissue specific, so it could be possible to utilize a specific SSU promoter to express *B.t.k* in only a subset of plant tissues. If for example *B.t* expression in certain cells was found to be deleterious to those cells. For example, for control of Colorado potato beetle in potato, it may be advantageous to use SSU promoters to direct *B.t.k* expression to the leaves but not to the edible tubers.

[0141] Utilizing SSU CTP sequences to localize *B.t* proteins to the chloroplast might also be advantageous. Localization of the *B.t* to the chloroplast could protect the protein from proteases found in the cytoplasm. This could stabilize the *B.t* protein and lead to higher levels of accumulation of active protein. *B.t* genes containing the CTP could be used in combination with the SSU promoter or with other promoters such as CaMV35S.

[0142] A variety of plant transformation vectors were constructed for the expression of *B.t.k* genes utilizing SSU promoters and SSU CTPs. The promoters and CTPs utilized were from the petunia SSU11a gene described by Turner et al. (1986) and from the *Arabidopsis* ats1A gene (an SSU gene) described by Krebbers et al. (1988) and by Ellonor et al. (1989). The petunia SSU11a promoter was contained on a DNA fragment that extended approximately 800 bp upstream of the SSU coding sequence. The *Arabidopsis* ats1A promoter was contained on a DNA fragment that extended approximately 1.8 kb upstream of the SSU coding sequence. At the upstream end convenient sites from the multicloner of pUC18 were used to move these promoters into plant transformation vectors such as pMON893. These promoter fragments extended to the start of the SSU coding sequence at which point an NcoI restriction site was engineered to allow insertion of the *B.t* coding sequence, replacing the SSU coding sequence.

[0143] When SSU promoters were used in combination with their CTP, the DNA fragments extended through the coding sequence of the CTP and a small portion of the mature SSU coding sequence at which point an NcoI restriction site was engineered by standard techniques to allow the in frame fusion of *B.t* coding sequences with the CTP. In particular, for the petunia SSU11a CTP, *B.t* coding sequences were fused to the SSU sequence after amino acid 8 of the mature SSU sequence at which point the NcoI site was placed. The 8 amino acids of mature SSU sequence were included because preliminary *in vitro* chloroplast uptake experiments indicated that uptake was of *B.t.k* was observed only if this segment of mature SSU was included. For the *Arabidopsis* ats1A CTP, the complete CTP was included plus 24 amino acids of mature SSU sequence plus the sequence gly-gly-arg-val-asn-cys-met-gln-alu-met, terminating in an NcoI site for *B.t* fusion. This short sequence reiterates the native SSU CTP cleavage site (between the cys and met) plus a short segment surrounding the cleavage site. This sequence was included in order to insure proper uptake into chloroplasts. *B.t* coding sequences were fused to this ats1A CTP after the met codon. *In vitro* uptake experiments with this CTP construction and other (non-*B.t*) coding sequences showed that this CTP did target proteins to the chloroplast.

[0144] When CTPs were used in combination with the CaMV 35S promoter, the same CTP segments were used. They were excised just upstream of the ATG start sites of the CTP by engineering of BglII sites, and placed downstream of the CaMV35S promoter in pMON893, as BglII to NcoI fragments. *B.t* coding sequences were fused as described above.

[0145] The wild type *B.t.k* HD-1 coding sequence of pMON9921 (see Figure 1) was fused to the ats1A promoter to make pMON1925 or the ats1A promoter plus CTP to make pMON1921. These vectors were used to transform tobacco plants, and the plants were screened for activity against tobacco hornworm. No toxic plants were recovered. This is surprising in light of the fact that toxic plants could be recovered, albeit at a low frequency, after transformation with pMON9921 in which the *B.t.k* coding sequence was expressed from the enhanced CaMV35S, promoter in pMON893, and in light of the fact that Ellonor et al. (1989) report that the ats1A promoter itself is comparable in strength to the CaMV35S promoter and approximately 10-fold stronger when the CTP sequence is included. At least for the wild-type *B.t.k* HD-1 coding sequence, this does not appear to be the case.

[0146] A variety of plant transformation vectors were constructed utilizing either the truncated synthetic HD-73 coding sequence of Figure 4 or the full length *B.t.k* HD-73 coding sequence of Figure 11. These are listed in the table below.

Table XV

Gene Constructs with CTPs			
Vector	Promoter	CTP	B.t.k. HD-73 Coding Sequence
pMON10806	En 35S	ats1A	truncated
pMON10814	En35S	SSU11a	full length
pMON10811	SSU11a	SSU11a	truncated
pMON10819	SSU11a	none	truncated
pMON10815	ats1A	none	truncated
pMON10817	ats1A	ats1A	truncated
pMON10821	En 35S	ats1A	truncated
pMON10822	En 35S	ats1A	full length
pMON10838	SSU11a	SSU11a	full length
pMON10839	ats1A	ats1A	full length

[0147] All of the above vectors were used to transform tobacco plants. For all of the vectors containing truncated B.t.k. genes, leaf tissue from these plants has been analyzed for toxicity to insects and B.t.k. protein levels by immunoassay. pMON10806, 10811, 10819 and 10821 produce levels of B.t.k. protein comparable to pMON5383 and pMON5390 which contain synthetic B.t.k. HD-73 coding sequences driven by the En 35S promoter itself with no CTP. These plants also have the insecticidal activity expected for the B.t.k. protein levels detected. For pMON10815 and pMON10817 (containing the ats1A promoter), the level of B.t.k. protein is about 5-fold higher than that found in plants containing pMON5383 or 5390. These plants also have higher insecticidal activity. Plants containing 10815 and 10817 contain up to 1% of their total soluble leaf protein as B.t.k. HD-73. This is the highest level of B.t.k. protein yet obtained with any of the synthetic genes.

[0148] This result is surprising in two respects. First, as noted above, the wild type coding sequences fused to the ats1A promoter and CTP did not show any evidence of higher levels of expression than for En 35S, and in fact had lower expression based on the absence of any insecticidal plants. Second, Ellonor et al. (1989) show that for two other genes, the ats1A CTP can increase expression from the ats1A promoter by about 10-fold. For the synthetic B.t.k. HD-73 gene, there is no consistent increase seen by including the CTP over and above that seen for the ats1A promoter alone.

[0149] Tobacco plants containing the full length synthetic HD-73 fused to the SSU11A CTP and driven by the En 35S promoter produced levels of B.t.k. protein and insecticidal activity comparable to pMON1518 which contains does not include the CTP. In addition, for pMON10518 the B.t.k. protein extracted from plants was observed by gel electrophoresis to contain multiple forms less than full length, apparently due the cleavage of the C-terminal portion (not required for toxicity) in the cytoplasm. For pMON10814, the majority of the protein appeared to be intact full length. Indicating that the protein has been stabilized from proteolysis by targeting to the chloroplast.

#### Example 10 -- Targeting of B.t. Proteins to the Extracellular Space or Vacuole through the Use of Signal Peptides

[0150] The B.t. proteins produced from the synthetic genes described here are localized to the cytoplasm of the plant cell, and this cytoplasmic localization results in plants that are insecticidally effective. It may be advantageous for some purposes to direct the B.t. proteins to other compartments of the plant cell. Localizing B.t. proteins in compartments other than the cytoplasm may result in less exposure of the B.t. proteins to cytoplasmic proteases leading to greater accumulation of the protein yielding enhanced insecticidal activity. Extracellular localization could lead to more efficient exposure of certain insects to the B.t. proteins leading to greater efficacy. If a B.t. protein were found to be deleterious to plant cell function, then localization to a noncytoplasmic compartment could protect these cells from the protein.

[0151] In plants as well as other eucaryotes, proteins that are destined to be localized either extracellularly or in several specific compartments are typically synthesized with an N-terminal amino acid extension known as the signal peptide. This signal peptide directs the protein to enter the compartmentalization pathway, and it is typically cleaved from the mature protein as an early step in compartmentalization. For an extracellular protein, the secretory pathway typically involves cotranslational insertion into the endoplasmic reticulum with cleavage of the signal peptide occurring at this stage. The mature protein then passes thru the Golgi body into vesicles that fuse with the plasma membrane thus releasing the protein into the extracellular space. Proteins destined for other compartments follow a similar pathway. For example, proteins that are destined for the endoplasmic reticulum or the Golgi body follow this scheme, but they are specifically retained in the appropriate compartment. In plants, some proteins are also targeted to the vacuole,

another membrane bound compartment in the cytoplasm of many plant cells. Vacuole targeted proteins diverge from the above pathway at the Golgi body where they enter vesicles that fuse with the vacuole.

[0152] A common feature of this protein targeting is the signal peptide that initiates the compartmentalization process. Fusing a signal peptide to a protein will in many cases lead to the targeting of that protein to the endoplasmic reticulum.

5 The efficiency of this step may depend on the sequence of the mature protein itself as well. The signals that direct a protein to a specific compartment rather than to the extracellular space are not as clearly defined. It appears that many of the signals that direct the protein to specific compartments are contained within the amino acid sequence of the mature protein. This has been shown for some vacuole targeted proteins, but it is not yet possible to define these sequences precisely. It appears that secretion into the extracellular space is the "default" pathway for a protein that 10 contains a signal sequence but no other compartmentalization signals. Thus, a strategy to direct *B.t.* proteins out of the cytoplasm is to fuse the genes for synthetic *B.t.* genes to DNA sequences encoding known plant signal peptides. These fusion genes will give rise to *B.t.* proteins that enter the secretory pathway, and lead to extracellular secretion or targeting to the vacuole or other compartments.

[0153] Signal sequences for several plant genes have been described. One such sequence is for the tobacco pathogenesis related protein PR1b described by Cornelissen et al. The PR1b protein is normally localized to the extracellular space. Another type of signal peptide is contained on seed storage proteins of legumes. These proteins are localized to the protein body of seeds, which is a vacuole like compartment found in seeds. A signal peptide DNA sequence for the beta subunit of the 7S storage protein of common bean (*Phaseolus vulgaris*), PvB, has been described by Doyle et al. Based on the published these published sequences, genes were synthesized by chemical synthesis of oligonucleotides that encoded the signal peptides for PR1b and PvB. The synthetic genes for these signal peptides corresponded exactly to the reported DNA sequences. Just upstream of the translational initiation codon of each signal peptide a BamHI and BgIII site were inserted with the BamHI site at the 5' end. This allowed the insertion of the signal peptide encoding segments into the BgIII site of pMON893 for expression from the En 35S promoter. In some cases to achieve secretion or compartmentalization of heterologous proteins, it has proved necessary to include some amino acid sequence beyond the normal cleavage site of the signal peptide. This may be necessary to insure proper cleavage of the signal peptide. For PR1b the synthetic DNA sequence also included the first 10 amino acids of mature PR1b. For PvB the synthetic DNA sequence included the first 13 amino acids of mature PvB. Both synthetic signal peptide encoding segments ended with NcoI sites to allow fusion in frame to the methionine initiation codon of the synthetic *B.t.* genes.

30 [0154] Four vectors encoding synthetic *B.t.k.* HD-73 genes were constructed containing these signal peptides. The synthetic truncated HD-73 gene from pMON5383 was fused with the signal peptide sequence of PvB and incorporated into pMON893 to create pMON10827. The synthetic truncated HD-73 gene from pMON5383 was also fused with the signal peptide sequence of PR1b to create pMON10824. The full length synthetic HD-73 gene from pMON10518 was fused with the signal peptide sequence of PvB and incorporated into pMON893 to create pMON10828. The full length 35 synthetic HD-73 gene from pMON10518 was also fused with the signal peptide sequence of PR1b and incorporated into pMON893 to create pMON10825.

[0155] These vectors were used to transform tobacco plants and the plants were assayed for expression of the *B.t.k.* protein by Western blot analysis and for insecticidal efficacy. pMON10824 and pMON10827 produced amounts of *B.t.k.* protein in leaf comparable to the truncated HD-73 vectors, pMON5383 and pMON5390. pMON10825 and 40 pMON10828 produced full length *B.t.k.* protein in amounts comparable to pMON10518. In all cases, the plants were insecticidally active against tobacco hornworm.

#### BIBLIOGRAPHY

45 [0156]

Adami, G. and Nevins, J. (1988) RNA Processing, Cold Spring Harbor Laboratory, p. 28.

Adang, et al., Molecular Strategies for Crop Protection (1987) pp. 345-353, Alan R. Liss, Inc.

50

Barton, K. A. et al., Plant Physiol. (1987), 85, 1103-1109.

55

Bevan, M. et al., Nature (1983) 304:184.

Brady, H. and Wold, W. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 224.

Brown, John W., Nucleic Acids Research (1988) Vol. 14, No. 24, p. 9549.

5 Calls, J. Fromm, M. and Walbot, V., Genes and Develop. (1987), 1:1183-1200.

10 Conway, L. and Wickens, M. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 40.

15 Cornelissen, B.J.C., et al., EMBO J. (1988) Vol. 5, No. 1, 37-40.

20 Daar, I. O. et al. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 45.

25 Dean, C. et al., Nucleic Acids Research (1986), Vol. 14, No. 5, p. 2229.

30 Dedrick, R., et al., The Journal of Biological Chemistry (1987), Vol. 262, No. 19, pp. 9098-1108.

35 Donovan, W. P. et al., The J. of Biol. Chem. (1988), Vol. 263, No. 1, pp. 561-567.

40 Doyle, J.J., et al., J. Biol. Chem. (1986), Vol. 261, No. 20, 9228-9236.

45 Ellenor, R.P., et al., Mol. Gen. Genet. (1989), 218:78-86.

50 Fischhoff, D. A. et al., Bio/Technology (1987), Vol. 5, p. 807.

55 Fraley, R. T. et al., Bio/Technology (1985) 3:629-635.

60 Fromm, M., Taylor, L. P. and Walbot, V., Nature (1986), 319:791-793.

65 Gallego, M. E. and Nadal-Ginard B. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 61.

70 Genovese, C. and Milcarek, C. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 62.

75 Gil, A. and Proudfoot, N. J., Nature (1984), Vol. 312, p. 473.

80 Goodall, G. et al. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 63.

85 Gross, et al. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 128.

90 Hampson, R. K. and Rottman, F. M. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 68.

95 Hanley, Brian A and Schuler, Mary A., Nucleic Acids Research (1988), Vol. 16, No. 14, p. 7159.

100 Helfman, D. M. and Ricci, W. M. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 219.

105 Herrera-Estrella, L. et al., Nature (1983), 303:209.

110 Hoekema, A. et al., Molecular and Cellular Biology (1987), Vol. 7, pp. 2914-2924.

115 Honee, G. et al., Nucleic Acids Research (1988), Vol. 16, No. 13.

120 Horsch, R. B. et al., Science (1985), 227:1229.

125 Jarret, R. L. et al., Physiol. Plant (1980), 49:177.

130 Jarret, R. L. et al., In Vitro (1981), 17:825.

135 Kay, R. et al., Science (1987), 236:1299-1302.

140 Kessler, M. et al. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 85.

145 Klee, H. J. et al., Bio/Technology (1985), 3:837-842.

5 Kozak, M., Nature (1984), 308:241-248.

10 Krebbers, E., et al., Plant Molecular Biology (1988), 11:745-759.

15 Kunkel, T. A., Proc. Natl. Acad. Sci. USA (1985), Vol. 82, pp. 488-492.

20 Marrone et al., J. Econ. Entomol. (1985), 78:290-293.

25 Marzluff, W. and Pandey, N. (1988), RNA Processing, Cold Spring Harbor Laboratory, p. 244.

30 McCormick, S. et al., Plant Cell Reports (1988), 5:81-84.

35 McDevitt, M. A. et al., Cell (1984), Vol. 37, pp. 993-999.

40 Murashige, T. and Skoog, F., Physiol. Plant (1962), 15:473.

45 Odell, J. et al., Nature (1985), 313:810.

50 Pandey, N. B. and Marzluff, W. F. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 133.

55 Proudfoot, N. J. et al. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 17.

60 Reines, D., et al., J. Mol. Biol. (1987) 196:299-312.

65 Sadofsky, M. and Alwine, J. C., Molecular and Cellular Biology (1984), Vol. 4, No. 8, pp. 1460-1468.

70 Sanders, P. R. et al., Nucleic Acids Research (1987), Vol. 15, No. 4, p. 1543.

75 Schuler, M. A. et al., Nucleic Acids Research (1982), Vol. 10, No. 24, pp. 8225-8244.

80 Shaw, G. & Kamen, R., Cell (1986), 46:659-667.

85 Shaw, G. and Kamen, R. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 220.

90 Trollinder, N. L. and Goodin, J. R., Plant Cell Reports (1987), 6:231-244.

95 Tsurushita, N. and Korn, L. J. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 215.

100 Turner, N.E., et al., Nucleic Acids Reg. (1986), Vol. 14:8, 3325.

105 Vaeck, M. et al., Nature (1987), Vol. 328, p. 33.

110 Velten et al., EMBO J. (1984), 3:2723-2730.

115 Velten & Schell, Nucleic Acids Research (1985), 13:6981-6998.

120 Visser, B. et al., Mol. Gen. Genet. (1988), 212:219-224.

125 Webb, K. J. et al., Plant Sci. Letters (1983), 30:1.

130 Wickens, M. and Stephenson, P., Science (1984), Vol. 226, p. 1045.

135 Wickens, M. et al. (1987), RNA Processing, Cold Spring Harbor Laboratory, p. 9.

140 Wlebauer, K. et al., Molecular and Cellular Biology (1988), Vol. 8, No. 5, pp. 2042-2051.

145 Yamamoto, T. and Iizuka, T., Archives of Biochemistry and Biophysics (1983), Vol. 227, No. 1, pp. 233-241.

## Claims

1. A method for modifying a wild-type structural gene sequence which encodes an insecticidal protein of *Bacillus thuringiensis* to enhance the expression of said protein in plants which comprises:
  - a) identifying regions within said sequence with greater than four consecutive adenine or thymine nucleotides;
  - b) modifying the regions of step (a) which have two or more polyadenylation signals within a ten base sequence to remove said signals while maintaining a gene sequence which encodes said protein; and
  - c) modifying the 15-30 base regions surrounding the regions of step (a) to remove major plant polyadenylation signals, consecutive sequences containing more than one minor polyadenylation signal and consecutive sequences containing more than one ATTAA sequence while maintaining a gene sequence which encodes said protein.
2. A method for modifying a wild-type structural gene sequence which encodes an insecticidal protein of *Bacillus thuringiensis* to enhance the expression of said protein in plants which comprises:
  - a) removing polyadenylation signals contained in said wild-type gene while retaining a sequence which encodes said protein; and
  - b) removing ATTAA sequences contained in said wild-type gene while retaining a sequence which encodes said protein.
3. A method of claim 2 further comprising the removal of self-complementary sequences and replacement of such sequences with nonself-complementary DNA comprising plant preferred codons while retaining a structural gene sequence encoding said protein.
4. A method of claims 1 to 3 further comprising the use of plant preferred sequences in the removal of the polyadenylation signals and ATTAA sequences.
5. A method of claims 1 to 3 in which the plant polyadenylation signals are selected from the group consisting of AAAAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAAA, ATGAAA, AAGCAT, ATTAAT, ATACAT, AAAATA, ATTTAA, AATTAA, AATACA and CATAAA.
6. A method for improving the expression of a heterologous gene in plants wherein said gene comprises a modified chimeric gene containing a promoter which functions in plant cells operably linked to a structural coding sequence and a 3' non-translated region containing a polyadenylation signal which functions in plants to cause the addition of polyadenylate nucleotides to the 3' end of the RNA, wherein said structural coding sequence encodes an insecticidal protein at least a portion of which was derived from a *Bacillus thuringiensis* protein, wherein said method comprises modifying said structural coding sequence so that said sequence has a DNA sequence which differs from the naturally occurring DNA sequence encoding said *Bacillus thuringiensis* protein and said structural coding sequence does not contain more than 5 consecutive nucleotides consisting of either adenine or thymine residues.
7. A method for improving the expression of a heterologous gene in plants wherein said gene comprises a modified chimeric gene containing a promoter which functions in plant cells operably linked to a structural coding sequence and a 3' non-translated region containing a polyadenylation signal which functions in plants to cause the addition of polyadenylate nucleotides to the 3' end of the RNA, wherein said structural coding sequence encodes an insecticidal protein at least a portion of which was derived from a *Bacillus thuringiensis* protein, wherein said method comprises modifying said structural coding sequence so that said sequence has a DNA sequence which differs from the naturally occurring DNA sequence encoding said *Bacillus thuringiensis* protein and has the following characteristics:

said structural coding sequence has a region which is complementary to the following sequence:

GGCTTGATTCTAGCGAACTCTCGATTCTCTGGTTGATGAGCTGGTC  
1 5 10 15 20 25 30 35 40 45

5 said region in said coding sequence having eliminated 2 AACCAA and 1 AATTAA sequence.

8. A method according to claim 7, wherein said structural coding sequence encodes an insecticidal protein at least a portion of which was derived from a *Bacillus thuringiensis kurstakls* HD-1.

10 9. A method according to claim 7 or 8, wherein the plant is a tobacco plant.

15 10. A modified chimeric gene containing a promoter which functions in plant cells operably linked to a structural coding sequence and a 3' non-translated region containing a polyadenylation signal which functions in plants to cause the addition of polyadenylate nucleotides to the 3' end of the RNA, wherein said structural coding sequence encodes an insecticidal protein at least a portion of which was derived from a *Bacillus thuringiensis* protein, wherein said structural coding sequence has a DNA sequence which differs from the naturally occurring DNA sequence encoding said *Bacillus thuringiensis* protein and is selected from:

20 A. A structural gene which encodes an insecticidal protein of *B.t.k.* HD-1 having the sequence:

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1	ATGGCTATAGAACTGGTTACACCCCAATCGATATTCCT	40
5		
41	TGTCGCTAACGCAATTCTTTGAGTGAATTGTTCCGG	80
10		
81	TGCTGGATTGTGTTAGGACTAGTTGATATTATCTGGGA	120
15		
121	ATTTTGGTCCCTCTCAATGGGACGCATTCTGTACAAA	160
20		
161	TTGAACAGCTCATCAACCAGAGAAATCGAAGAGTCGCTAG	200
25		
201	GAATCAAGCCATTCTAGATTAGAAGGACTAACGCAATCTT	240
30		
241	TATCAAATTACGCAGAACCTTTAGAGAGTGGGAGGCAG	280
35		
281	ATCCTACTAATCCAGCATTAAAGAGAAAGAGATGCGTATTCA	320
40		
321	ATTCAATGACATGAAACAGTGCCCTTACAACCGCTATTCTT	360
45		
361	CTTTTGCAAGTCAAAATTATCAAGTTCTCTCCCTCCG	400
50		
401	TGTACGTTCAAGCTGCCAACCTCACCTCTCAGTTTGAG	440
55		
441	AGATGTTCAAGTGTGTTGGACAAAGGTGGGGATTGATGCC	480
481	GCGACTATCAATAGTCGTTATAATGATTTAACTAGGCTTA	520

5	521	TTGGCAACTATACAGATCATGCTGTACGCTGGTACAATAC	560
10	561	GGGATTAGAGCGTGTATGGGACCGGATTCTAGAGATTGG	600
15	601	ATCAGGTACAACCAACAGTTCAGAAGAGAGCTTACACTAATG	640
20	641	TATTAGATATCGTTCTCTATTTCCGAACTATGATAGTAG	680
25	681	AACGTATCCAATTCGAACAGTTCCCAATTAAACAGAGAA	720
30	721	ATTTATACAAACCCAGTATTAGAAAATTTGATGGTAGTT	760
35	761	TTCGAGGCTCGGCTCAGGGCATAGAAGGAAGTATTAGGAG	800
40	801	TCCACATTTGATGGATATACTTAATAGTATAACCATCTAT	840
45	841	ACCGATGCTCATAGAGGAGAACTACTACTGGTCCGGTCACC	880
50	881	AGATCATGGCTTCTCCTGTAGGGTTTCGGGCCAGAATT	920
55	921	CACTTTCCGTATATGAACTATGGAAATGCAGCTCCA	960
60	961	CAACAACGTATTGTTGCTCAACTAGGTCAAGGGCGTGTATA	1000
65	1001	GAACATTATCGTCCACCTTATATAGAAGACCTTTAACAT	1040
70	1041	CGGGATCAACAACCAACAACTATCTGTTCTGACGGGACA	1080
75	1081	GAATTGCTTATGAAACCTCCTCAAATTGCCATCCGCTG	1120

5	1121	TATAACAGAAAAAGCGGAACGGTAGATTGCTGGATGAAAT	1160
10	1161	ACCGCCACAGAATAACAACGTGCCACCTAGGCAAGGATT	1200
15	1201	AGTCATCGATTAAGCCATGTTCAATGTTCGTTAGGCT	1240
20	1241	TTAGTAATAGTAGTGTAAGTATAATAAGAGCTCCTATGTT	1280
25	1281	CTCTGGATACATCGTAGTGCTGAGTTCAACAAACATCATC	1320
30	1321	CCTTCATCACAAATCACCCAAATCCCACTCACCAAGTCTA	1360
35	1361	CTAATCTTGGCTCTGGAACCTCTGTCGTTAAAGGACCAAGG	1400
40	1401	ATTTACAGGAGGAGATATTCTCGAAGAACCTCACCTGGC	1440
45	1441	CAGATTCAACCTTAAGAGTAAATATTACTGCACCATTAT	1480
50	1481	CACAAAGATATCGGGTAAGAATTGCTACGCTTACCCAC	1520
55	1521	AAACCTTCAGTTCCACACATCAATTGACGGAAGACCTATT	1560
60	1561	AATCAGGGAAATTTTCAGCAACTATGAGTAGTGGAGTA	1600
65	1601	ATTTACAGTCCGGAAGCTTACGACTGTAGGTTTACTAC	1640
70	1641	TCCGTTAACCTTCAAATGGATCAAGTGTATTCAGTTA	1680
75	1681	AGTGCTCATGTCCTCAATTCAAGGCAATGAAGTTATAG	1720
80	1721	ATCGAATTGAATTGTTCCGGCA	1743

B. A structural gene which encodes an insecticidal protein of *B.t.k.* HD-73 having the sequence:

1	ATGGCCATTGAAACCGGTTACACTCCCATCGACATCTCCT	40
5		
41	TGTCCCTGACACAGTTCTGCTCAGCGAGTTGGTGCAGG	80
10		
81	TGCTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGT	120
15		
121	ATCTTGGTCCATCTCAATGGGATGCATTCCCTGGTGCAAA	160
20		
161	TTGAGCAGTTGATCAACCAGAGGATCGAAGAGTTGCCAG	200
25		
201	GAACCAAGGCCATCTCTAGGTTGGAGGGATTGAGCAATCTC	240
30		
241	TACCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCG	280
35		
281	ATCCTACTAACCCAGCTCTCCGCAGGGAAATGCGTATTCA	320
40		
321	ATTCAACGACATGACAGCGCCTTGACCACAGCTATCCCA	360
45		
361	TTGTTCCAGTCCAGAACTACCAAGTTCCCTCTTGTCCG	400
50		
401	TGTACGTTCAAGCAGCTAATCTCACCTCAGCGTGCTTCG	440
55		

441	AGACGTTAGCGTGTGGGCAAAGGTGGGATTGATGCT	480
481	GCAACCATCAATAGCGTTACAACGACCTTACTAGGCTGA	520
521	TTGGAAACTACACCGACCACGCTGTTGTTACAACAC	560
561	TGGCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGG	600
601	ATTAGATACAACCAGTTCAAGGAGAGAATTGACCCCTACAG	640
641	TTTGGACATTGTGTCTCTTCCCGAACTATGACTCCAG	680
681	AACCTACCCCTATCCGTACAGTGTCCCACTTACCAAGAGAA	720
721	ATCTATACTAACCCAGTTCTTGAGAACCTCGACGGTAGCT	760
761	TCCGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAG	800
801	CCCACACTTGATGGACATCTGAACAGCATAACTATCTAC	840
841	ACCGATGCTCACAGAGGGAGAGTATTACTGGTCTGGACACC	880
881	AGATCATGGCCTCTCCAGTTGGATTCAAGCGGGCCCGAGTT	920
921	TACCTTCCCTCTATGGAACATATGGAAACGCCGCTCCA	960
961	CAACAAACGTATCGTTGCTCAACTAGGTCAGGGTGTCTACA	1000
1001	GAACCTTGTCTTCCACCTTGTACAGAAGACCCCTCAATAT	1040

5	1041	CGGTATCAACACCAGCACTTCCGTTCTGACGGAACA	1080
10	1081	GAGTCGCCTATGGAACCTCTTAACCTGCCATCCGCTG	1120
15	1121	TTTACAGAAAGAGCGGAACCGTTGATTCCCTGGACGAAAT	1160
20	1161	CCCACCCACAGAACAAACAATGTGCCACCCAGGCAAGGATTC	1200
25	1201	TCCCCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGAT	1240
30	1241	TCAGCAACAGTTCCGTGAGCATCATCAGAGCTCTATGTT	1280
35	1281	CTCTTGGATAACACCGTAGTGCTGAGTTCAACAAACATCATC	1320
40	1321	GCATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAA	1360
45	1361	ACTTCTCTCAACGGTTCTGTCAATTCAAGGACCAGGATT	1400
50	1401	CACTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAAT	1440
55	1441	AACATTCAAGATAGAGGGTATATTGAAGTTCCAATTCACT	1480
60	1481	TCCCATCCACATCTACCAAGATATAGAGTTCGTGTGAGGTA	1520
65	1521	TGCTTCTGTGACCCCTATTCACCTCAACGTTAATTGGGT	1560
70	1561	AATTCACTCCATCTCTCCAATACAGTTCCAGCTACAGCTA	1600
75	1601	CCTCCTGGATAATCTCAAATCCAGCGATTTCGGTTACTT	1640

5  
1641 TGAAAGTGCCAATGCTTTACATCTTCACTCGGTAACATC 1680  
1681 GTGGGTGTTAGAAACTTACTGGGACTGCAGGAGTGATTA 1720  
10 1721 TCGACAGATTGAGTTCAATTCCAGTTACTGCAACACTCGA 1760  
15 1761 GGCTGAG 1767.

C. A structural gene encoding a insecticidal protein of *B.t.k* HD-1 having the sequence:

20 1 ATGGACAAACAACCCAAACATCAACGAATGCATTCCATACA 40  
25 41 ACTGCTTGACTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80  
81 ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG 120  
30 121 TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCAGGTG 160  
161 CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT 200  
35 201 CTTTGGTCCATCTCAATGGGATGCATTCTGGTGCAAATT 240  
40 241 GAGCAGTTGATCAACCAGAGGGATCGAAGAGTTGCCAGGA 280  
281 ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA 320  
45 321 CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT 360

361	CCTACTAACCCAGCTCTCCGCGAGGAAATCGTATTCAAT	400
401	TCAACCGACATGAACAGCGCTTGACCACAGCTATCCCATT	440
441	GTTCGCAGTCCAGAAGTACCAAGTTCTCTTGTCCGTG	480
481	TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG	520
521	ACGTTAGCGTGTGGCAAGGTTGGGATTGATGCTGC	560
561	AACCACATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600
601	GGAAACTACACCGACCACGCTGTTGTTGACAAACACTG	640
641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680
681	TAGATACAACCAAGTTAGGAGAGATTGACCCCTCACAGTT	720
721	TTGGACATTGTGTCTCTTCCCAGAACTATGACTCCAGAA	760
761	CCTACCCATCCGTACAGTGTCCAACCTTACCAAGAGAAAT	800
801	CTATACTAACCCAGTTCTTGAGAACCTCGACGGTAGCTTC	840
841	CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
881	CACACTTGATGGACATCTTGAACAGCATAACTATCACAC	920
921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG	960

961	ATCATGGCCTCTCCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
1001	CCTTTCTCTATGGAACATGGAAACGCCGCTCCACA	1040
1041	ACAACGTATCGTTGCTCAACTAGGTCAAGGGTGTACAGA	1080
1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCCTCAATATCG	1120
1121	GTATCACAACCAGCAACTTCCGTTCTGACGGAACAGA	1160
1161	GTTCGCCTATGGAACCTCTTCTAATTGCCATCCGCTGTT	1200
1201	TACAGAAAGAGCGGAACCGTTGATTCTTGGACGAAATCC	1240
1241	CACCAAGAACAAATGTGCCACCCAGGCAAGGATTCTC	1280
1281	CCACAGGTTGAGCCACGTTGATGTTCCGTTCCGGATTG	1320
1321	AGCAACAGTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
1361	CATGGATTCACTGTTAGTGTGAGTCAACAATATCATTCC	1400
1401	TTCCCTCTCAAATCACCCAAATCCCATTGACCAAGTCTACT	1440
1441	AACCTTGGATCTGGAACCTCTGTCGTGAAAGGACCCAGGCT	1480
1481	TCACAGGAGGTGATATTCTTAGAAGAACCTCTCCGGCCA	1520
1521	GATTAGCACCCCTCAGAGTTAACATCACTGCACCACTTCT	1560

1561 CAAAGATATCGTGTAGGATTGTTACGCATCTACCACTA 1600  
 1601 ACTTGCAATTCCACACCTCCATCGACGGAAGGCCTATCAA 1640  
 1641 TCAGGGTAACCTCTCGCAACCATGTCAAGCGGCAGAAC 1680  
 1681 TTGCAATCCGGCAGCTTCAGAACCGTCGGTTCACTACTC 1720  
 1721 CTTTCAACTTCTCTAACGGATCAAGCGTTTCACCCCTAG 1760  
 1761 CGCTCATGTGTTCAATTCTGCAATGAAGTGTACATTGAC 1800  
 1801 CGTATTGAGTTGTGCCCTGCCGAAGTTACCTTGAGGCTG 1840  
 1841 AGTAC 1845

D. A structural gene encoding an insecticidal protein derived from *B.t.k* HD-73 having the sequence:

1 ATGGACACAAACCCAAACATCAACGAATGCATTCCATACA 40  
 41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80  
 81 ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG 120  
 121 TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG 160  
 161 CTGGGTTCGTTCTCGGACTACTTGACATCATCTGGGTAT 200

5	201	CTTTGGTCCATCTCAATGGGATGCATTCTGGTCAAATT	240
10	241	GAGCAGTTGATCAACCAGAGGGATCGAAGAGTTCGCCAGGA	280
15	281	ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA	320
20	321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
25	361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT	400
30	401	TCAACGACATGAACAGCGCTTGACCACAGCTATCCCATT	440
35	441	GTTCGCAGTCCAGAACTACCAAGTCTCTCTTGTCCGTG	480
40	481	TACGTTCAAGCAGCTAACCTTCACCTCAGCGTGTTCGAG	520
45	521	ACGTTAGCGTGTGGGCAAAGGTGGGATTCGATGCTGC	560
50	561	AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600
55	601	GGAAACTACACCGACCACGCTGTTCGTTGGTACAACACTG	640
60	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680
65	681	TAGATACAACCAGTTCAAGAGAGAAATTGACCCCTCACAGTT	720
70	721	TTGGACATTGTGTCTCTTCCCAGAACTATGACTCCAGAA	760
75	761	CCTACCCCTATCCGTACAGTGTCCCAACTTACCAAGAGAAAT	800

5	801	CTATACTAACCCAGTTCTTGAGAACCTCGACGGTAGCTTC	840
10	841	CGTGGTTCTGCCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
15	881	CACACTTGACATCTGAAACAGCATAACTATCTACAC	920
20	921	CGATGCTCACAGAGGAGAGTATTACTGGCTGGACACCAG	960
25	961	ATCATGGCCTCTCCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
30	1001	CCTTCCTCTATGAACTATGGAAACCGCCGCTCCACA	1040
35	1041	ACAACGTATCGTTGCTCAACTAGGTCAAGGTGTCTACAGA	1080
40	1081	ACCTTGTCTTCCACCTGTACAGAAGACCCCTCAATATCG	1120
45	1121	GTATCAACAACCAAGCAACTTCCGTCTTGACGGAACAGA	1160
50	1161	GTTCGCCATATGAAACCTCTTCTAACCTGCCATCCGCTGTT	1200
55	1201	TACAGAAAGAGCGGAACCGTTGATTCCCTGGACGAAATCC	1240
60	1241	CACCAAGAACAAATGTGCCACCCAGGCRAGGATTCTC	1280
65	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTTC	1320
70	1321	AGCAACAGTCCGTGAGCATCATCAGAGCTCTATGTTCT	1360
75	1361	CTTGGATACACCGTAGTGCTGAGTTAACACATCATCGC	1400

5 1401 ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC 1440  
 1441 TTTCTCTTCAACGGTCTGTCATTCAGGACCAGGATTCA 1480  
 10 1481 CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA 1520  
 15 1521 CATTAGAATAGAGGGTATATTGAAGTTCCAATTCACTTC 1560  
 1561 CCATCCACATCTACCAGATATAGAGTTCGTGTGAGGTATG 1600  
 20 1601 CTTCTGTGACCCCTATTCACCTCAACGTTAATTGGGTAA 1640  
 1641 TTCATCCATCTTCTCCAATACAGTTCCAGCTACAGCTACC 1680  
 25 1681 TCCTTGGATAATCTCCAATCCAGCGATTCGGTTACTTTG 1720  
 1721 AAAGTGCCATGCTTTACATCTTCACTCGGTAAACATCGT 1760  
 30 1761 GGGTGTAGAAACTTGTGGACTGCAGGAGTGATTATC 1800  
 1801 GACAGATTGAGTTCAATTCACTGCAACACTCGAGG 1840  
 35 1841 CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTAAATGCG 1880  
 40 1881 CTGTTACGTCTACAAACCAGCTTGGACTCAAGACAAATG 1920  
 45 1921 G 1921;

E. A structural gene encoding the full-length insecticidal protein of *B.t.k.* HD-73 having the sequence:

1	ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA	40
41	ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA	80
81	ACCGATTGAAACCGGTTACACTCCCATCGACATCTCCTTG	120
121	TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG	160
161	CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT	200
201	CTTTGGTCCATCTCAATGGGATGCATTCCCTGGTCAAATT	240
241	GAGCAGTTGATCAACCAAGAGGGATCGAAGAGTTCGCCAGGA	280
281	ACCAGGCCATCTCTAGGTTGAAAGGATTGAGCAATCTCTA	320
321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT	400
401	TCAACGACATGAACAGCGCTTGACCACAGCTATCCCATT	440
441	GTTCGCAGTCCAGAACTACCAAGTTCCCTCTTGTCCGTG	480
481	TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTCGAG	520
521	ACGTTAGCGTGTGGCAAAGGTGGGATTGATGCTGC	560
561	AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600

5	601	GGAAACTACACCGACCACGCTGTTGGTACAAACACTG	640
10	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680
15	681	TAGATACAACCAAGTCAGGAGAGAATTGACCCACAGTT	720
20	721	TTGGACATTGTGTCCTCTTCCGAACATATGACTCCAGAA	760
25	761	CCTACCCCTATCCGTACAGTGTCCCAACTTACCAAGAGAAAT	800
30	801	CTATACTAACCCAGTTCTTGAGAACCTCGACGGTAGCTTC	840
35	841	CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
40	881	CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC	920
45	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAAG	960
50	961	ATCATGGCCTCTCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
55	1001	CCTTTCCCTCTATGAACTATGGAAACCGCCGCTCCACA	1040
60	1041	ACAACGTATCGTTGCTCAACTAGGTCAAGGGTGTACAGA	1080
65	1081	ACCTTGTCTTCCACCTTGACAGAAGACCCCTCAATATCG	1120
70	1121	GTATCAACAACCAAGCAACTTCCGTTCTGACGGAACAGA	1160
75	1161	GTTGCCCTATGGAACCTCTAACTTGGCATCCGCTGTT	1200

5	1201	TACAGAAAGAGCGGAACCGTTGATTCCCTTGGACGAAATCC	1240
10	1241	CACCAACAGAACAAACATGTGCCACCCAGGCAAGGATCTC	1280
15	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTTC	1320
20	1321	AGCAACAGTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
25	1361	CTTGGATAACCCGTAGTGCTGAGTCAACAACATCATCGC	1400
30	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
35	1441	TTTCTCTTCAACGGTCTGTCATTCAAGGACCAGGATTCA	1480
40	1481	CTGGTGGAGACCTCGTTAGACTCACAGCAGTGGAAATAA	1520
45	1521	CATTAGAAATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
50	1561	CCATCCACATCTACCAAGATATAGAGTTCTGTGAGGTATG	1600
55	1601	CTTCTGTGACCCCTATTCAACCTCAACGTTAATTGGGTAA	1640
	1641	TTCATCCATCTTCTCCAATACAGTTCCAGCTACAGCTACC	1680
	1681	TCCTTGGATAATCTCCAATCCAGCGGATTTGGTTACTTTG	1720
	1721	AAAGTCCAATGCTTTACATCTTCACTCGGTAAACATCGT	1760
	1761	GGGTGTTAGAAACTTTAGTGGGACTGCAGGAGTGATTATC	1800

5	1801	GACAGATTCGAGTTCATTCAGTTACTGCAACACTCGAGG	1840
10	1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
15	1881	GCTGTTACGTCTACAAACCAGCTCGGCCTCAAGACCAAT	1920
20	1921	GTGACGGATTATCATATTGATCAAGTGTCCAATTGGTGA	1960
25	1961	CCTACCTCAGCGATGAGTTCTGTCGGATGAAAGCGAGA	2000
30	2001	ATTGTCCGAGAAAAGTCAAACATGCGAAGCGACTCAGTGAT	2040
35	2041	GAACGCAATTACTCCAAGATTCAAATTCAAGACATTA	2080
40	2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120
45	2121	TACCATCCAGGGAGGTGACGACGTGTTCAAGGAGAACTAC	2160
50	2161	GTCACACTATCAGGTACCTTGATGAGTGCTATCCAACAT	2200
55	2201	ACCTCTACCAGAACGATCGACGAGTCCAAGTTGAAAGCCTT	2240
60	2241	TACCCGTTATCAATTAGAGGGTATATCGAAGATAAGTCAA	2280
65	2281	GACCTCGAGATCTACCTCATCCGCTACAATGCAAAACATG	2320
70	2321	AAACAGTAAATGTGCCAGGTACGGGTTCTTATGCCGCT	2360
75	2361	TTCAAGCCCCAAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400

5	2401	CGATGCGGCCACACCTTGAATGGAATCCTGACTTAGATT	2440
10	2441	GTTCTGTAGGGATGGAGAAAAGTGTGCCCATTCGCA	2480
15	2481	TCATTCTCCTTAGACATTGATGTAGGATGTACAGACTTA	2520
20	2521	AATGAGGACCTAGGTGTATGGGTGATCTTAAGATTAAGA	2560
25	2561	CGCAAGATGGGCACCGAACAGACTAGGAACTAGAGTTCT	2600
30	2601	CGAAGAGAAACCATTAGTAGGAGAACCGCTAGCTCGTGTG	2640
35	2641	AAAAGAGCGGAGAAAAAATGGAGAGACAAACGTGAGAAGT	2680
40	2681	TGGAATGGGAGACCAACATCGTCTACAAAGAGGGCAAAAGA	2720
45	2721	ATCTGTAGATGCTTATTGTAAACTCTCAATATGATCAA	2760
50	2761	TTACAGCGGATACGAATATTGCCATGATTGATGCCAG	2800
55	2801	ATAAACGTGTTCATAGCATTGAGAACGTTATCTGCGTGA	2840
60	2841	GCTGTCGTGATTCCGGGTGTCAATGCGGCTATTTTGAA	2880
65	2881	GAATTAGAAGGGCGTATTTCACTGCATTCTCCCTACG	2920
70	2921	ATGCCAGAACGTCAAGAACGGTACTTCACAATGG	2960
75	2961	CTTATCCTGCTGGAACGTGAAAGGGCATGATGTAGAA	3000

5	3001	GAACAAAACAACCAACGTTCGGTCTTGTGTCGGAAAT	3040
10	3041	GGGAAGCAGAAGTGTACAAGAAGTCGTGTCGTCCGGG	3080
15	3081	TCGTGGCTATATCCTTCGTGTCACAGCGTACAAGGAGGG	3120
20	3121	TATGGAGAAGGTTGGTAACCATTCATGAGATCGAGAACAA	3160
25	3161	ATACAGACGAACTGAAGTTAGCAACTGCGTAGAAGAGGA	3200
30	3201	AATCTATCCAATAAACCGTAACGTGTAATGATTATACT	3240
35	3241	GTAAATCAAGAAGAATAACGGAGGTGCGTACACTTCGTA	3280
40	3281	ATCGAGGATATAACGAAGCTCCTCCGTACCGCTGATTA	3320
45	3321	TGCGTCAGTCTATGAAGAAAATCGTATACAGATGGACGA	3360
50	3361	AGAGAGAAATCCTTGTGAATTAAACAGAGGGTATAGGGATT	3400
55	3401	ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA	3440
60	3441	ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
65	3481	GAAACGGAAGGAACATTATCGTGGACAGCGTGGATTAC	3520
70	3521	TCCTTATGGAGGAA 3534.	

F. A structural gene encoding a full-length insecticidal protein of *B.t.k* HD-73 having the sequence:

1	ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA	40
5		
41	ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA	80
10		
81	ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG	120
15		
121	TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG	160
20		
161	CTGGGTTCGTTCTGGACTAGTTGACATCATCTGGGTAT	200
25		
201	CTTGGTCCATCTCAATGGGATGCATTCTGGTGCAAATT	240
30		
241	GAGCAGTTGATCAACCAAGAGGGATCGAAGAGTTCGCCAGGA	280
35		
281	ACCAAGGCCATCTCTAGGTTGGAAGGGATTGAGCAATCTCTA	320
40		
321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
45		
361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT	400
50		
401	TCAACGACATGAAACAGCCCTTGACCACAGCTATCCCATT	440
55		
441	GTTCGCAGTCCAGAACTACCAAGTTCTCTTGTCCGTG	480
60		
481	TACGTTCAAGCAGCTAATCTTCACCTCAGCGTCTTCGAG	520

5	521	ACGTTAGCGTGTGGGCAAAGGTGGGATTGGATGCTGC	560
10	561	AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600
15	601	GGAAACTACACCGACCACGCTGTTGGTACAACACTG	640
20	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680
25	681	TAGATAACAACCAAGTTCAAGGAGAGAATTGACCTCACAGTT	720
30	721	TTGGACATTGTGTCTCTTCCCGAACTATGACTCCAGAA	760
35	761	CCTACCCCTATCCGTACAGTGTCCCAACTTACCAAGAGAAAT	800
40	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
45	841	CGTGGTTCTGCCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
50	881	CACACTTGATGGACATCTGAAACAGCATAACTATCTACAC	920
55	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG	960
60	961	ATCATGGCCTCTCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
65	1001	CCTTCCTCTATGAACTATGGAAACGCCGCTCCACA	1040
70	1041	ACAACGTATCGTTGCTCAACTAGGTCAAGGGTGTACAGA	1080
75	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCCTCAATATCG	1120

5	1121	GTATCAACAACCAGCAACTTCCGTTCTGACGGAACAGA	1160
10	1161	GTTCGCCCTATGGAACCTCTTCTAACCTGCCATCCGCTGTT	1200
15	1201	TACAGAAAGAGCGGAACCGTTGATTCCTGGACGAAATCC	1240
20	1241	CACCAAGAGAACAAATGTGCCACCCAGGCAAGGATTCTC	1280
25	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTG	1320
30	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
35	1361	CTTGGATAACCCGTAGTGCTGAGTTCAACAAACATCATCGC	1400
40	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
45	1441	TTTCTCTAACGGTTCTGTCAATTTCAGGACCAGGATTCA	1480
50	1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
55	1521	CATTCAAGATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
60	1561	CCATCCACATCTACCAAGATATAGAGTTCGTGTGAGGTATG	1600
65	1601	CTTCTGTGACCCCTATTCAACCTCAACGTTAATTGGGGTAA	1640
70	1641	TTCATCCATCTTCTCCAATACAGTTCCAGCTACAGCTACC	1680
75	1681	TCCTTGGATAATCTCCAATCCAGCGATTCGGTTACTTTG	1720

1721	AAAGTGCCAATGCTTTACATCTTCACTCGGTAAACATCGT	1760
1761	GGGTGTTAGAAACTTACTGGGACTGCAGGAGTGATTATC	1800
1801	GACAGATTGAGTTCATTCCAGTTACTGCAACACTCGAGG	1840
1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
1881	GCTGTTACGTCTACAAACCAACTAGGGCTAAAAACAAAT	1920
1921	GTAACGGATTATCATATTGATCAAGTGTCCAATTAGTTA	1960
1961	CGTATTATCGGATGAATTTGTCCTGGATGAAAAGCGAGA	2000
2001	ATTGTCGAGAAAGTCAAACATGCGAAGCGACTCAGTGAT	2040
2041	GAACCGAATTTACTCCAAGATTCAAATTCAAAGACATTA	2080
2081	ATAGGCAACCAGAACGTGGTGGGGCGGAAGTACAGGGAT	2120
2121	TACCATCCAAGGAGGGATGACGTATTTAAAGAAAATTAC	2160
2161	GTCACACTATCAGGTACCTTGATGAGTGCTATCCAACAT	2200
2201	ATTTGTATCAAAAATCGATGAATCAAATTAAAAGCCTT	2240
2241	TACCCGTTATCAATTAAGAGGGTATATCGAAGATAGTCAA	2280
2281	GACTTAGAAATCTATTTAATTCGCTACAATGCAAAACATG	2320

5	2321	AAACAGTAAATGTGCCAGGTACGGGTTCTTATGGCCGCT	2360
10	2361	TTCAGCCAAAGTCCAATCGAAAGTGTGGAGAGCCGAAT	2400
15	2401	CGATGCCGCCACACCTTGAATGGAATCCTGACTTAGATT	2440
20	2441	GTTCGTGTAGGGATGGAGAAAAGTGTGCCCATCATTCCA	2480
25	2481	TCATTTCTCTTAGACATTGATGTAGGATGTACAGACTTA	2520
30	2521	AATGAGGACCTAGGTGTATGGGTGATCTTAAGATTAAGA	2560
35	2561	CGCAAGATGGCACCCCAAGACTAGGAAATCTAGAGTTCT	2600
40	2601	CGAAGAGAAACCATTAGTAGGAGAACCGCTAGCTCGTGTG	2640
45	2641	AAAAGACGGAGAAAAATGGAGAGAACCGTAAAAAT	2680
50	2681	TGGAATGGAAACAAATATCGTTATAAAGAGGCAAAAGA	2720
55	2721	ATCTGTAGATGCTTTATTGTAAACTCTCAATATGATCAA	2760
	2761	TTACAAGCGGATACGAATATTGCCATGATTGCGGCAG	2800
	2801	ATAAACGTGTTCATAGCATTGAGAAGCTTATCTGCCCTGA	2840
	2841	GCTGTCTGTGATTCCGGGTGTCAATGCGGTATTTTGAA	2880
	2881	GAATTAGAAGGGCGTATTTCACTGCATTCTCCCTATATG	2920

5	2921	ATCCGAGAAATGTCATTAAAATGGTATTAAATAATGG	2960
10	2961	CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA	3000
15	3001	GAACAAAACAACCAACGTCGGTCCCTGTTCCCGGAAT	3040
20	3041	GGGARGCAGAAGTGTACAAGAAGTTCGTGTCTGTCCGGG	3080
25	3081	TCGTGGCTATATCCTCGTGTACAGCGTACAAGGAGGGA	3120
30	3121	TATGGAGAAGGTTGCGTAACCATTGAGATCGAGAACAA	3160
35	3161	ATACAGACGAACGTAAAGTTAGCAACTGCGTAGAAGAGGA	3200
40	3201	AATCTATCCAAAATAACACGGTAACGTGTAATGATTATACT	3240
45	3241	GTAAATCAGAAGAATACGGAGGTGCGTACACTCTCGTA	3280
50	3281	ATCGAGGGATATAACGAAGCTCCTCCGTACCAAGCTGATTA	3320
55	3321	TGCGTCAGTCTATGAAAGAAAAATCGTATAACAGATGGACGA	3360
60	3361	AGAGAGAATCCTGTGAATTAAACAGAGGGTATAGGGATT	3400
65	3401	ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA	3440
70	3441	ATACTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
75	3481	GAAACGGAAGGAACATTATCGTGGACAGCGTGGATTAC	3520
80	3521	TCCTTATGGAGGAA 3534.	

G. A structural gene encoding a full-length insecticidal protein of *B.t.k.* HD-73 having the sequence:

1	ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA	40
41	ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA	80
81	ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG	120
121	TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG	160
161	CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT	200
201	CTTGGTCCATCTCAATGGGATGCATTCTGGTGCATT	240
241	GAGCAGTTGATCAACCAGAGGATCGAAGAGTTGCCAGGA	280
281	ACCAGGCCATCTCTAGGTTGAAAGGATTGAGCAATCTCTA	320
321	CCAAATCTATGCAGAGAGCTTCAGAGACTGGGAAGCCGAT	360
361	CCTACTAACCCAGCTCTCCGCAGGAAATGCGTATTCAAT	400
401	TCAACCGACATGAAACAGCGCCTTGACCACAGCTATCCCATT	440

5	441	GTTCGCAGTCAGAACTACCAAGTCCCTCTTGTCCGTG	480
10	481	TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTCGAG	520
15	521	ACGTTAGCGTGTGGCAAGGTGGGATTGATGCTGC	560
20	561	AACCATAATAGCCCTTACAACGACCTTACTAGGCTGATT	600
25	601	GGAAACTACACCGACCACGCTGTTGGTACAACACTG	640
30	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680
35	681	TAGATAACCAACCAAGTTCAAGGAGAGAATTGACCCCTCACAGTT	720
40	721	TTGGACATTGTGTCTCTTCCCGAACTATGACTCCAGAA	760
45	761	CCTACCCCTATCCGTACAGTGTCCCCAACTTACCAAGAGAAAT	800
50	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
55	841	CGTGGTTCTGCCAAGGTATCGAACAGCTCCATCAGGAGCC	880
60	881	CACACTTGATGGACATCTTGAAACAGCATAACTATCTACAC	920
65	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG	960
70	961	ATCATGGCCTCTCCAGTTGATTCAAGCGGGCCCGAGTTA	1000
75	1001	CCTTTCCCTCTATGGAACATGGAAACGCCGCTCCACA	1040

1041	ACAAACGTATCGTTGCTCAACTAGGTCAGGGTGTACAGA	1080
1081	ACCTTGTCTTCCACCTTGTACAGAAAGACCCCTCAATATCG	1120
1121	GTATCAAACACCAGCAACTTCCGTTCTGACGGAACAGA	1160
1161	GTTCGCCTATGGAACCTCTTCTAACCTGCCATCCGCTGTT	1200
1201	TACAGAAAGAGCGGAAACCGTTGATTCCTGGACGAAATCC	1240
1241	CACCAACAGAACAAATGTGCCACCCAGGAAGGATTCTC	1280
1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTTC	1320
1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
1361	CTTGGATACACCGTAGTGCTGAGTCACAAACATCATCGC	1400
1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
1441	TTTCTCTCAACGGTCTGTCATTCAAGGACCAGGATTCA	1480
1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
1521	CATTCAAAATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
1561	CCATCCACATCTACCAGATATAGAGTTCGTGTGAGGTATG	1600
1601	CTTCTGTGACCCCTATTCAACCTCAACGTTAATTGGGGTAA	1640

5	1641	TTCATCCATCTTCTCCAATACAGTTCCAGCTACAGCTACC	1680
10	1681	TCCTTGGATAATCTCCAATCCAGCGATTCGGTTACTTTG	1720
15	1721	AAAGTGCCAATGCTTTACATCTTCACTCGGTAAACATCGT	1760
20	1761	GGGTGTTAGAAAACCTTAGTGGGACTGCAGGAGTGATTATC	1800
25	1801	GACAGATTGAGTTCATTCAGTTACTGCAACACTCGAGG	1840
30	1841	CTGAGTACAACCTTGAGAGAGCCCAGAAGGCTGTGAACGC	1880
35	1881	CCTCTTACCTCCACCAATCAGCTGGCTTGAAAACTAAC	1920
40	1921	GTTACTGACTATCACATTGACCAAGTGTCCAACTTGGTCA	1960
45	1961	CCTACCTTAGCGATGAGTTCTGCCCTGACCGAGAACCGTGA	2000
50	2001	ACTCTCCGAGAAAAGTTAACACGCCAACGCGTCTAGCGAC	2040
55	2041	GAGAGGAATCTTGCAGACTCCAACCTCAAAGACATCA	2080
	2081	ACAGGCAGCCAGAACGTGGTTGGGGTGGAACGCCGGAT	2120
	2121	CACCATCCAAGGAGGCCAGATGTGTTCAAGGAGAACTAC	2160
	2161	GTCACCCCTCTCCGGAACCTTCGACCGAGTGCTACCCCTACCT	2200
	2201	ACTTGTACCAAGAGATCGATGAGTCCAAACTCAAAGCCTT	2240

5	2241	CAACAGGTATCAACTTAGAGGCTACATCGAAGACAGCCAA	2280
10	2281	GACCTTGAATCTACTCGATCAGGTACAATGCCAAGCACG	2320
15	2321	AGACCGTGAATGTCCCAGGTACTGGTCCCTCTGGCCACT	2360
20	2361	TTCTGCCAATCTCCATTGGGAAGTGTGGAGAGCCTAAC	2400
25	2401	AGATGCCCTCCACACCTTGAGTGGAAATCTGACTTGGACT	2440
30	2441	GCTCCTGCAGGGATGGCGAGAAGTGTGCCACCATTCTCA	2480
35	2481	TCACTTCTCCTTGGACATCGATGTGGGATGTACTGACCTG	2520
40	2521	AATGAGGACCTCGGAGTCTGGGTCACTTCAAGATCAAGA	2560
45	2561	CCCAAGACGGACACCGAACAGACTTGGCAACCTTGAGTTCT	2600
50	2601	CGAAGAGAACCATGGTCGGTGAAGCTCTCGCTCGTGTG	2640
55	2641	AAGAGAGCAGAGAAGAAGTGGAGGGACAAACGTGAGAAC	2680
	2681	TCGAATGGGAAACTAACATCGTTACAGGGAGGCCAAAGA	2720
	2721	GTCCCGTGGATGCTTGTTCGTGAACCTCCAAATATGATCAG	2760
	2761	TTGCAAGCCGACACCAACATGCCATGATCCACGCCGCAG	2800
	2801	ACAAACGTGTGCACAGCATTGAGGCTTACTTGCCTGA	2840

5	2841	GTTGTCGTGATCCCTGGTGTGAACGCTGCCATCTCGAG	2880
10	2881	GAACTTGAGGGACGTATCTTACCGCATTCTCCTTGTACG	2920
15	2921	ATGCCAGAAACGTCATCAAGAACGGTACTTCAACAATGG	2960
20	2961	CCTCAGCTGCTGGAATGTGAAAGGTATGTGGACGTGGAG	3000
25	3001	GAACAGAACAACTCAGCGTTCCCTGGTTGTGCCTGAGT	3040
30	3041	GGGARGCTGAAGTGTCCCAAGAGGTTAGAGTCTGTCCAGG	3080
35	3081	TAGAGGCTACATTCTCCGTGTGACCGCTTACAAGGAGGGA	3120
40	3121	TACGGTGAGGGTTGCGTGAACCATCCACGAGATCGAGAAC	3160
45	3161	ACACCGACGAGCTTAAGTTCTCCAAC TGCGTCGAGGAAGA	3200
50	3201	AATCTATCCAACAACACCGTTACTTGCAACGACTACACT	3240
55	3241	GTGAATCAGGAAGAGTACGGAGGTGCCTACACTAGCCGTA	3280
60	3281	ACAGAGGTTACAACGAAGCTCCTCCGTTCTGCTGACTA	3320
65	3321	TGCCTCCGTGTACGAGGAGAAATCCTACACAGATGGCAGA	3360
70	3361	CGTGAGAACCCCTTGCAGTTCAACAGAGGTTACAGGGACT	3400
75	3401	ACACACCACTTCCAGTTGGCTATGTTACCAAGGAGCTTGA	3440

5 3441 GTACTTCCTGAGACCGACAAAGTGTGGATCGAGATCGGT 3480

10 3481 GAAACCGAGGGAACCTTCATCGTGGACAGCGTGGAGCTTC 3520

15 3521 TCTTGATGGAGGAA 3534

H. A structural gene which encodes an insecticidal protein of *B.tu* having the sequence:

16 1 ATGACTGCAGACAAACAACACCGAAGCCCTCGACAGTTCTA 40

20 41 CCACTAAGGATGTTATCCAGAAGGGTATCTCCGGTGTGGG 80

25 81 AGACCTCTTGGCGTGGTTGGATTCCCTTCGGTGGAGCC 120

121 CTCGTGAGCTTCTATACAAACTTCTCAACACCATTGGC 160

30 161 CAAGCGAGGACCCCTGGAAAGCATTCATGGAGCAAGTTGA 200

35 201 AGCTCTTATGGATCAGAAGATTGCAGATTATGCCAAGAAC 240

241 AAGGCCTTGGCAGAACTCCAGGGCCTTCAGAACAAATGTGG 280

40 281 AGGACTACGTGAGTGCATTGTCAGCTGGCAGAAGAACCC 320

321 TGTTAGCTCCAGAAATCCTCACAGCCAAGGTAGGATCAGA 360

45 361 GAGTTGTTCTCTCAAGCCGAATCCCACTTAGAAATTCCA 400

50

55

401	TGCCTAGCTTGCTATCTCCGGTTACGAGGTTCTTCCCT	440
441	CACTACCTATGCTCAAGCTGCCAACACCCACTTGTTCCTC	480
481	CTTAAGGACGCTCAAATCTATGGAGAAGAGTGGGGATACG	520
521	AGAAAGAGGACATTGCTGAGTTCTACAAGCGTCACTTAA	560
561	GCTCACCAAGAGTACACTGACCATTGCGTGAATGGTAT	600
601	AACGTTGGTCTCGATAAGCTCAGAGGCTTCCCTACGAGT	640
641	CTTGGGTGAACCTCAACAGATAACAGGAGAGAGATGACCTT	680
681	GACTGTGCTCGATCTTATCGCACTCTTCCCTTGTACGGAT	720
721	GTGAGACTCTACCCAAAGGAAGTAAAAACTGAGCTTACCA	760
761	GAGACGTGCTCACTGACCCATTGTCGGAGTCACAAACCT	800
801	TAGGGTTATGGAACTACCTTCAGCAATATCGAAACTAC	840
841	ATTAGGAAACCACATCTTCGACTATCTTCACAGAATTG	880
881	AATTCCACACAAGGTTCAACCCAGGATACTATGGTAAACGA	920
921	CTCCTCAACTATTGGTCCCGTAACATGTTCCACCAGA	960
961	CCAAGCATTGGATCTAATGACATCATCACATCTCCCTCT	1000

5 1001 ATGGTAACAAGTCCAGTGAACCTGTGCAGAACCTTGAGTT 1040  
 1041 CAACGGCGAGAAAGTCTATAGAGCCGTCGCAAACACCAAT 1080  
 10 1081 CTCGCTGTGGCCATCCGAGTTACTCAGGGTCACAA 1120  
 15 1121 AGGTGGAGTTAGTCAGTATAACGATCAGACCGATGAGGC 1160  
 1161 CAGCACCCAGACTTACGACTCCAACGTAACGTTGGCGCA 1200  
 20 1201 GTCTCTTGGGATTCTATCGACCAATTGCCCTCCAGAAACCA 1240  
 1241 CAGACGAACCATTGGAGAAGGGCTACAGCCACCAACTTAA 1280  
 25 1281 CTATGTGATGTGCTTCTTGATGCAAGGTTCCAGAGGGACC 1320  
 1321 ATTCCACTGTTGACCTGGACACACAAGTCCGTGGACTTCT 1360  
 30 1361 TCAACATGATCGATAGCAAGAAGATCACTCAACTTCCCTT 1400  
 1401 GGTGAAAGCCTACAAGCTGCAATCTGGTGCTTCGTTGTC 1440  
 35 1441 GCAGGGTCCCAGATTCACTGGAGGTGACATCATCCAGTGCA 1480  
 40 1481 CAGAGAACGGCAGCCAGCTACTATCTACGTGACACCTGA 1520  
 45 1521 TGTGTCTTACTCTCAGAAGTACAGGGCACGTATTCAATTAC 1560  
 1561 GCATCTACCAGCCAGATCACCTCACACTCAGCTTGGATG 1600

50

55

5  
 1601 GAGCACCCCTCAACCAGTATTACTTGCACAGACCATCAA 1640  
 1641 CAAAGGTGACACTCTCACATACAATAGCTCACTGGCA 1680  
 10  
 1681 AGTTTCAGCACACCAATTGAACTCTCAGGCAACAATCTTC 1720  
 15  
 1721 AGATCGGCGTCACCGGTCTCAGCGCCGGAGACAAAGTCTA 1760  
 1761 CATCGACAAGATTGAGTTCATCCCAGTGAAC 1791

20 I. A structural gene which encodes an insecticidal protein of *B.t. entomocidus* having the sequence:

25  
 1 ATGGAGGGAGAACCAACCAAAACCAATGCATTCCATACAACT 40  
 41 GCTTGAGTAACCCAGAAGAGGTATTGCTTGATGGAGAACG 80  
 30  
 81 CATTCAACCGGTAACTCTTCATCGACATCTCCTTGTCC 120  
 121 TTGGTCCAGTTCTGGTCAGCAACTTCGTGCCAGGTGGTG 160  
 35  
 161 GGTTCCCTTGTCCGGACTAATTGACTTCGTCTGGGTATCGT 200  
 201 TGGTCCATCTCAATGGGATGCATTCTGGTGCAAATTGAG 240  
 40  
 241 CAGTTGATCAACGAGAGGATCGCTGAGTTGCCAGGAACG 280  
 45  
 281 CTGCCATCGCTAATTGGAAGGATTGGCAATAACTTCAA 320

5	321	CATCTATGTGGAGGCCTCAAAGAGTGGGAAGAGGGACCT	360
10	361	AAACAACCCAGAGACCCGCACTAGGGTGATCGACAGATCA	400
15	401	GAATCTTGGACGGCCTCTGGAGAGAGATATCCATCCTT	440
20	441	CAGAACATCTGGCTTCGAAGTTCTCTTGTCCGTGTAC	480
25	481	GCTCAAGCAGCTAATCTCACCTCGCTATCCTCGAGACA	520
30	521	GTGTCATCTTGGGAAAGGTGGGATTGACCCTATCAA	560
35	561	CGTCAATGAGAATTACAACAGACTTATCAGGCACATTGAC	600
40	601	GAGTACGCCGACCACGTGCTAACACCTACAACCGTGGCT	640
45	641	TGACAATCTCCCTAAGTCTACTTATCAAGATTGGATTAC	680
50	681	CTACAACAGGTTGAGGGAGAGACTTGACCCCTCACAGTTTG	720
55	721	GACATTGCAGCTTCTTCCCGAACTATGACAACAGGAGAT	760
	761	ACCCCTATCCAACCAGTGGTCAACTTACCCAGAGAAGTCTA	800
	801	TACTGACCCACTTATCAACTTCAACCCCTCAGTTGCAAAGT	840
	841	GTCGCCAACTTCCCACATTCAACGTCATGGAGTCCAGCC	880
	881	GTATCAGGAACCCACACTTGTGACATCTTGAACAAACCT	920

5	921	TAATATCTCACCGATTGGTTCAGCGTTGGCGTAACCTC	960
10	961	TATTGGGGTGGACACAGGGTCACTCTCTCTTATTGGAG	1000
15	1001	GTGGGAACATTACCTCTCCTATCTATGGACGTGAGGCAA	1040
20	1041	CCAGGAGCCACCACGTAGTTCACCTCAACGGTCCAGTC	1080
25	1081	TTCAGAACCTTGTCTAACCTACCTTGAGATTGCTCCAGC	1120
30	1121	AACCTTGGCCAGCTCCACCTTCACCTTAGAGGTGTTGA	1160
35	1161	GGGC GTT GAG TT CT CT ACT CCT ACCA ACT CCT CACT TAC	1200
40	1201	AGAGGTAGAGGAACCGTTGATT CCTTGACCGA ACT CCCAC	1240
45	1241	CAGAGGACAATAGCGTGCCACCCAGGGAAAGGCTACTCCCA	1280
50	1281	CAGGTTGTGCCACGCAACCTCGTGCAGCGTTCCGGAACT	1320
55	1321	CCATT CCTCACTACAGGAGTTGTGTCTCATGGACTGATC	1360
60	1361	GTAGTGCTACTCTCACTAAATACCAATTGATCCCGAGAGGAT	1400
65	1401	CAATCAAATCCCATTGGTCAAGGGTTCCGTGTGTGGGA	1440
70	1441	GGAACCTCTGTCACTCACAGGACCAGGCTCACAGGAGGTG	1480
75	1481	ATATTCTTAGAAGAACACTTTGGCGACTTGTGAGCCT	1520

5	1521	CCAAGTTAACATCAACTCTCCAATTACTCAAAGATACTGT	1560
10	1561	CTCAGGTTCGTTACGCATCTCCCGTGACGCTAGAGTC	1600
15	1601	TCGTGCTCACCGGAGCCAGCTTCTACCGGTGTCGGTGGACA	1640
20	1641	AGTCTCCGTGAACATGCCACTCCAGAAGACTATGGAGATC	1680
25	1681	GGCGAGAACTTGACATCCAGGACCTTCAGATAACCCGACT	1720
30	1721	TCTCTAACCCCTTCAGTTCCGTGCCAACCTGACATCAT	1760
35	1761	TGGCATTAGCGAACACACCTCTCTTGGAGCTGGTAGCATC	1800
40	1801	TCATCTGGCGAATTGTACATTGACAAGATTGAGATCATTC	1840
45	1841	TTGCCGACGCTACCTCGAGGCTGAGTCTGACCTTGAGAG	1880
50	1881	AGCCCGAGGCTGTGAACGCCCTTTACCTCCTCTAAT	1920
55	1921	CAGATTGGCTTGAAGAACTGACGTTACTGACTATCACATTG	1960
60	1961	ACCAAGTGTCCAACTTGGTCGACTGCCCTAGCGATGAGTT	2000
65	2001	CTGCCCTCGACCGAGAACCGTGAACCTCTCGAGAAAGTTAAA	2040
70	2041	CACGCCAACGCGTCTCAGCGACGAGAGGAATCTTGCAG	2080
75	2081	ACCCCAACTTCAGAGGCATCACAGGCAGCCAGACCGTGG	2120

5	2121	TTGGAGAGGAAGCACCACATCACCATCCAAGGAGGCAC	2160
10	2161	GATGTGTTCAAGGAGAACTACGTACCCCTCCAGGAACGTG	2200
15	2201	TGGACGAGTGCTACCCCTACCTACTTGTACCAGAACATCGA	2240
20	2241	TGAGTCACAACTCAAAGCCTACACCAGGTATGAACCTTAA	2280
25	2281	GGCTACATCGAACAGACAGCCAAGACCTTGAAATCTACCTCA	2320
30	2321	TCAGGTACATGCCAACGACAGGAGATCGTGAATGTCCCAGG	2360
35	2361	TACTGGTCCCCCTGGCCACTTCTGCCAATGCCATT	2400
40	2401	GGGAAGTGTGGAGAGCCTAACAGATGCGCTCCACACCTTG	2440
45	2441	AGTGGAACCTGACTTGGACTGCTCTGCAGGGATGGCGA	2480
50	2481	GAAGTGTGCCACCATTCTCATCACCTCACCTGGACATC	2520
55	2521	GATGTGGATGTACTGACCTGAATGAGGACCTCGGAGTCT	2560
60	2561	GGGTCATTTCAAGATCAAGACCCAAGACGGACACGCAAG	2600
65	2601	ACTTGGCAACCTTGAGTTCTCGAAGAGAAACCATTGCTC	2640
70	2641	GGTGAAGCTCGCTCGTGTGAAGAGAGCAGAGAAGAAGT	2680
75	2681	GGAGGGACAAACGTGAGAAACTCCAACTCGAGACTAACAT	2720

5	2721	CGTTACAAGGAGGCCAAAGAGTCCGTGGATGCTTGTTC	2760
10	2761	GTGAACTCCAATATGATAGGTTCAAGTGGACACCAACA	2800
15	2801	TCGCCATGATCCACCGCTGCAGACAAACGTGTGCACAGGAT	2840
20	2841	TCGTGAGGCTTACTTGCCTGACTTGTCCGTGATCCCTGGT	2880
25	2881	GTGAAACGCTGCCATCTCGAGGAACTTGAGGGACGTATCT	2920
30	2921	TTACCGCATACTCCTTGTACGATGCCAGAACGTCATCAA	2960
35	2961	GAACGGTGACTTCAACAATGCCCTTGTGCTGGAATGTG	3000
40	3001	AAAGGTATGTGGACGTGGAGGAACAGAACAAATCACCGTT	3040
45	3041	CCGTCCCTGGTTATCCCTGAGTGGGAAGCTGAAGTGTCCCA	3080
50	3081	AGAGGTTAGAGTCTGTCCAGGTAGAGGCTACATTCTCCGT	3120
55	3121	GTGACCGCTTACAAGGAGGGATACGGTGAGGGTTGCGTGA	3160
	3161	CCATCCACGAGATCGAGGACAACACCGACGAGCTTAAGTT	3200
	3201	CTCCAATGCGTCGAGGAAGAAGTCTATCCCAACACACCC	3240
	3241	GTTACTTGCAACAACACTACACTGGGACCCAGGAAGAGTACG	3280
	3281	AAGGTACCTACACTAGCCGTAACCAAGGTTACGACGAAAGC	3320

5                   3321 TTACGGAAACAATCCTTCGTTCTGCTGACTATGCCCTCC   3360  
 10                3361 GTGTACGAGGAGAAATCCTACACAGATGGCAGACGTGAGA   3400  
 15                3401 ACCCTTGGAGTCCAACAGAGGTTACGGTACTACACACC   3440  
 20                3441 ACTTCCAGCAGGCTATGTTACCAAGGACCTTGAGTACTTT   3480  
 25                3481 CCTGAGACCGACAAAGTGTGGATCGAGATCGGTGAAACCG   3520  
 30                3521 AGGGAACCTTCATCGTGGACAGCGTGGAGCTCTCTTGAT   3560  
 35                3561 GGAGGAA 3567

25                J. A structural gene which encodes a P2 Insecticidal protein having the sequence:

30                1 ATGGACACAAACGTCTGAACTCTGGTGAACAACCATCT   40  
 35                41 GCGACGCATAACAACGTCGTGGCTACGATCCATTCAAGCTT   80  
 40                81 CGAACACAAGAGCCTCGACACTATTCAAGAAGGAGTGGATG   120  
 45                121 GAATGGAAACGTACTGACCACTCTCTACGTGCGACCTG   160  
 50                161 TGGTTGGAACAGTGTCCAGCTTCTCAAGAAGGTGG   200  
 55                201 CTCTCTCATCGGAAAACGTATCTGTCCGAACCTGGGGT   240

5	241	ATCATTTCCATCTGGGTCCACTAATCTCATGCAAGACA	280
10	281	TCTTGAGGGAGACCGAACAGTTCTCAACCAGCGTCTCAA	320
15	321	CACTGATACTTGGCTAGAGTCACCGCTGAGTTGATCGGT	360
20	361	CTCCAAGCAAACATTGGTGGAGTCACCGCAAGTGGACA	400
25	401	ACTTCTTGAATCCAACCTCAGAACATCCTGTGCCTCTTCCAT	440
30	441	CACTTCTTCCGTGAACACTATGCAGCAACTCTTCCCTAAC	480
35	481	AGATTGGCTCAGTTCTAGATTCAAGGGCTACCAAGTTGCTCC	520
40	521	TTCTTCCACTCTTGTCAAGGCTGCCAACATGCACTTGTC	560
45	561	CTTCATACGTGACGTGATCCTAACCGCTGACGAATGGGA	600
50	601	ATCTCTGCAGCCACTCTTAGGACATACAGAGACTACTTGA	640
55	641	GGAACATACACTCGTGTGATTACTCCAACATTGCATCAACAC	680
60	681	TTATCAGACTGCCTTCGTGGACTCAATACTAGGCTTCAC	720
65	721	GACATGCTTGAGTTCAAGGACCTACATGTTCTTAACGTGT	760
70	761	TTGAGTACGTCAGCATTGGAGTCTCTCAAGTACCAAGAG	800
75	801	CTTGATGGTGTCTGGAGCCAATCTTACGCCCTGGC	840

5	841	AGTGGACCACAGCAAACTCAGAGCTTCACAGCTCAGAACT	880
10	881	GGCCATTCTTGTATAGCTTGTCCAAGTCAACTCCAACTA	920
15	921	CATTCTCAGTGGTATCTCTGGGACCAAGACTCTCCATAACC	960
20	961	TTTCCCAACATTGGTGGACTTCAGGCTCCACTACAACCC	1000
25	1001	ATAGCCTTAACCTCTGCCAGAGTGAACATACAGTGGAGGTGT	1040
30	1041	CAGCTCTGGATTGATTGGTGCAACTAACTTGAACCACAAAC	1080
35	1081	TTCATTTGCTCCACCGCTTGCCACCTCTGAGCACACCGT	1120
40	1121	TTGTGAGGTCCCTGGCTTGACAGCGGTACTGATGCCGAAGG	1160
45	1161	AGTTGCTACCTCTACAAACTGGCAAACCGAGTCCTCCAA	1200
50	1201	ACCACTCTTAGCCTTCGGTGTGGAGCTTCTCTGCACGTG	1240
55	1241	GGAATTCAAACACTACTTCCAGACTACTTCATTAGGAACAT	1280
60	1281	CTCTGGTGTCTCTCGTCATCAGGAATGAAGACCTCACC	1320
65	1321	CGTCCACTTCATTACAACCAAGATTAGGAACATCGAGTCTC	1360
70	1361	CATCCGGTACTCCAGGAGGTGCAAGAGCTTACCTCGTGTG	1400
75	1401	TGTCCATAACAGGAAGAACACATCTACGCTGCCAACGAG	1440

5 1441 AATGGCACCATGATTCACCTTGCACCAAGAGATTACACTG 1480  
 10 1481 GATTCAACCATCTCTCCAATCCATGCTACCCAAGTGAACAA 1520  
 15 1521 TCAGACACGGCACCTTCATCTCCGAAAAGTCGGAAATCAA 1560  
 20 1561 GGTGACTCCITGAGGTTGAGCAATCCAACACTACCGCTA 1600  
 25 1601 GGTACACTTTGAGAGGCAATGGAACAGCTACACCTTTA 1640  
 30 1641 CTTGAGAGTTAGCTCCATTGGTACTCCACCATCCGTGTT 1680  
 35 1681 ACCATCAACGGACGTGTTACACAGTCTCTAATGTGAACA 1720  
 40 1721 CTACAACGAAACATGATGGCTTAACGACAACGGAGCCAG 1760  
 45 1761 ATTCAAGGACATCAACATTGGCAACATCGTGGCCTCTGAC 1800  
 50 1801 AACACTAACGTTACTTGGACATCAATGTGACCCCTCAATT 1840  
 55 1841 CTGGAACTCCATTGATCTCATGAACATCATGTTGTGCC 1880  
 60 1881 AACTAACCTCCCTCCATTGTAC 1902 ; or

45 K. A structural gene sequence encoding a, fusion protein comprising the N-terminal 610 amino acids of *B.t.k.*  
 HD-1 and the C-terminal 567 amino acids of *B.t.k.* HD-73, said gene having the sequence:

1	ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA	40
5		
41	ACTGCTTGAGTAACCCAGAAAGTTGAAGTACTTGGTGGAGA	80
10		
81	ACCGATTGAAACCGGTTACACTCCCATCGACATCTCCTTG	120
15		
121	TCCCTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG	160
20		
161	CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT	200
25		
201	CTTTGGTCCATCTCAATGGGATGCATTCCCTGGTGCAAATT	240
30		
241	GAGCAGTTGATCAACCAAGAGGATCGAAGAGTTGCCAGGA	280
35		
281	ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA	320
40		
321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
45		
361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT	400
50		
401	TCAACCGACATGAACAGCGCTTGACCACAGCTATCCCTT	440
55		

5 441 GTTCGCAGTCCAGAACTACCAAGTTCTCTCTTGTCCGTG 480  
 10 481 TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGTTCGAG 520  
 15 521 ACGTTAGCGTGTGGGCAAAGGTGGGATTCGATGCTGC 560  
 20 561 AACCATCAATAGCCGTACAACGACCTTAAGGCTGATT 600  
 25 601 GGAAACTACACCGACCACGCTGTTCGTTGGTACAACACTG 640  
 30 641 GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT 680  
 35 681 TAGATACAACCAGTTCAAGGAGAGAATTGACCCCTCACAGTT 720  
 40 721 TTGGACATTGTGTCTCTTCCCGAACTATGACTCCAGAA 760  
 45 761 CCTACCCATCCGTACAGTGTCCCAACTTACCGAGAAAT 800  
 50 801 CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC 840  
 55 841 CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC 880  
 60 881 CACACTTGATGGACATCTGAACAGCATAACTATCTACAC 920  
 65 921 CGATGCTCACAGAGGGAGGTATTACTGGCTGGACACCAG 960  
 70 961 ATCATGCCCTCTCCAGTTGATTCAAGCGGGCCCGAGTTA 1000  
 75 1001 CCTTTCCCTCTATGGAACATGGAAACGCCGCTCCACA 1040

5	1041	ACAAACGTATCGTTGCTCAACTAGGTCAAGGGTGTCTACAGA	1080
10	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCCTCAATATCG	1120
15	1121	GTATCAACAAACCAGCAACTTCCGTTCTGACCGAACAGA	1160
20	1161	GTTCGCCCTATGGAACCTCTTCTAACTTGCCATCCGCTGTT	1200
25	1201	TACAGAAAGACCGGAACCGTTGATTCTTGGACGAAATCC	1240
30	1241	CACCAACAGAACAAATGTGCCACCCAGGCAAGGATTCTC	1280
35	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTG	1320
40	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCTATGTTCT	1360
45	1361	CATGGATTCATCGTAGTGCTGAGTTCAACAAATCATCATTCC	1400
50	1401	TTCCCTCTCAAATCACCCAAATCCCATTGACCAAGTCTACT	1440
55	1441	AACCTTGGATCTGGAACCTCTGTCGTGAAAGGACCAGGCT	1480
	1481	TCACAGGAGGTGATATTCTTAGAAGAACTTCTCTGGCCA	1520
	1521	GATTAGCACCCCTCAGAGTTAACATCACTGCACCACTTCT	1560
	1561	CAAAGATATCGTGTCAAGGATTGTTACGCATCTACCACTA	1600
	1601	ACTTGCAATTCCACACCTCCATCGACGGAGGCCTATCAA	1640

5	1641	TCAGGGTAACCTCTCGCAACCATGTCAAGCGGCAGCAAC	1680
10	1681	TTGCAATCCGGCAGCTTCAGAACCGTCGGTTCACTACTC	1720
15	1721	CTTCAACTCTCTAACGGATCAAGCGTTTACCCCTAG	1760
20	1761	CGCTCATGTGTTCAATTCTGGCAATGAAGTGTACATTGAC	1800
25	1801	CGTATTGAGTTGTCCTGCCGAAGTTACCCCTCGAGGCTG	1840
30	1841	AGTACAACCTTGAGAGAGGCCAGAACGGCTGTAAACGCCCT	1880
35	1881	CTTACCTCCACCAATCAGCTGGCTTGAAACTAACGTT	1920
40	1921	ACTGACTATCACATTGACCAAGTGTCCAACCTGGTCACCT	1960
45	1961	ACCTAGCGATGAGTTCTGCCTCGACGAGAACGGCTGAAC	2000
50	2001	CTCCGAGAAAGTTAACACGCCAACGGCTCTCAGCGACGAG	2040
55	2041	AGGAATCTCTGCAAGACTCCAACCTCAAAGACATCAACA	2080
	2081	GGCAGCCAGAACGTGGTTGGGTGGAAGCACGGGATCAC	2120
	2121	CATCCAAGGAGGCACGATGTCAAGGAGAACACTACGTC	2160
	2161	ACCCCTCCGGAACCTTCGACGAGTGCTACCCCTACCTACT	2200
	2201	TGTACCAGAAGATCGATGAGTCCAACCTCAAAGCCTTCAC	2240

5	2241	CAGGTATCAACTTAGAGGCTACATCGAAGACAGCCAAGAC	2280
10	2281	CTTGAATCTACTCGATCAGGTACAATGCCAAGCACGAGA	2320
15	2321	CCGTGAATGTCCCAGGTACTGGTTCCCTCTGGCCACTTTC	2360
20	2361	TGCCCAATCTCCCATTGGGAAGTGTGGAGAGCCTAACAGA	2400
25	2401	TGGCCTCCACACCTTGAGTGGAACTCTGACTTGGACTGCT	2440
30	2441	CCTGCAGGGATGGCGAGAAGTGTGCCACCATTCTCATCA	2480
35	2481	CTTCTCCTGGACATCGATGTGGATGACTCACCTGAAAT	2520
40	2521	GAGGACCTCGGAGTCTGGTCATCTCAAGATCAAGACCC	2560
45	2561	AAGACGGACACGCAAGACTGGCAACCTTGAGTTCTCGA	2600
50	2601	AGAGAAACCATTGGTCGGTGAAGCTCTCGCTCGTGTGAAG	2640
55	2641	AGAGCAGAGAAGAAGTGGAGGGACAAACGTGAGAAACTCG	2680
60	2681	AATGGGAAACTAACATCGTTACAAGGAGGCCAAAGAGTC	2720
65	2721	CGTGGATGCTTGTTCGTGAACCTCCAATATGATCAGTTG	2760
70	2761	CAAGCCGACACCAACATGCCATGATCCACGCCAGACA	2800
75	2801	AACGTGTGCACAGCATTGAGGCTTACTTGCTGAGTT	2840

5	2841	GTCCGTGATCCCTGGTGTGAAACGCTGCCATCTCGAGGAA	2880
10	2881	CTTGAGGGACGTATCTTACCGCATTCTCCTGTACGATG	2920
15	2921	CCAGAACGTCATCAAGAACGGTACTTCACAAATGGCCT	2960
20	2961	CAGCTGCTGGAATGTGAAAGGTATGTGGACGTGGAGGAA	3000
25	3001	CAGAACAAATCAGCGTCCGTCCTGGTTGTGCCTGAGTGGG	3040
30	3041	AAGCTGAAGTGTCCAAAGAGGTTAGAGTCTGTCCAGGTAG	3080
35	3081	AGGCTACATTCTCCGTGTGACCGCTTACAAGGAGGGATAC	3120
40	3121	GGTGAGGGTTGCGTGACCATCCACGAGATCGAGAACACA	3160
45	3161	CCGACGAGCTTAAGTCTCCAATGCCGTCGAGGAARGAAAT	3200
50	3201	CTATCCAAACAACACCGTTACTTGCAACGACTACACTGTG	3240
55	3241	AATCAGGAAGAGTACGGAGGTGCCTACACTAGCCGTAAACA	3280
	3281	GAGGTTACAACGAAGCTCCTCCGTTCTGCTGACTATGC	3320
	3321	CTCCGTGTACGAGGGAGAAATCCTACACAGATGGCAGACGT	3360
	3361	GAGAACCCCTGCGAGTTAACAGAGGTTACAGGGACTACA	3400
	3401	CACCACTTCCAGTTGGCTATGTTACCAAGGAGCTTGAGTA	3440

5 3441 CTTTCCTGAGACCGACAAAGTGTGGATCGAGATCGGTGAA 3480

10 3481 ACCGAGGGAACCTTCATCGTGGACAGCGTGGAGCTCTCT 3520

15 3521 TGATGGAGGAA 3531.

15 **Patentansprüche**

1. Verfahren zur Modifizierung einer Wildtyp-Struktur-Gensequenz, welche für ein insektizides Protein von *Bacillus thuringiensis* codiert, zur Verbesserung der Expression dieses Proteins in Pflanzen, welches umfasst:
  - a) das Identifizieren von Regionen innerhalb dieser Sequenz mit mehr als vier aufeinander folgenden Adenin- oder Thymin-Nukleotiden;
  - b) das Modifizieren der Regionen von Schritt (a), die zwei oder mehr Polyadenylierungssignale innerhalb einer Zehn-Basen-Sequenz aufweisen, um diese Signale zu entfernen, wobei eine Gensequenz, die für dieses Protein codiert, beibehalten wird; und
  - c) das Modifizieren der 15-30-Basen-Regionen, die die Regionen von Schritt (a) umgeben, um Pflanzen-Polyadenylierungs-Hauptsignale, aufeinander folgende Sequenzen, die mehr als ein untergeordnetes Polyadenylierungssignal enthalten, und aufeinander folgende Sequenzen, die mehr als eine ATTTA-Sequenz enthalten, zu entfernen, wobei eine Gensequenz, die für dieses Protein codiert, beibehalten wird.
2. Verfahren zur Modifizierung einer Wildtyp-Struktur-Gensequenz, welche für ein insektizides Protein von *Bacillus thuringiensis* codiert, zur Verbesserung der Expression dieses Proteins in Pflanzen, welches umfasst:
  - a) das Entfernen von Polyadenylierungssignalen, die in diesem Wildtyp-Gen enthalten sind, wobei eine Sequenz, die für dieses Protein codiert, beibehalten wird; und
  - b) das Entfernen von ATTTA-Sequenzen, die in diesem Wildtyp-Gen enthalten sind, wobei eine Sequenz, die für dieses Protein codiert, beibehalten wird.
3. Verfahren nach Anspruch 2, welches weiters das Entfernen von selbstkomplementären Sequenzen und das Ersetzen solcher Sequenzen durch nicht-selbstkomplementäre DNA, welche von Pflanzen bevorzugte Codons aufweist, wobei eine Struktur-Gensequenz, die für dieses Protein codiert, beibehalten wird.
4. Verfahren nach den Ansprüchen 1 bis 3, welches weiters die Verwendung von von Pflanzen bevorzugten Sequenzen beim Entfernen der Polyadenylierungssignale und ATTTA-Sequenzen umfasst.
5. Verfahren nach den Ansprüchen 1 bis 3, bei welchem die Pflanzen-Polyadenylierungssignale ausgewählt sind aus der Gruppe bestehend aus AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AAG-CAT, ATTAAT, ATACAT, AAAATA, ATTAAA, AATTAA, AATACA und CATAAA.
6. Verfahren zur Verbesserung der Expression eines heterologen Gens in Pflanzen, wobei dieses Gen ein modifiziertes chlöräres Gen aufweist, das einen Promotor enthält, der in Pflanzenzellen wirkt, der operabel mit einer strukturellen Codiersequenz und einer 3'-nicht-translatierten Region, die ein Polyadenylierungssignal enthält, in Pflanzen wirkt, um die Addition von Polyadenylylat-Nukleotiden an das 3'-Ende der RNA zu bewirken, verbunden ist, wobei die strukturelle Codiersequenz für ein insektizides Protein codiert, von welchem mindestens ein Teil von einem *Bacillus-thuringiensis*-Protein stammte, wobei das Verfahren das Modifizieren dieser strukturellen Codier-

sequenz umfasst, so dass diese Sequenz eine DNA-Sequenz aufweist, die sich von der natürlicherweise vorkommenden DNA-Sequenz, welche für dieses *Bacillus-thuringiensis*-Protein codiert, unterscheidet und diese strukturelle Codiersequenz nicht mehr als 5 aufeinander folgende Nukleotide aufweist, die entweder aus Adenin- oder aus Thymin-Resten bestehen.

5 7. Verfahren zur Verbesserung der Expression eines heterologen Gens in Pflanzen, wobei dieses Gen ein modifiziertes chlöräres Gen aufweist, das einen Promotor enthält, der in Pflanzenzellen wirkt, der operabel mit einer strukturellen Codiersequenz und einer 3'-nicht-translatierten Region, die ein Polyadenyllierungssignal enthält, das 10 in Pflanzen wirkt, um die Addition von Polyadenylat-Nukleotiden an das 3'-Ende der RNA zu bewirken, verbunden ist, wobei diese strukturelle Codiersequenz für ein Insektizides Protein codiert, von welchem mindestens ein Teil 15 von einem *Bacillus-thuringiensis*-Protein stammt, wobei das Verfahren das Modifizieren dieser strukturellen Codiersequenz umfasst, so dass diese Sequenz eine DNA-Sequenz besitzt, die sich von der natürlicherweise vorkommenden DNA-Sequenz, die für das *Bacillus-thuringiensis*-Protein codiert, unterscheidet und die folgenden Merkmale hat:

15 diese strukturelle Codiersequenz hat eine Region, die zur folgenden Sequenz komplementär ist

GGCTTGATTCTAGCGAACTCTTCGATTCTCTGGTTGATGAGCTGTT  
 20 1 5 10 15 20 25 30 35 40 45

wobei in der Codiersequenz dieser Region 2 AACCAA- und 1 AATTAA-Sequenz eliminiert sind.

25 8. Verfahren nach Anspruch 7, wobei die strukturelle Codiersequenz für ein Insektizides Protein codiert, von welchem mindestens ein Teil von einem *Bacillus thuringiensis kurstaki* HD-1 stammt.

9. Verfahren nach Anspruch 7 oder 8, wobei die Pflanze eine Tabakpflanze ist.

30 10. Modifiziertes chlöräres Gen, das einen Promotor enthält, welcher in Pflanzenzellen wirkt, der operabel mit einer strukturellen Codiersequenz und einer 3'-nicht-translatierten Region, die ein Polyadenyllierungssignal enthält, welches 35 in Pflanzen wirkt, um die Addition von Polyadenylat-Nukleotiden am 3'-Ende der RNA zu bewirken, verbunden ist, wobei diese strukturelle Codiersequenz für ein Insektizides Protein codiert, von welchem mindestens ein Teil von einem *Bacillus thuringiensis*-Protein stammt, wobei diese strukturelle Codiersequenz eine DNA-Sequenz aufweist, die sich von der natürlicherweise vorkommenden DNA-Sequenz, welche für dieses *Bacillus thuringiensis*-Protein codiert, unterscheidet und ausgewählt ist aus:

A. einem Struktur-Gen, welches für ein Insektizides Protein von *B.t.k* HD-1 codiert, mit der Sequenz:

40

45

50

55

1	ATGGCTATAGAAACTGGTTACACCCCAATCGATATTCCT	40
5		
41	TGTCGCTAACGCAATTCTTTGAGTGAAATTGTCGGG	80
10		
81	TGCTGGATTGTGTTAGGACTAGTTGATATTCTGGGG	120
15		
121	ATTTTGCTCCCTCTCAATGGGACGCATTCTTGACAAA	160
20		
161	TTGAACAGCTCATCAACCAGAGAATCGAAGAGTCGCTAG	200
25		
201	GAATCAAGCCATTCTAGATTAGAAGGACTAAGCAATCTT	240
281	TATCAAATTACGCAGAATCTTTAGAGAGTGGAAAGCAG	280
321	ATTCATGACATGAACAGTGCCTTACAACCGCTATTCTT	360
361	CTTTTGCAAGTCAAAATTATCAAGTTCTCTCCCTCCG	400
40		
401	TGTACGTTCAAGCTGCCAACCTCCACCTCTCAGTTTGAG	440
45		
441	AGATGTTCAAGTGGACAAAGGTGGGATTGATGCC	480
481	GCGACTATCAATAGTCGTTATAATGATTAACTAGGCTTA	520
521	TTGGCAACTATACAGATCATGCTGTACGCTGGTACAATAC	560
561	GGGATTAGAGCGTGTATGGGGACCGGAACTCTAGAGATTGG	600
601	ATCAGGTACAACCAGTTCAAGAGAGCTTACACTAACTG	640
641	TATTAGATATCGTTCTCTATTCCGAACTATGATAGTAG	680
681	AACGTATCCAATTGAAACAGTTCCCAATTAAACAAGAGAA	720

5	721	ATTTATACAAACCCAGTATTAGAAAATTTGATGGTAGTT	760
10	761	TTCGAGGCTCGGCTCAGGGCATACAAGGAAGTATTAGGAG	800
15	801	TCCACATTTGATGGATATACCTAATAGTATAACCACATCTAT	840
20	841	ACGGATGCTCATAGAGGAGAACTACTACTGGTCCGGTCACC	880
25	881	AGATCATGGCTTCTCCTGTAGGGTTTCGGGCCAGAATT	920
30	921	CACTTTCCGCTATATGAACTATGGAAATGCAGCTCCA	960
35	961	CAACAACGTATTGTTGCTCACTAGGTCAAGGGCGTGTATA	1000
40	1001	GAACATTATCGTCCACCTTATATAGAACCTTTAACAT	1040
45	1041	CGGGATCAACAACCAACAACATATCTGTTCTGACGGGACA	1080
50	1081	GAATTGCTTATGGAACCTCCTCAAATTGCCATCCGCTG	1120
55	1121	TATACAGAAAAGCGGAACGGTAGATTGCTGGATGAART	1160
60	1161	ACCGCCACAGAATAACACGTGCCACCTAGGCAAGGATTT	1200
65	1201	AGTCATCGATTAAGCCATGTTCAATGTTGCTTCAGGCT	1240
70	1241	TTAGTAATAGTAGTGTAAAGTATAATAAGAGCTCCTATGTT	1280
75	1281	CTCTTGGATAACATCGTAGTGCTGAGTTCAACACATCATC	1320
80	1321	CCTTCATCACAAATCACCCAAATCCCACTCACCAAGTCTA	1360
85	1361	CTAACTTGGCTCTGGAACCTCTGTCGTTAAAGGACCAGG	1400
90	1401	ATTTACAGGAGGAGATATTCTTCAAGAACCTCACCTGGC	1440

1441 CAGATTCACCTTAAGAGTAAATATTACTGCACCAATT 1480  
 1481 CACAAAGATATCGGTAAAGAATTGGCTACGCTTCTACAC 1520  
 1521 AAACCTTCAGTTCCACACATCAATTGACGGAAGACCTATT 1560  
 1561 AATCAGGGAAATTTTCAGCAACTATGAGTAGTAGGGAGTA 1600  
 1601 ATTTACAGTCCGGAAAGCTTTAGGACTGTAGGTTTACTAC 1640  
 1641 TCCGTTAACCTTCAAATGGATCAAGTGTATTACGTTA 1680  
 1681 AGTGCCTCATGTCCTCAATTCAAGGAATGAAGTTATATAG 1720  
 1721 ATCGAATTGAATTGGTCCGGCA 1743

B. einem Struktur-Gen, welches für ein Insektizides Protein von *B.t.k* HD-73 codiert, mit der Sequenz:

1 ATGGCCATTGAAACCGGTTACACTCCCATCGACATCTCCT 40  
 41 TGTCCCTGACACAGTTCTGCTCAGCGAGTCGTGCCAGG 80  
 81 TGCTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGT 120  
 121 ATCTTGGTCCATCTCAATGGGATGCATTCTGGTGCAA 160  
 161 TTGAGCAGTTGATCAACCAGAGGATCGAAGAGTTGCCAG 200  
 201 GAACCAGGCCATCTCTAGGTTGGAGGGATTGAGCAATCTC 240  
 241 TACCAAATCTATGCAGAGAGCTCAGAGAGTGGGAAGCCG 280  
 281 ATCCTACTAACCCAGCTCTCCGCGAGGAATGCGTATTCA 320

5	321	ATTCAGGACATGAACAGCGCTTGACCACAGCTATCCCA	360
10	361	TTGTTCCAGTCCAGAACTACCAAGTCCCTCTTGTCGG	400
15	401	TGTACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCG	440
20	441	AGACGTTAGCGTGGTGGCAAAGGTGGGATTGATGCT	480
25	481	GCAACCATCAATAGCCGTTACAACGACCTTACTAGGCTGA	520
30	521	TTGGAAACTACACCGACCACGCTGTCGTTGGTACAACAC	560
35	561	TGGCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGG	600
40	601	ATTAGATAACACCAGTTCAAGGAGAGAATTGACCCCTACAG	640
45	641	TTTTGGACATTGTGTCTCTTCCCGAACTATGACTCCAG	680
50	681	AACCTACCCCTATCCGTACAGTGTCCCAACTTACCAAGAGAA	720
55	721	ATCTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCT	760
60	761	TCCGTTGCTGCCAACGGTATCGAAGGCTCCATCAGGAG	800
65	801	CCCACACTTGATGGACATCTTGAACAGCATAACTATCTAC	840
70	841	ACCGATGCTCACAGAGGAGTATTACTGGTCTGGACACC	880
75	881	AGATCATGGCTCTCCAGTTGGATTCAAGCGGGCCCGAGTT	920
80	921	TACCTTCCCTCTATGGAACTATGGAAACGCGCTCCA	960
85	961	CAACACGTATCGTTGCTCAACTAGGTCAAGGTGTCTACA	1000
90	1001	GAACCTTGTCTTCCACCTTGATCAGAAAGACCCCTCAATAT	1040
95	1041	CGGTATCAACAACCAGCAACTTCCGTTCTGACGGAACA	1080

5 1081 GAGTCGCCTATGAAACCTCTTCTAACTTGCCTCCGCTG 1120  
 1121 TTTACAGAAAGAGCGGAACCGTTGATTCTTGGACGAAAT 1160  
 10 1161 CCCACCCACAGAACAAACATGTGCCACCCAGGCAAGGATTC 1200  
 15 1201 TCCCCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGAT 1240  
 1241 TCAGGAAACAGTTCCGTGAGCATCATCAGAGCTCTATGTT 1280  
 20 1281 CTCTTGGATAACACCGTAGTGCTGAGTTCAACAAACATCATC 1320  
 1321 GCATCCGATAGTATTACTCAAATCCCTGCAGTGAAAGGGAA 1360  
 25 1361 ACTTTCTCTTCAACGGTTCTGTCATTCAGGACCAGGATT 1400  
 1401 CACTGGTGGAGACCTCGTTAGACTCAAACAGCAGTGGAAAT 1440  
 30 1441 AACATTCAAGAATAGAGGGTATATTGAAGTTCCAATTCACT 1480  
 1481 TCCCCATCCACATCTACCAAGATATAGAGTTCGTGTGAGGTA 1520  
 35 1521 TGCTTCTGTGACCCCTATTCAACCTCAACGTTAATTGGGGT 1560  
 1561 AATTCACTCCATCTTCTCCAATACAGTTCCAGCTACAGCTA 1600  
 40 1601 CCTCCCTGGATAATCTCCAATCCAGCGATTTCGGTTACTT 1640  
 1641 TGAAAGTGCCAATGCTTTACATCTTCACTCGGTAACATC 1680  
 45 1681 GTGGGTGTTAGAACTTTAGTGGGACTGCAGGGAGTGTATA 1720  
 50 1721 TCGACAGATTGAGTTCAATTCCAGTTACTGCAACACTCGA 1760  
 55 1761 GGCTGAG 1767.

C. einem Struktur-Gen, das für ein Insektizides Protein von B.t.k. HD-1 codiert, mit der Sequenz:

5	1	ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA	40
10	41	ACTGCTTGAGTAACCCAGAAGTGAAGTACTTGGTGGAGA	80
15	81	ACGCATTGAAACCGGTTACACTCCCATGCACATCTCCTTG	120
20	121	TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG	160
25	161	CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT	200
30	201	CTTTGGTCCATCTCAATGGGATGCCATTCTGGTGCAATT	240
35	241	GAGCAGTTGATCAACCCAGAGGATCGAAGAGTTGCCAGGA	280
40	281	ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA	320
45	321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
50	361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT	400
55	401	TCAACGACATGAACAGCGCCTTGACCACAGCTATCCCATT	440
60	441	GTTCGCAGTCCAGAACTACCAAGTTCTCTCTTGTCCGTG	480
65	481	TACGTTCAAGCAGCTAATCTCACCTCAGCGTGCCTCGAG	520
70	521	ACGTTAGCGTGTGGCAAAGGTGGGATTGCGATGCTGC	560
75	561	AACCATCAATAGCGTTACAACGACCTTACTAGGCTGATT	600
80	601	GGAAACTACACCGACCACGCTGTTGTTGGTACAACACTG	640
85	641	GCTTGGAGCGTGTCTGGGTCCCTGATTCTAGAGATTGGAT	680

5	681	TAGATACAACCAAGTTCAGGAGAGAATTGACCCCTCACAGTT	720
10	721	TTGGACATTGTGTCTCTCTTCCCGAACTATGACTCCAGAA	760
15	761	CCTACCCCTATCCGTACAGTGTCCCAACTTACCGAGAGAAAT	800
20	801	CTATACTAACCCAGTTCTGAGAACCTTCGACGGTAGCTTC	840
25	841	CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
30	881	CACACTTGATGGACATCTTGAACAGCATAACTATCACAC	920
35	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG	960
40	961	ATCATGGCCTCTCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
45	1001	CCTTTCCCTCTATGGAACATATGGAAACGCGCCCTCCACA	1040
50	1041	ACAACGTATCGTTGCTCAACTAGGTCAAGGTGTCTACAGA	1080
55	1081	ACCTTGCTTCCACCTTGACAGAAGACCCCTCAATATCG	1120
60	1121	GTATCAACAAACCAAGCAACTTCCGTTCTTGACGGAACAGA	1160
65	1161	GTTGCCCTATGGAACCTCTTCTAACCTGCCATCGCTGTT	1200
70	1201	TACAGAAAGCGGAAACCGTTGATTCCCTGGACGAAATCC	1240
75	1241	CACCAACAGAACAAACATGTGCCACCCAGGCAAGGATTCTC	1280
80	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTG	1320
85	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
90	1361	CATGGATTCATCGTAGTGTGAGTTCAACAAATATCATTCC	1400
95	1401	TTCCCTCTAAATCACCCAAATCCCATTGACCAAGTCTACT	1440

1441 AACCTTGGATCTGGAACCTCTGCGTGAAGGACCAAGGCT 1480  
 1481 TCACAGGAGGTGATATTCTTAGAAGAACCTCTGGCCA 1520  
 1521 GATTAGCACCCCTCAGAGTTAACATCACTGCACCACTTCT 1560  
 1561 CAAAGATATCGTGTAGGATTCGTTACGCATCTACCACTA 1600  
 1601 ACTTGCAATTCCACACCTCCATCGACGGAAGGCCTATCAA 1640  
 1641 TCAGGGTAACCTCTCCGCAACCATGTCAGCGGCAGCAAC 1680  
 1681 TTGCAATCCGGCAGCTTCAGAACCGTCGGTTCACTACTC 1720  
 1721 CTTTCAACTTCTCTAACGGATCAAGCGTTTCACCCCTAG 1760  
 1761 CGCTCATGTGTTCAATTCTGGCAATGAAGTGTACATTGAC 1800  
 1801 CGTATTGAGTTGTGCCCTGCCRAAGTTACCTTCGAGGCTG 1840  
 1841 AGTAC 1845.

D. einem Struktur-Gen, das für ein Insektizides Protein codiert, das von *B.t.k* HD-73 stammt, mit der Sequenz:

1 ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA 40  
 41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80  
 81 ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG 120  
 121 TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGGCAGGTG 160  
 161 CTGGGTTCGTCTCGGACTAGTTGACATCATCTGGGTAT 200

5	201	CTTGGTCCATCTCAATGGGATGCATTCTGGTGC <del>AA</del> ATT	240
10	241	GAGCAGTTGATCAACCAGAGGATCGAAGAGTTCGCCAGGA	280
15	281	ACCAAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA	320
20	321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
25	361	CCTACTAACCCAGCTCTCCGGAGGAAATGCGTATTCAAT	400
30	401	TCAACGACATGAAACAGGCCCTTGACCACAGCTATCCCATT	440
35	441	GTTCGCAGTCCAGAACTACCAAGTTCTCTTGTCCGTG	480
40	481	TACGTTCPAGCAGCTATTCTCACCTCAGCGTGCTTCGAG	520
45	521	ACGTTAGCGTGTGGCAAGGTGGGATTCGATGCTGC	560
50	561	AACCATCAATAGCCGTTACACGACCTTACTAGGCTGATT	600
55	601	GGAAACTACACCGACCACGCTGTTCGTTGGTACAACACTG	640
60	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680
65	681	TAGATACAACCAAGTTCAAGGAGAGATTGACCCCTCACAGTT	720
70	721	TTGGACATTGTTGTCCTCTTCCGAACTATGACTCCAGAA	760
75	761	CCTACCCCTATCCGTACAGTGTCCCAACTTACCAAGAGAAAT	800
80	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
85	841	CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
90	881	CACACTTGATGGACRTCTTGAAACAGCATTAACATATCTACAC	920
95	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAG	960

5	961	ATCATGGCCTCTCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
10	1001	CCTTCTCTCTATGGAACTATGGAAACCGCCCTCCACA	1040
15	1041	ACAAACGTATCGTGTCAACTAGGTAGGGTGTCTACAGA	1080
20	1081	ACCTGTCTTCCACCTGTACAGAAGACCCCTTCATATCG	1120
25	1121	GTATCAACAACCAGCAACTTTCCGTTCTGACGGAACAGA	1160
30	1161	GTTCGCCTATGAAACCTCTTCTAACCTGCCATCCGCTGTT	1200
35	1201	TACAGAAAGAGCGGAACCGTTGATTCTTGGACGAAATCC	1240
40	1241	CACCAACAGAACAAATGTGCCACCCAGGCAAGGATTCTC	1280
45	1281	CCACAGGTTGAGCCACGTTGTCATGTTCCGTTCCGGATT	1320
50	1321	AGCACACAGTTCCGTGAGCATCATCAGAGCTCTATGTTCT	1360
55	1361	CTTGGATAACCCGTAGTGCTGAGTTCAACAAACATCATCGC	1400
	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAGGGAAAC	1440
	1441	TTTCCTTCACGGTTCTGTCATTTCAGGACCAGGATTCA	1480
	1481	CTGGTGGAGACCTCGTAGACTCAACAGCAGTGGAAATAA	1520
	1521	CATTCAAAATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
	1561	CCATCCACATCTACCAAGATAATAGAGTTGCGTGTGAGGTATG	1600
	1601	CTTCTGTGACCCCTATTCAACCTCAACGTTAATTGGGGTAA	1640
	1641	TTCATCCATCTCTCCAATACAGTCCAGCTACAGCTACC	1680
	1681	TCCTTGGATAATCTCCAATCCAGCGATTTCGGTTACTTTG	1720

1721 AAAGTCCAATGCTTTACATCTTCACTCGGTAACATCGT 1760  
 1761 GGGTGTAGAAACTTGTGGACTGCAGGAGTGATTATC 1800  
 1801 GACAGATTGAGTTCAATTCCAGTTACTGCAACACTCGAGG 1840  
 1841 CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTAAATGCG 1880  
 1881 CTGTTTACGTCTACAAACAGCTGGACTCAAGACAAATG 1920  
 1921 G 1921

E. einem Struktur-Gen, das für das Insektizide Protein von *B.t.k* HD-73 in dessen gesamter Länge codiert, mit der Sequenz:

1 ATGGACAAACAACCCAAACATCAACGAATGCATTCCATACA 40  
 41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80  
 81 ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG 120  
 121 TCCTTGACACAGTTCTGCTCAGCGAGTTCTGTGCCAGGTG 160  
 161 CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT 200  
 201 CTTTGGTCCATCTCAATGGGATGCATTCTGGTGCATT 240  
 241 GAGCAGTTGATCAACCAGAGGGATCGAAGAGTTCGCCAGGA 280  
 281 ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA 320  
 321 CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT 360  
 361 CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT 400  
 401 TCAACGACATGAACAGCGCCTTGACCACAGCTATCCCATT 440

441	GTTCGCAGTCCAGAACTACCAAGTCCCTCTCTTGTCCGTG	480
481	TACCTTCAGCAGCTAACTTTCACCTCAGCGTGCTTCGAG	520
521	ACGTTAGCGTGTGGGCAAAGGTGGGATTGATGCTGC	560
561	AACCATCAATAGCGTTACAACGACCTTACTAGGCTGATT	600
601	GGAAACTACACCGACCACGCTGTTGTTGTAACACTG	640
641	GCTGGAGCGTGTCTGGGTCCCTGATTCTAGAAGATTGGAT	680
681	TAGATACAACCAAGTCAGGAGAGAATTGACCCCTCACAGTT	720
721	TTGGACATTGTGTCTCTTCCCGAACTATGACTCCAGAA	760
761	CCTACCCCTATCCGTACAGTGTCCCACCTTACCAAGAGAAAT	800
801	CTATACAAACCCAGTTCTTGAGAACCTCGACGGTAGCTTC	840
841	CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
881	CACACTTGATGGACATCTGAACAGCATAACTATCTACAC	920
921	CGATGCTCACAGAGGGAGAGTATTACTGGTCTGGACACCAAG	960
961	ATCATGGCCTCTCAGTTGGATTCAAGCGGGCCCCAGTTTA	1000
1001	CCTTCCTCTATGAACTATGGGAAACGCCGCTCCACA	1040
1041	ACAAACGTATCGTTGTCACACTAGGTCAAGGGTGCTACAGA	1080
1081	ACCTTGTCTCCACCTGTACAGAAGACCCCTCAATATCG	1120
1121	GTATCAACAACCGAACCTTCCGTTCTGACGGAACAGA	1160
1161	GTTGCCCTATGGAACCTCTTAACCTGGCATCCGCTGTT	1200

5	1201	TACAGAAAAGAGCGGAACCGTTGATTCTTGGACGAAATCC	1240
10	1241	CAACACAGAACAAATGTGCCACCCAGGCAAGGATTCTC	1280
15	1281	CCACAGGTTGAGCCACGTCCTCATGTTCCGTTCCGGATT	1320
20	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
25	1361	CTTGGATAACCCGTAGTGTGAGTTCAACAAACATCATCGC	1400
30	1401	ATCCGATACTTACTCAAAATCCCTGCAGTGAAGGGAAAC	1440
35	1441	TTTCTCTCAACGGTCTGTCAATTCAAGGACCAGGATTCA	1480
40	1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
45	1521	CATTAGAATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
50	1561	CCATCCACATCTACCAAGATAAGACTTCGTGTGAGGTATG	1600
55	1601	CTTCTGTGACCCCTATTCAACCTCAACGTTAATTGGGGTAA	1640
60	1641	TTCACTCATCTTCTCCAATACAGTCCAGCTACACCTACC	1680
65	1681	TCCTGGATAATCTCCAATCCAGCCATTCCGGTTACTTTG	1720
70	1721	AAAGTCCAATGCTTTACATCTTCACTCGTAACATCGT	1760
75	1761	GGGTGTTAGAAACTTAGTGGACTGCAGGAGTGATTATC	1800
80	1801	GACAGATTGAGTTCACTCCAGTTACTGCAACACTCGAGG	1840
85	1841	CTGAATATAATCTGGAAAGAGCGCGAGAAGGCGGTGAATGC	1880
90	1881	GCTGTTACGTCTACAAACCAAGCTCGGCCTCAAGACCAAT	1920
95	1921	GTGACGGATTATCATATTGATCAAGTGTCCAACCTGGTGA	1960

5	1961	CCTACCTCAGCGATGAGTTCTGTCTGGATGAAAAGCGAGA	2000
10	2001	ATTGTCCGAGAAAAGTCAAACATCGGAAGCGACTCAGTGAT	2040
15	2041	GAACCGCAATTACTCCAAGATTCAAATTCAAAGACATTA	2080
20	2081	ATAGGCACCCAGAACCGTGGTGGGCGGAAAGTACAGGGAT	2120
25	2121	TACCATCCAGGGAGGTGACCGACGTGTTCAAGGAGAACTAC	2160
30	2161	GTCACACTATCAGGTACCTTGATGAGTGCTATCCAACAT	2200
35	2201	ACCTCTACCAGAAGATCGACGAGTCCAAGTTGAAAGCCTT	2240
40	2241	TACCCGTTATCAATTAAAGGGGTATATCGAAGATAAGTCAA	2280
45	2281	GACCTCGAGATCTACCTCATCCGCTACAATGCAAAACATG	2320
50	2321	AAACAGTAAATGTGCCAGGTACGGTTCCCTTATGCCGCT	2360
55	2361	TTCAGCCCCAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400
60	2401	CGATGCGCGCCACACCTTGAATGGAATCCTGACTTAGATT	2440
65	2441	GTTCGTGTAGGGATGGAGAAAAGTGTGCCCATCTTCGCA	2480
70	2481	TCATTTCTCCTTAGACATTGATGTAGGATGTACAGACTTA	2520
75	2521	AATGAGGACCTAGGTGTATGGGTGATCTTAAGATTAAGA	2560
80	2561	CGCAAGATGGGCACGCAAGACTAGGGAATCTAGAGTTCT	2600
85	2601	CGAAGAGAAAACCATTAGTAGGAGAAGCGCTAGCTCGTGTG	2640
90	2641	AAAAGAGCGGAGAAAAATGGAGAGACAAACGTGAGAAAGT	2680
95	2681	TGGAATGGGAGACCAACATCGTCTACAAAGAGGCAAAAGA	2720

2721	ATCTGTAGATGCTTATTGTAACTCTCAATATGATCAA	2760
2761	TTACAAGCGGATACGAATTGCCCCATGATTCAATGGGGCAG	2800
2801	ATAAAACGTGTTCATAGCATTGAGAAGCTTATCTGCCTGA	2840
2841	GCTGTCTGTGATTCCGGGTGTCAATGGGGCTATTTTGAA	2880
2881	GAATTAGAAGGGCGTATTTCACTGCATTCTCCCTCTACG	2920
2921	ATGCCAGAAAACGTCATCAAGAACGGTGACTTCACAAATGG	2960
2961	CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA	3000
3001	GAACAAAACAACCAACCGTCCGGTCTTGTGTTCCGGAAT	3040
3041	GGGAAGCAGAAGTGTACAAGAACGTTCTGTCTGTCGGG	3080
3081	TCGTGGCTATATCCTTCGTGTACACCGCTACAGGAGGGA	3120
3121	TATGGAGAAGGTTGCGTAACCATTGAGATCGAGAACCA	3160
3161	ATACAGACGAACGTGAACTTGTGAACTGCGTAGAACAGAGGA	3200
3201	AATCTATCCAAATAACACGGTAACCGTGTAAATGATTATAC	3240
3241	GTAAATCAAGAAGAACCGGAGGTGCGTACACTTCGTGTA	3280
3281	ATCGAGGATATAACGAAGCTCCTTCGTACCGCTGATTA	3320
3321	TGCGTCAGTCTATGAAAGAAAATCGTATACAGATGGACGA	3360
3361	AGAGAGAACCTTGTGAATTAAACAGAGGGTATAGGGATT	3400
3401	ACACGCCACTACCGAGTTGGTTATGTGACAAAAGAATTAGA	3440
3441	ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480

3481 GAAACGGAAGGAACATTTATCGTGGACAGCGTGGATTAC 3520

5 3521 TCCTTATGGAGGAA 3534.

10 F. einem Struktur-Gen, das für ein Insektizides Protein von *B.t.k.* HD-73 in dessen gesamter Länge codiert, mit der Sequenz:

15 1 ATGGACAAACAACCCAAACATCAACGAATGCAATTCCATACA 40

20 41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80

25 81 ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCCTG 120

30 121 TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG 160

35 161 CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT 200

40 201 CTTTGGTCCATCTCAATGGGATGCATTCTGGTGCAAATT 240

45 241 GAGCAGTTGATCAACCAGAGGATCGAAGAGTTGCCAGGA 280

50 281 ACCAGGCCATCTCTAGGTTGAAAGGATTGAGCAATCTCTA 320

55 321 CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT 360

361 CCTACTAACCCAGCTCTCCGGAGGGAAATGCGTATTCAAT 400

401 TCAACGACATGAACAGCGCCTTGACCACAGCTATCCCATT 440

441 GTTCGCAGTCCAGAACTACCAAGTTCCCTCTTGTCCGTG 480

481 TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG 520

55 521 ACGTTAGCGTGTGTTGGGCAAAGGTGGGATTGATGCTGC 560

5	561	AACCATCAATAGCCGTTACAACGACCTTACTAGGGCTGATT	600
10	601	GGAAACTACACCGACCACGCTGTTGGTGGTACAACACTG	640
15	641	GCTTGGAGCGTGTCTGGGTCCCTGATTCTAGAGATTGGAT	680
20	681	TAGATACAACCAAGCTTCAAGGAGAGAAATTGACCCCTCACAGTT	720
25	721	TTGGACATTGTGTCTCTTCCCGAACTATGACTCCAGAA	760
30	761	CCTACCCCTATCCGTACAGTGCCCCAACTTACCAAGAGAAAT	800
35	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
40	841	CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
45	881	CACACTTGATGGACATCTTGAACACGATAACTATCTACAC	920
50	921	CGATGCTCACAGAGGAGAGTATTACTGGCTGGACACCAAG	960
55	961	ATCATGGCCTCTCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
60	1001	CCTTCCTCTATGGAACTATGGAAACCGCCGCTCCACA	1040
65	1041	ACAACGTATCGTTGCTCAACTAGGTCAAGGGTGTCTACAGA	1080
70	1081	ACCTGTCTTCCACCTTGTACAGAAGACCCCTCAATATCG	1120
75	1121	GTATCAACAACCAACCAACTTCCGTTCTGACGGAACAGA	1160
80	1161	GTTGCCCTATGGAACCTCTTCTAACTTGCCTCCGCTGTT	1200
85	1201	TACAGAAAGAGCGGAACCGTTGATTCCTGGACGAAATCC	1240
90	1241	CACCAACAGAACAAATGTGCCACCCAGGCAAGGATTCTC	1280
95	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTC	1320

5	1321	AGCAACAGTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
10	1361	CTTGGATACACCGTAGTGTGAGTCAACAACATCATCGC	1400
15	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
20	1441	TTTCTCTCAACGGTCTGTCAATTCAAGGACCAGGATTCA	1480
25	1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
30	1521	CATTCAAGAATAGAGGGTATATTGAAGTCCAATTCACTTC	1560
35	1561	CCATCCACATCTACCAAGATATAGAGTTCGTGTGAGGTATG	1600
40	1601	CTTCTGTGACCCCTATTACACCTCAACGTTAATTGGGTAA	1640
45	1641	TTCATCCATCTCTCCAATACAGTCCAGCTACAGCTACC	1680
50	1681	TCCTTGGATAATCTCAATCCAGCGATTTCGGTTACTTTG	1720
55	1721	AAAGTGCCAATGCTTTACATCTTCACTCGGTAAACATCGT	1760
60	1761	GGGTGTTAGAAACTTTAGTGGACTCAGGAGTGATTATC	1800
65	1801	GACAGATTGAGTTCAATTCCAGTTACTGCACACTCGAGG	1840
70	1841	CTGAATATAATCTGGAAGAGCGCAGAAGGGGTGAATGC	1880
75	1881	GCTGTTACGTCTACAAACCAACTAGGGCTAAAACAAAT	1920
80	1921	GTAACGGATTATCATATTGATCAAGTGTCCAATTAGTTA	1960
85	1961	CGTATTTATCGGATGAATTTGTCTGGATGAAAAGCGAGA	2000
90	2001	ATTGTCCGAGAAAGTCACATGCCAAGCGACTCAGTGAT	2040
95	2041	GAACGCAATTACTCCAAGATTCAAATTCAAAGACATTA	2080

5	2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120
10	2121	TACCATCCAAGGAGGGGATGACGTATTTAAGAATAATTAC	2160
15	2161	GTCACACTATCAGGTACCTTGATGAGTGCTATCCAACAT	2200
20	2201	ATTTGTATCAAAAATCGATGAATCAAATTAAAGCCCTT	2240
25	2241	TACCCGTTATCAATTAAGAGGGTATATCGAAGATAGTCAA	2280
30	2281	GACTTAGAAAATCTATTTAATTCGCTACAAATGCAAAACATG	2320
35	2321	AAACAGTAAATGTGCCAGGTACGGGTTCCCTATGCCGCT	2360
40	2361	TTCAGCCCAAAGTCCAATCGAAAGTGTGGAGAGCCGAAT	2400
45	2401	CGATGCGCGCACCTTGAATGGAATCCTGACTTAGATT	2440
50	2441	GTTCGTGTAGGGATGGAGAAAATGTGCCCATCTCGCA	2480
55	2481	TCATTTCTCTTAGACATTGATGTAGGATGTACAGACTTA	2520
	2521	AATGAGGACCTAGGTGTATGGGTGATCTTAAGATTAAGA	2560
	2561	CGCAAGATGGGCACCCAAGACTAGGGAAATCTAGAGTTCT	2600
	2601	CGAAGAGAAACCATTAGTAGGAGAACCGCTAGCTCGTGTG	2640
	2641	AAAAGAGCGGAGAAAAATGGAGAGACAAACGTGAAAAT	2680
	2681	TGGAATGGGAAACAAATATCGTTTAAAGAGGGCAAAAGA	2720
	2721	ATCTGTAGATGCTTTATTTGTAACCTCTCAATATGATCAA	2760
	2761	TTACAAGCGGATACGAATATTGCCATGATTGATGCCAG	2800
	2801	ATAAACGTGTCATAGCATTGAGAGCTTATCTGCCCTGA	2840

5 2841 GCTGCTGTGATTCCGGGTGTCAATGCGGCTATTTTGAA 2880  
 10 2881 GAATTAGAAGGGCGTATTTCACTGCATTCTCCCTATATG 2920  
 15 2921 ATGCGAGAAATGTCATTAAAAATGGTGATTTAATAATGG 2960  
 20 2961 CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA 3000  
 25 3001 GAACAAAACAACCAACGTTCGGTCTTGTGTTCCGGAAAT 3040  
 30 3041 GGGAAAGCAGAAGTGTACAAGAAGTTCGTGTCTGTCGGG 3080  
 35 3081 TCGTGGCTATATCCTCGTGTACAGCGTACAAGGAGGGA 3120  
 40 3121 TATGGAGAAGGTTGCGTAACCATTATGAGATCGAGAACAA 3160  
 45 3161 ATACAGACGAACGTAAAGTTAGCAACTGCGTAGAAGAGGA 3200  
 50 3201 AATCTATCCAAAATAACACGGTAACGTGTAAATGATTATACT 3240  
 55 3241 GTAAATCAAGAAGAATACGGAGGTGCGTACACTTCGTA 3280  
 60 3281 ATCGAGGATATAACGAAGCTCCTCCGTACCGCTGATTA 3320  
 65 3321 TGCCTCAGTCTATGAAGAAAATCGTATACAGATGGACGA 3360  
 70 3361 AGAGAGAATCCTGTGAATTAAACAGAGGGTATAGGGATT 3400  
 75 3401 ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA 3440  
 80 3441 ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA 3480  
 85 3481 GAAACGGAAGGAACATTATCGTGGACAGCGTGGAAATTAC 3520  
 90 3521 TCCTTATGGAGGAA 3534,

G. einem Struktur-Gen, das für ein Insektizides Protein von *B.t.k* HD-73 in dessen gesamter Länge codiert, mit der Sequenz:

5	1	ATGGACAACAAACCCAAACATCAACGAATGCATTCCATACA	40
10	41	ACTGCTTGACTAACCCAGAAGTTGAAGTACTTGGTGGAGA	80
15	81	ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG	120
20	121	TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGGCAGGTG	160
25	161	CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT	200
30	201	CTTTGGTCCATCTCAATGGGATGCAATTCTGGTGCAAATT	240
35	241	GAGCAGTTGATCAACCAACAGAGGATCGAAGAGTTCCCAAGGA	280
40	281	ACCAAGGCCATCTCTAGGTTGAAAGGATTGAGCAATCTCTA	320
45	321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
50	361	CCTACTAACCCAGCTCTCGCGAGGAAATGCGTATTCAAT	400
55	401	TCAACGACATGAACAGCGCTTGACCACAGCTATCCCATT	440
60	441	GTTCGCAGTCCAGAACTACCAAGTTCTCTCTTGCCGTG	480
65	481	TACGTTCAAGCAGCTAACCTTCACCTCAGCGTGCCTCGAG	520
70	521	ACGTTAGCGTGTGGCAAAGGTGGGATTGCGATGCTGC	560
75	561	AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600
80	601	GGAAACTACACCGACCAACGCTGTTGTTGGTACAACACTG	640
85	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680

5	681	TAGATACACCAGTTCAAGGAGAGAATTGACCCCTCACAGTT	720
10	721	TTGGACATTGTCTCTCTTCCCGAAGCTATGACTCCAGAA	760
15	761	CCTACCCCTATCCGTACAGTGTCCCAACTTACCAAGAGAAAT	800
20	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
25	841	CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
30	881	CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC	920
35	921	CGATGCTCACAGAGGGAGAGTATTACTGGTCTGGACACCAG	960
40	961	ATCATGGCCCTCTCCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
45	1001	CCTTCCCTCTATGGAACATATGGAAACGCCGCTCCACA	1040
50	1041	ACAACGTTATCGTTGCTCAACTAGGTCAAGGGTCTACAGA	1080
55	1081	ACCTTGTCTCCACCTTGACAGAAGACCCCTTCATATCG	1120
60	1121	GTATCAACAACCAGCAACTTCCGTTCTGACGGAACAGA	1160
65	1161	GTTGCCCTATGGAACCTCTTCTAATTGCCATCCGCTGTT	1200
70	1201	TACAGAAAGAGCGGAACCGTTGATTCTGGACGAAATCC	1240
75	1241	CACACAGAACAAACATGTGCCACCCAGGCAAGGATTCTC	1280
80	1281	CCACAGGTTGAGCCACGTTGTCATGTTCCGTTCCGGATTC	1320
85	1321	AGCAACAGTCCGTGACCATCATCAGAGCTCTATGTTCT	1360
90	1361	CTTGGATAACCCGTAGTGCTGAGTTAACAAACATCATCGC	1400
95	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440

5	1441	TTTCCTTCACGGTCTGTCATTCAGGACCAGGATTCA	1480
10	1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
15	1521	CATTCAAGAATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
20	1561	CCATCCACATCTACCAAGATATAGAGTTCTGTGAGGTATG	1600
25	1601	CTTCGTGACCCCTATTCACTCAACGTTAATTGGGGTAA	1640
30	1641	TTCATCCATCTCTCAATACAGTCCAGCTACAGCTACC	1680
35	1681	TCCTTGGATAATCTCCAATCCAGCGATTCTGGTTACTTG	1720
40	1721	AAAGTCCAATGCTTTACATCTCACTCGGTAACATCGT	1760
45	1761	GGGTGTTAGAAACTTAGTGGGACTGCAGGAGTGATTATC	1800
50	1801	GACAGATTGAGTTCAITCCAGTTACTGCAACACTCGAGG	1840
55	1841	CTGAGTACAACCTTGAGAGAGGCCAGAAGGCTGTGAACGC	1880
60	1881	CCTCTTACCTCCACCAATCAGCTGGCTTGAAAACTAAC	1920
65	1921	GTTACTGACTATCACATTGACCAAGTGTCCAACTTGGTCA	1960
70	1961	CCTACCTTAGCGATGAGTTCTGCCTCGACGAGAACGTGA	2000
75	2001	ACTCTCCGAGAAAGTTAACACGCCAACCGTCTCAGCGAC	2040
80	2041	GAGAGGAATCTCTGCAAGACTCCAACITCAAAGACATCA	2080
85	2081	ACAGGCAGCCAGAACGTGGTTGGGTGGAAGCACCGGGAT	2120
90	2121	CACCATCCAAGGAGGGCAGCATGTTCAAGGGAGAACTAC	2160
95	2161	GTCACCCCTCTCCGGAACCTTCGACGAGTGTACCCCTACCT	2200

5

2201	ACTTGTACCAGAAGATCGATGAGTCCAACTCAAAGCCTT	2240
2241	CACCAAGGTATCAACTTAGAGGCTACATCGAAGACAGCCAA	2280
2281	GACCTTGAAATCTACTCGATCAGGTACAAATGCCAAGCAGC	2320
2321	AGACCGTGAATGTCCCAGGTACTGGTCCCTCTGCCACT	2360
2361	TTCTGCCAATCTCCATTGGGAAGTGTGGAGAGCCCTAAC	2400
2401	AGATGCGCTCCACACCTTGAGTGGAAATCCTGACTTGGACT	2440
2441	GCTCCTGCAGGGATGGCGAGAAGTGTGCCACCATTCTCA	2480
2481	TCACTTCTCCTTGGACATCGATGGGATGTACTGACCTG	2520
2521	AATGAGGACCTCGGAGTCTGGTCATCTTCAGATCAAGA	2560
2561	CCCAAGACGGACACCCAAGACTTGGCAACCTTGAGTTCT	2600
2601	CGAAGAGAAACCATTGGCGGTGAAGCTCTCGCTCGTGTG	2640
2641	AAGAGAGCAGAGAAGAAGTGGAGGGACAAACGTGAGAAC	2680
2681	TCGAATGGGAAACTAACATCGTTACAAGGAGGCCAAGA	2720
2721	GTCCCTGGATGCTTGTTCGTGAACCTCCAAATATGATCAG	2760
2761	TTGCAAGCCGACACCAACATGCCATGATCCACGCCAG	2800
2801	ACAAACGTGTGCACAGCATTCTGAGGCTTACTTGCCTGA	2840
2841	GTTGTCCTGATCCCTGGTGTGAACGCTGCCATCTCGAG	2880
2881	GAACCTGAGGGACCTATCTTACCGCATTCTCCTGTACG	2920
2921	ATGCCAGAACGTCTACAGAACGGTGACTTCAACAATGG	2960

2961 CCTCAGCTGCTGGATGTGAAAGGTATGTGGACGTGGAG 3000  
 3001 GAACAGAACAAATCAGCGTCCGTCCTGGTTGTGCCCTGAGT 3040  
 3041 GGGAAAGCTGAAGTGTCCCAAGAGGTTAGAGTCTGTCCAGG 3080  
 3081 TAGAGGCTACATTCTCCGTGTGACCCCTTACAAGGAGGGA 3120  
 3121 TACGGTGAGGGTTGCGTGACCATCCACGAGATCGAGAACAA 3160  
 3161 ACACCGACGAGCTTAAGTTCTCAACTGCGTCGAGGAAGA 3200  
 3201 AATCTATCCAAACACACCGTTACTTGCAACGACTACACT 3240  
 3241 GTGAATCAGGAAGAGTACGGAGGTGCCTACACTAGCCGTA 3280  
 3281 ACAGAGGTTACAACGAAGCTCCTCCGTTCTGCTGACTA 3320  
 3321 TGCCTCCGTGTACGAGGGAGAAATCCTACACAGATGGCAGA 3360  
 3361 CGTGAGAACCCCTTGGAGTTCAACAGAGGTTACAGGGACT 3400  
 3401 ACACACCACCTCCAGTTGGCTATGTTACCAAGGAGCTTGA 3440  
 3441 GTACTTTCTGAGACCGACAAAGTGTGGATCGAGATCGGT 3480  
 3481 GAAACCGAGGGAACCTTCATCGTGGACAGCGTGGAGCTTC 3520  
 3521 TCTTGATGGAGGAA 3534

50 H. einem Struktur-Gen, das für ein Insektizides Protein von *B.t.t.* codiert, mit der Sequenz:

55 1 ATGACTGCAGACAAACAACACCGAACGCCCTCGACAGTTCTA 40  
 41 CCACTAAGGATGTATCCAGAAGGGTATCTCCGTTGTGGG 80

81	AGACCTCTGGCGTGGTTGGATTCCTCGGTGGAGCC	120
121	CTCGTGAGCTCTATACAAACTTCTCAACACCAATTGGC	160
161	CAAGCGAGGACCCCTGGAAAGCATTCATGGAGCAAGTTGA	200
201	AGCTCTTATGGATCAGAAGATTGCAGATTATGCCAAGAAC	240
241	AAGGCTTGGCAGAACTCCAGGGCCTTCAGARCAAATGTGG	280
281	AGGACTACGTGAGTGCATTGTCAGCTGGCAGAAGAACCC	320
321	TGTTAGCTCCAGAAATCCTCACAGCCAAGGTAGGATCAGA	360
361	GAGTTGTTCTCAAGCCGAATCCACTTCAGAAATTCCA	400
401	TGCCCTAGCTTGCTATCTCCGGTTACGAGGTTCTTTCTC	440
441	CACTACCTATGCTCAAGCTGCCAACACCCACTTGTCTC	480
481	CTTAAGGACGCTAAATCTATGGAGAAGAGTGGGGATAAG	520
521	AGAAAGAGGACATTGCTGAGTTCTACAAGCGTCAACTTAA	560
561	GCTCACCCAAGAGTACACTGACCATTGCGTGAAATGGTAT	600
601	AACTGGTCTCGATAAGCTCAGAGGCTCTTCTACGAGT	640
641	CTTGGGTGAACTCAACAGATAACAGGAGAGAGATGACCTT	680
681	GACTGTGCTCGATCTTATCGCACTCTTCCCTGTACGAT	720
721	GTGAGACTCTACCCAAAGGAAGTGAAAAGTGAGCTTACCA	760
761	GAGACGTGCTCACTGACCCATTGCGGAGTCAACAAACCT	800
801	TAGGGTTATGAACTACCTTCAGCAATATCGAAAACCTAC	840

841	ATTAGGAAACCACATCTCTTCGACTATCTTCACAGAAATC	880
881	AATTCCACACAAGGTTCAACCAGGATACTATGGTAACGA	920
921	CTCCTCACTATTGGTCCGGTAACTATGTTCCACCAGA	960
961	CCAAGGATGGATCTAATGACATCATCACATCTCCCTTCT	1000
1001	ATGGTAACAAGTCCAGTGAACCTGTGCAGAACCTTGAGTT	1040
1041	CAACGGCAGAAAGTCTATAGGCCGTCGCAAAACACCAAT	1080
1081	CTCGCTGTGTGGCATCCGCAAGTTACTCAGGCCTCACAA	1120
1121	AGGTGGAGTTAGTCAGTATAACGATCAGACCGATGAGGC	1160
1161	CAGCACCCAGACTTACGACTCCAACGTAACGTTGGCGCA	1200
1201	GTCTCTGGGATTCTATCGACCAATTGCTCCAGAAACCA	1240
1241	CAGACGAACCATTGGAGAAGGGCTACAGCCACCAACTAA	1280
1281	CTATGTGATGTCTTCTTGATGCAAGGTTCCAGAGGGACC	1320
1321	ATTCAGTGTGACCTGGACACACAAGTCCGTGGACTTCT	1360
1361	TCAACATGATCGATAGCAAGAAGATCACTCAACTTCCCTT	1400
1401	GGTGAAAGCCTACAAGCTGCAATCTGGTGTCCGTTGTC	1440
1441	GCAGGTCCCAGATTCACTGGAGGTGACATCATCCAGTGCA	1480
1481	CAGAGAACGGCAGCGCAGCTACTATCTACGTGACACCTGA	1520
1521	TGTGTCTTACTCTCAGAAGTACAGGGCACGTATTCAATTAC	1560
1561	GCATCTACCAGCCAGATCACCTTACACTCAGCTTGGATG	1600

5 1601 GAGCACCCCTCAACCAGTATTACTTGTACAAGACCATCAA 1640  
 1641 CAAAGGTGACACTCTCACATACAATAGCTTCAGGGCA 1680  
 10 1681 AGTTTCAGCACACCATTGAACTCTCAGGCAACAATCTTC 1720  
 15 1721 AGATCGGCGTCACCGGCTCAGCGCCGGAGACAAAGTCTA 1760  
 1761 CATCGACAAGATTGAGTTCATCCCAGTGAAC 1791

20 I. einem Struktur-Gen, das für ein Insektizides Protein von *B. t. entomocidus* codiert, mit der Sequenz:

25 1 ATGGAGGAGAACACCAAAACCAATGCATTCCATACAACT 40  
 41 GCTTGAGTAACCCAGAAGAGGTATTGCTTGATGGAGAACG 80  
 30 81 CATTTCACCGGTAACTCTTCCATCGACATCTCCTTGTCC 120  
 121 TTGGTCCAGTTCTGGTCAGCAACTTGTGCCAGGTGGTG 160  
 35 161 GGTTCTTGTGGACTAATTGACTTCGTCTGGGTATCGT 200  
 201 TGGTCCATCTCAATGGGATGCATTCTGGTGCAAATTGAG 240  
 40 241 CAGTTGATCAACGAGAGGATCGCTGAGTCGCCAGGAACG 280  
 45 281 CTGCCATCGCTAACCTGGAAGGATTGGCAATAACTTCAA 320  
 321 CATCTATGTGGAGGCCTTCAAAGAGTGGGAAGAGGACCT 360  
 50 361 AACAAACCCAGAGACCCGCACTAGGGTGTGACAGATTCA 400  
 401 GAATCTTGGACGGCCTCTTGGAGAGAGATATCCCATCCTT 440  
 55 441 CAGAATCTCTGGCTTCGAAGTTCCCTCTTGTCCGTGTAC 480

481	GCTCAAGCAGCTAATCTCACCTCGTATCCTCGAGACA	520
521	GTGTCATCTTGGGAAAGGTGGGATTGACCACTATCAA	560
561	CGTCAATGAGAATTACAACAGACTTATCAGGCACATTGAC	600
601	GAGTACGCCGACCACTGTGCTAACACCTACAAACCTGGCT	640
641	TGAACAATCTCCCTAAGTCTACTTATCAAGATTGGATTAC	680
681	CTACAAACAGGTTGAGGGAGAGACTTGACCCCTCACAGTTTG	720
721	GACATTGCAGCTTCTTCCCGAACTATGACAACAGGAGAT	760
761	ACCCATCCAACCACTGGGTCAACTTACCAAGAGAAGTCTA	800
801	TACTGACCCACTTATCAACTTCAACCCCTCAGTTGCAAAGT	840
841	GTCGCCAACCTCCCACATTCAACGTCAATGGAGTCCAGCC	880
881	GTATCAGGAACCCACACTTGGTTGACATCTTGAACACCT	920
921	TACTATCTTACCGATTGGTTCAAGCGTTGGCGTAACCTC	960
961	TATTGGGTGGACACAGGGTCATCTCCTCTCTTATTGGAG	1000
1001	GTGGGAACATTACCTCTCTATCTATGGACGTGAGGCAA	1040
1041	CCAGGAGCCACCACGTAGTTCACCTCAACGGTCCAGTC	1080
1081	TTCAGAACCTTGTCTAACCTTACCTTGAGATTGCTCCAGC	1120
1121	AACCTTGGCCAGCTCCACCTTCAACCTTAGAGGTGTTGA	1160
1161	GGGCAGTTGAGTTCTCTACCTCACCAACTCCTCACTTAC	1200
1201	AGAGGTAGAGGAACCGTTGATTCTTGAACCGAACCTCCAC	1240

5	1241	CAGAGGACAATAGCGTGCCACCCAGGGAGGGCTACTCCCA	1280
10	1281	CAGGTGTCGCCACGCAACCTCGTGCAGCGTCCCGGAACCT	1320
15	1321	CCATTCCTCACTACAGGAGTTGTGTTCTCATGGACTGATC	1360
20	1361	GTAGTGTACTCTCACTAACATTGATCCCGAGAGGAT	1400
25	1401	CAATCAAATCCCATTGGTCAAGGGTTCCGTGTGTTGGGA	1440
30	1441	GGAACCTCTGTCACTCACAGGACCAGGGCTCACAGGAGGTG	1480
35	1481	ATATTCTTAGAAGAAAACACTTTGGCGACTTGTGAGCCT	1520
40	1521	CCAAGTTAACATCAACTCTCAATTACTCAAAGATATCGT	1560
45	1561	CTCAGGTTCTGTTACGCATCTCCCGTACGCTAGAGTCA	1600
50	1601	TCGTGCTCACCGGAGCAGCTCTACCGGTGTCGGTGGACA	1640
55	1641	AGTCTCCGTGAACATGCCACTCCAGAAGACTATGGAGATC	1680
60	1681	GGCGAGAACTTGACATCCAGGACCTTCAGATACACCCACT	1720
65	1721	TCTCTAACCCCTTCAGTTCCGTGCCAACCTGACATCAT	1760
70	1761	TGGCATTAGCGAACACCTCTCTTGGAGCTGGTAGCATC	1800
75	1801	TCATCTGGCGAATTGACATTGACAAGATTGAGATCATTC	1840
80	1841	TTGCCGACGCTACCTCGAGGCTGAGTCTGACCTTGAGAG	1880
85	1881	AGCCCAGAAGGCTGTGAAACGCCCTTTACCTCTTAAT	1920
90	1921	CAGATTGGCTTGAAACTGACGTTACTGACTATCACATTG	1960
95	1961	ACCAAGTGTCCAACCTGGTCGACTGCCCTAGCGATGAGTT	2000

5	2001	CTGCCCTGACGAGAAGCGTGAACCTCTCGAGAAAGTTAAA	2040
10	2041	CACGCCAAGCGTCTCAGCGACGAGAGGAATCTCTTCAAG	2080
15	2081	ACCCCAACTTCAGAGGCATCAACAGGCAGCCAGACCGTGG	2120
20	2121	TTGGAGAGGAAGCACCGACATCACCATCCAAGGAGGGCAG	2160
25	2161	GATGTGTTCAAGGAGAACTACGTCACCCCTCCAGGAACGTG	2200
30	2201	TGGACGAGTGCTACCCCTACCTACTTGTACCGAGAAGATCGA	2240
35	2241	TGAGTCAAAATCTAAAGCCTACACCAGGTATGAACCTTAA	2280
40	2281	GGCTACATCGAAGACAGCCAAGACCTTGAAATCTACCTCA	2320
45	2321	TCAGGTACAATGCCAACGACAGATCGTGAATGTCCCAGG	2360
50	2361	TACTGGTCCCTCTGCCACTTTCTGCCAAATGCCATT	2400
55	2401	GGGAAGTGTGGAGAGCCTAACAGATGCCCTCACACCTTG	2440
60	2441	AGTGGAAATCTGACTTGGACTGCTCTGCAGGGATGGCGA	2480
65	2481	GAAGTGTGCCACCATTCTCATCACTTCACCTTGGACATC	2520
70	2521	GATGTGGATGTACTGACCTGAATGAGGACCTCGGAGTCT	2560
75	2561	GGGTCACTTCAGATCAAGACCCAAGACGGACACCGCAAG	2600
80	2601	ACTTGGCAACCTTGAGTTCTCGAAGAGAAACCATTGCTC	2640
85	2641	GGTGAAGCTCTCGCTCGTGTGAAGAGAGCAGAGAAGAGT	2680
90	2681	GGAGGGACAAACGTGAGAAAACCTCAACTCGAGACTAACAT	2720
95	2721	CGTTTACAAGGAGGCCAAAGAGTCCGTGGATGCTTGTTC	2760

5	2761	GTGAACTCCAAATATGATAGTTGCAAGTGGACACCAACA	2800
10	2801	TCCGCCATGATCCACCGCTGCAGACAAACGTGTGCAACAGGAT	2840
15	2841	TCGTGAGGCTTACTTGCCTGAGTTGTCCTGATCCCTGGT	2880
20	2881	GTGAAACGCTGCCATCTTCGAGGAACTTGAGGGACGTATCT	2920
25	2921	TTACCGCATACTCCTGTACCGATGCCAGAACGTCATCAA	2960
30	2961	GAACGGTGACTTCACAAATGGCCTCTGTGCTGGAATGTG	3000
35	3001	AAAGGTCACTGGACCTGGAGGAACAGAACAAATCACCGTT	3040
40	3041	CCGTCTGGTTATCCCTGAGTGGGAAGCTGAAGTGTCCCC	3080
45	3081	AGAGGTTAGAGTCTGTCCAGGTAGAGGCTACATTCTCCGT	3120
50	3121	GTGACCGCTTACAAGGAGGGATAACGGTGAGGGTTGCGTGA	3160
55	3161	CCATCCACGAGATCGAGGACAAACACCGACGAGCTTAAGTT	3200
60	3201	CTCCAACCTCGCTCGAGGAAGAAGTCTATCCAAACACACC	3240
65	3241	GTTACTTGCAACAACACTACACTGGGACCCAGGAAGAGTACG	3280
70	3281	AAGGTACCTACACTACCGTAACCAAGGTTACGACGAAGC	3320
75	3321	TTACGGAAACAATCCTCCGTTCTGCTGACTATGCCCTCC	3360
80	3361	GTGTACGAGGAGAAATCCTACACAGATGGCAGACGTGAGA	3400
85	3401	ACCCCTGCGAGTCCAACAGAGGTTACGGTGACTACACACC	3440
90	3441	ACTTCCAGCAGGCTATGTTACCAAGGGACCTTGAGTACTTT	3480
95	3481	CCTGAGACCGACAAAGTGTGGATCGAGATCGGTGAAACCG	3520

3521 AGGGAACCTTCATCGTGGACAGCGTGGAGCTCTCTTGAT 3560

5 3561 GGAGGAA 3567

10 J. einem Struktur-Gen, das für ein Insektizides P2-Protein codiert, mit der Sequenz:

15 1 ATGGACAAACAGTCTTGAACCTCTGGTAGAACAAACCATCT 40

20 41 GCGACGCATAACAGTCGTGGCTACGATCCATTCAAGCTT 80

25 81 CGAACACAAGAGCCTCGACACTATTCAAGAAGGAGTGGATG 120

121 GAATGGAAACGTACTGACCCTCTCTACGTCGCACCTG 160

161 TGGTTGGAACAGTGTCCAGCTTCTCTCAAGAAGGTCGG 200

30 201 CTCTCTCATCGGAAAACGTATCTTGTCCGAACCTGGGGT 240

241 ATCATCTTCCATCTGGGTCCACTAAATCTCATGCAAGACA 280

35 281 TCTTGAGGGAGACCGAACAGTTCTCAACCAGCGTCTCAA 320

40 321 CACTGATACTTGGCTAGAGTCACGCTGAGTTGATCGGT 360

361 CTCCAAGCAAACATTGAGTTCAACCAGCAAGTGGACA 400

45 401 ACTTCTTGAATCCAACCTCAGAACATCTGTGCCCTTTCCAT 440

441 CACTTCTCCGTAAACACTATGCAGCAACTCTTCCTCAAC 480

481 AGATTGCCTCAGTTCAAGTCAAGGCTACCGATTGCTCC 520

55 521 TTCTTCCACTCTTGCTCAGGCTGCCAACATGCACTTGTC 560

561 CTTCATACGTGACGTGATCCTCAACGCTGACGGAAATGGGA 600

5	601	ATCTCTGCAGCCACTCTAGGACATACAGAGACTACTTGA	640
10	641	GGAACACTACACTCGTGATTACTCCAACATTGCATCAACAC	680
15	681	TTATCAGACTGCCTTCGTGGACTCAATACTAGGCTTCAC	720
20	721	GACATGTTGAGTTCAGGACCTACATGTTCCCTAACGTGT	760
25	761	TTGAGTACGTCAGCATTTGGAGTCCTCAAGTACCAAGAG	800
30	801	CTTGATGGTGTCTCTGGAGCCAATCTCTACGCCCTGGC	840
35	841	AGTGGACCACAGCAAACCTCAGAGCTTACAGCTCAGAACT	880
40	881	GGCCATTCTTGATAGCTTGTCCAAGTCAACTCCAACTA	920
45	921	CATTCTCAGTGGTATCTCTGGGACCAGACTCTCCATAACC	960
50	961	TTTCCCAACATTGGTGGACTTCCAGGCTCCACTACAACCC	1000
55	1001	ATAGCCTTAACCTCTGCCAGAGTGAACCTACAGTGGAGGTGT	1040
60	1041	CAGCTCTGGATTGATTGGTGCACAACTTAACCTGAACCAAC	1080
65	1081	TTCAATTGCTCCACCGTCTTGCCACCTCTGAGCACACCGT	1120
70	1121	TTGTGAGGTCTGGCTTGACAGCGGTACTGATCGCGAAGG	1160
75	1161	AGTTGCTACCTCTACAAACTGGCAAACCGAGTCCTCCAA	1200
80	1201	ACCACTCTAGCCTCGGTGTGGAGCTTCTCTGCACGTG	1240
85	1241	GGAATTCAAACACTTTCCAGACTACTTCATTAGGAACAT	1280
90	1281	CTCTGGTGTCTCTCGTCATCAGGAATGAAGACCTCACC	1320
95	1321	CGTCCACCTCATTACACCAAGATTAGGAACATCGAGTCTC	1360

1361 CATCCGGTACTCCAGGAGGTGCAAGAGCTTACCTCGTGTGTC 1400  
 1401 TGTCCATAACAGGAAGAACACATCTACGCTGCCAACGAG 1440  
 1441 AATGGCACCATGATTCACCTTGACCAAGAGATTACACTG 1480  
 1481 GATTCAACCATCTCTCAATCCATGCTACCCAAAGTGAACAA 1520  
 1521 TCAGACACGCACCTTCATCTCCGAAAAGTCGGAAATCAA 1560  
 1561 GGTGACTCCTTGAGGTTCGAGCAATCCAACACTACCGCTA 1600  
 1601 GGTACACTTTGAGAGGCAATGAAACAGCTACAACCTTTA 1640  
 1641 CTTGAGAGTTAGCTCCATTGGTAACTCCACCATCCGTGTT 1680  
 1681 ACCATCAACGGACGTGTTACACAGTCTCTAATGTGAACA 1720  
 1721 CTACAACGAACAATGATGGCGTTAACGACAACGGAGCCAG 1760  
 1761 ATTCAAGGACATCAACATGGCAACATCGTGGCTCTGAC 1800  
 1801 AACACTAACGTTACTTGGACATCAATGTGACCCCTCAATT 1840  
 1841 CTGGAACCTCCATTGATCTCATGAACATCATGTTGTGCC 1880  
 1881 AACTAACCTCCCTCATTGTAC 1902

oder

K. einer Struktur-Gen-Sequenz, die für ein Fusionsprotein codiert, das die N-terminalen 610 Aminosäuren von  
 B.t.k. HD-1 und die C-terminalen 567 Aminosäuren von B.t.k. HD-73 aufweist, welches Gen die Sequenz hat:

1 ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA 40

41	ACTGCTTGAGTAACCCAGAAGTGAAGTACTTGGTGGAGA	80
81	ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG	120
121	TCCTTGACACAGTTCTGCTCAGCGAGTTCTGTGCCAGGTG	160
161	CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT	200
201	CTTTGGTCCATCTCAATGGGATGCATTCTGGTGCAAATT	240
241	GAGCAGTTGATCAACCCAGAGGGATCGAAGAGTTGCCAGGA	280
281	ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA	320
321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGGCCAT	360
361	CCTACTAACCCAGCTCTCCGCGAGGAATGCGTATTCAAT	400
401	TCAACGACATGAACAGCGCCTTGACCACAGCTATCCATT	440
441	GTTCGCAGTCCAGAACTACCAAGTTCTCTTGTCCGTG	480
481	TACGTTCAAGCAGCTAATCTCACCTCAGCGTGTCTCGAG	520
521	ACGTTAGCGTCTGGCAAAAGGTGGGATTCGATGCTGC	560
561	AACCATCAATAGCCGTTACACGACCTTACTAGGCTGATT	600
601	GGAAACTACACCGACCACGCTGTTCTGGTACAACACTG	640
641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680
681	TAGATACAACCACTGAGAGAATTGACCCCTCACAGTT	720
721	TTGGACATTGTTGTCCTCTCCGAACTATGACTCCAGAA	760
761	CCTACCCCTATCCGTACAGTGTCCCAACTTACCAAGAGAAAT	800

5 801 CTATACTAACCCAGTTCTTGAGAACCTCGACGGTAGCTTC 840  
 10 841 CGTGGTTCTGCCCAAGGTATCGAAGGCTCCATCAGGAGCC 880  
 15 881 CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC 920  
 20 921 CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCG 960  
 25 961 ATCATGGCCTCTCAGTTGGATTCAAGCGGGCCCCAGTTA 1000  
 30 1001 CCTTCCCTCTATGGAACATATGGGAAACGCCGCTCCACA 1040  
 35 1041 ACAACGTATCGTTGCTCAACTAGGTCAAGGGTGTCTACAGA 1080  
 40 1081 ACCTTGTCTTCCACCTTGTACAGAAGACCCCTTCATAATCG 1120  
 45 1121 GTATCAACACCCAGCAACTTCCGTTCTGACGGAAACAGA 1160  
 50 1161 GTTCGCCTATGGAACCTCTTCTAACTTGCCATCCGCTGTT 1200  
 55 1201 TACAGAAAAGAGCGGAACCGTTGATTCCCTTGGACGGAAATCC 1240  
 1241 CACCAACAGAACAAACATGTGCCACCCAGGCAAGGATTCTC 1280  
 1281 CCACAGGTGAGCCACGTCTCCATCTCCGTTCCGGATTC 1320  
 1321 AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT 1360  
 1361 CATGGATTCACTCGTAGTGCTGAGTTCAACAATCATTCC 1400  
 1401 TTCCCTCTCAAAATCACCCAAATCCATTGACCAAGTCTACT 1440  
 1441 AACCTTGGATCTGGAACCTCTGTCGTGAAAGGACCCAGGCT 1480  
 1481 TCACAGGAGGTGATATTCTTAGAAGAACCTCTCCGGCCA 1520

1521	GATTAGCACCCCTCAGAGTTAACATCACTGCACCACCTTCT	1560
1561	CAAAGATATCGTGTCAAGGATTGGTACGCATCTACCACTA	1600
1601	ACTTGCAATTCCACACCTCCATCGACGGAAGGCCTATCAA	1640
1641	TCAGGGTAACCTCTCCGCAACCATGTCAAGCGGCAGCAAC	1680
1681	TTGCAATCCGGCAGCTTCAGAACCGTCGGTTCACTACTC	1720
1721	CTTTCACCTCTCTAACGGATCAAGCGTTTCACCCCTAG	1760
1761	CGCTCATGTGTTCAATTCTGGCAATGAAGTGTACATTGAC	1800
1801	CGTATTGAGTTTGTGCCCTGCCAAGTTACCCCTGAGGCTG	1840
1841	AGTACAACCTTGAGAGAGGCCAGAAGGCTGTGAACGCCCT	1880
1881	CTTACCTCCACCAATCAGCTTGGCTTGAAACTAACGTT	1920
1921	ACTGACTATCACATTGACCAACTGTCCAACCTGGTCACCT	1960
1961	ACCTTAGCGATGAGTTCTGCCCTGACGAGAACGCGTGAAC	2000
2001	CTCCGAGAAAGTTAACACGCCAACGCGTCTCAGCGACGAG	2040
2041	AGGAATCTTGCAGACTCCAACCTCAAAGACATCAACA	2080
2081	GGCAGCCAGAACGTTGGTGGGTGAAAGCACCGGGATCAC	2120
2121	CATCCAAGGAGGCCGACGATGTCTCAAGGAGAACTACGTC	2160
2161	ACCCCTCCGGAACTTTCGACGAGTGCTACCCCTACCTACT	2200
2201	TGTACCAAGAGATCGATGAGTCCAAACTCAAAGCCTTCAC	2240
2241	CAGGTATCAACTTAGAGGCTACATCGAAGACAGCCAAGAC	2280

5	2281	CTTGAAATCTACTCGATCAGGTACAATGCCAAGCACGAGA	2320
10	2321	CCGTGAATGTCCCAGGTACTGGTTCCCTCTGGCCACTTTC	2360
15	2361	TGCCCAATCTCCATTGGGAAGTGTGGAGAGCCTAACAGA	2400
20	2401	TGCGCTCCACACCTTGAGTGGAAATCCTGACTTGGAAGTGT	2440
25	2441	CCTGCAGGGATGGCGAGAAGTGTGCCACCATTCTCATCA	2480
30	2481	CTTCTCTGGACATCGATGTGGATGTACTGACCTGAAT	2520
35	2521	GAGGACCTCGGAGTCTGGGTCACTTCAGATCAAGACCC	2560
40	2561	AAGACGGACACGCAAGACTTGGCACCTTGAGTTCTCGA	2600
45	2601	AGAGAAACCATTGGTCGGTGAAGCTCTCGCTCGTGTGAAG	2640
50	2641	AGACCAGAGAAGAAGTGGAGGGACAACCGTGAGAAACTCG	2680
55	2681	AATGGAAACTAACATCGTTACAAGGAGGCCAAGACTC	2720
60	2721	CGTGGATGCTTTGTTCTGTGAACCTCCAAATATGATCAGTTG	2760
65	2761	CAAGCCGACACCAACATGCCATGATCCACGCCGCAGACA	2800
70	2801	AACCTGTGACAGCATTCTGTGAGGCTTACTTGCTGAGTT	2840
75	2841	GTCCGTGATCCCTGGTGTGAACGCTGCCATCTCGAGGAA	2880
80	2881	CTTGAGGGACGTATCTTACCGCATTCTCCTTGTACGATG	2920
85	2921	CCAGAAACGTCAAGAACCGGTGACTTCACAAATGGCCT	2960
90	2961	CAGCTGCTGGAATGTGAAAGGTCACTGGACGTGGAGGAA	3000
95	3001	CAGAACAAATCAGCGTTCCGTCCTGGTGTGCCTGAGTGGG	3040

5                   3041 AAGCTGAAGTGTCCAAGAGGTTAGAGTCTGTCCAGGTAG   3080

10                  3081 AGGCTACATTCTCCGTGTGACCGCTTACAAGGACGGATAC   3120

15                  3121 GGTGAGGGTTGCGTGACCATCCACGAGATCGAGAACAAACA   3160

20                  3161 CCGACGAGCTTAAGTCTCCAACTGCGTCGAGGAAGAAAT   3200

25                  3201 CTATCCCAACAAACACCGTTACTTGCAACGACTACACTGTG   3240

30                  3241 AATCAGGAAGAGTACCGGAGGTGCCTACACTAGCCGTAACA   3280

35                  3281 GAGGTTACAACGAAGCTCCTCCGTTCTGCTGACTATGC   3320

40                  3321 CTCCGTGTACGGAGGAGAAATCCTACACAGATGGCAGACGT   3360

45                  3361 GAGAACCCCTTGCAGTTCAACAGAGGTTACAGGGACTACA   3400

50                  3401 CACCACTTCCAGTTGGCTATGTTACCAAGGAGCTTGAGTA   3440

55                  3441 CTTTCCTGAGACCGACAAAGTGTGGATCGAGATCGGTGAA   3480

60                  3481 ACCGAGGGAACCTTCATCGTGGACAGCGTGGAGCTCTCT   3520

65                  3521 TGATGGAGGAA 3531.

## Revendications.

1. Procédé de modification d'une séquence de gène de structure du type sauvage qui code une protéine insecticide de *Bacillus thuringiensis* afin d'activer l'expression de ladite protéine chez des plantes qui comprend :

50                  a) l'identification de régions à l'intérieur de ladite séquence comprenant plus de quatre nucléotides consécutifs d'adénine ou de thymine,

55                  b) la modification des régions de l'étape a) qui comportent deux ou plusieurs signaux de polyadénylation à l'intérieur d'une séquence de dix bases afin d'éliminer lesdits signaux tout en conservant une séquence de gène qui code ladite protéine, et

60                  c) la modification des régions de 15 à 30 bases entourant les régions de l'étape a) afin d'éliminer les signaux majeurs de polyadénylation de plantes, les séquences consécutives contenant plus d'un signal mineur de polyadénylation et les séquences consécutives contenant plus d'une séquence ATTAA tout en conservant une séquence de gène qui code ladite protéine.

2. Procédé de modification d'une séquence de gène de structure du type sauvage qui code une protéine Insecticide de *Bacillus thuringiensis* afin d'activer l'expression de ladite protéine chez des plantes qui comprend :

- l'élimination des signaux de polyadénylation contenus dans ledit gène de type sauvage tout en conservant une séquence qui code ladite protéine, et
- l'élimination des séquences ATTTA contenues dans ledit gène de type sauvage tout en conservant une séquence qui code ladite protéine.

3. Procédé selon la revendication 2, comprenant en outre l'élimination des séquences autocomplémentaires et le remplacement de telles séquences par de, l'ADN non autocomplémentaire comprenant des codons préférés des plantes tout en conservant une séquence de gène de structure codant ladite protéine.

4. Procédé selon les revendications 1 à 3, comprenant en outre l'utilisation des séquences préférées des plantes au cours de l'élimination des signaux de polyadénylation et des séquences ATTTA.

5. Procédé selon les revendications 1 à 3, dans lequel les signaux de polyadénylation des plantes sont choisis parmi le groupe constitué de AATAAA, AATAAT, AACCAA, ATATAA, AATCAA, ATACTA, ATAAAA, ATGAAA, AACCAT, ATTAAT, ATACAT, AAAATA, ATTTAA, AATACA et CATAAA.

6. Procédé destiné à améliorer l'expression d'un gène hétérologue chez des plantes dans lequel ledit gène comprend un gène chimère modifié comprenant un promoteur qui agit dans les cellules végétales liées de façon fonctionnelle à une séquence de structure codante et à une région 3' non traduite contenant un signal de polyadénylation qui agit chez des plantes pour provoquer l'addition de nucléotides de polyadénylate sur l'extrémité 3' de l'ARN, dans lequel ladite séquence de structure codante code une protéine insecticide dont une partie au moins est dérivée d'une protéine de *Bacillus thuringiensis*, dans lequel ledit procédé comprend la modification de ladite séquence de structure codante de sorte que ladite séquence comporte une séquence d'ADN qui diffère de la séquence d'ADN apparaissant dans la nature codant ladite protéine de *Bacillus thuringiensis* et ladite séquence de structure codante ne contient pas plus de 5 nucléotides consécutifs constitués de restes soit adénine, soit thymine.

7. Procédé d'amélioration de l'expression d'un gène hétérologue chez des plantes dans lequel ledit gène comprend un gène chimère modifié comprenant un promoteur qui agit dans des cellules végétales liées de façon fonctionnelle à une séquence de structure codante et à une région 3' non traduite contenant un signal de polyadénylation qui agit chez des plantes pour provoquer l'addition de nucléotides de polyadénylate sur l'extrémité 3' de l'ARN, dans lequel ladite séquence de structure codante code une protéine insecticide dont au moins une partie est dérivée d'une protéine de *Bacillus thuringiensis*, dans lequel ledit procédé comprend la modification de ladite séquence de structure codante de sorte que ladite séquence comporte une séquence d'ADN qui diffère de la séquence d'ADN qui apparaît dans la nature codant ladite protéine de *Bacillus thuringiensis* et présente les caractéristiques suivantes :

ladite séquence de structure codante comporte une région qui est complémentaire de la séquence suivante :

GGCTTGATTCTAGCGAACTCTCGATTCTCTGGTTGATGAGCTGTTCT  
 1 5 10 15 20 25 30 35 40 45

ladite région dans ladite séquence codante ayant éliminé 2 séquences AACCAA et 1 séquence ATTTA.

8. Procédé selon la revendication 7, dans lequel ladite séquence de structure codante code une protéine insecticide dont au moins une partie est dérivée de *Bacillus thuringiensis kurstaki* HD-1.

9. Procédé selon la revendication 7 ou 8, dans lequel la plante est un plan de tabac.

10. Gène chimère modifié contenant un promoteur qui agit dans des cellules végétales liées de façon fonctionnelle à une séquence de structure codante et à une région 3' non traduite contenant un signal de polyadénylation qui agit chez des plantes pour provoquer l'addition de nucléotides de polyadénylate sur l'extrémité 3' de l'ARN, dans lequel ladite séquence de structure codante code une protéine insecticide dont au moins une partie est dérivée d'une

protéine de *Bacillus thuringiensis*, dans lequel ladite séquence de structure codante comporte une séquence d'ADN qui diffère de la séquence d'ADN apparaissant dans la nature codant ladite protéine de *Bacillus thuringiensis* et est choisie à partir de :

5 A. Un gène de structure qui code une protéine insecticide de *B.t.k* HD-1 comportant la séquence :

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1	ATGGCTATAGAAACTGGTTACACCCCAATGGATAATTCCCT	40
5		
41	TGTGGCTAACGCAATTCTTTGAGTGAATTGTCGGG	80
10		
81	TGCTGGATTGTGTTAGGACTAGTTGATATTATCTGGGA	120
15		
121	ATTTTGGTCCCTCTAATGGGACGCATTCTTGTACAAA	160
20		
161	TTGAACAGCTCATCRAACCAGAGAATCGAAGAGTTGCTAG	200
25		
201	GAATCAAGCCATTCTAGATTAGAAGGACTAACGAACTTT	240
30		
241	TATCAAATTACGCAGAACATTAGAGAGTGGGAAGCAG	280
35		
281	ATCCTACTAATCCAGCATTAAGAGAACAGATGCGTATTCA	320
40		
321	ATTCAATGACATGACAGTGCCTTACAACCGCTATTCCCT	360
45		
361	CTTTTGAGTTCAAAATTATCAAGTTCCCTCTCCCTCTCCG	400
50		
401	TGTACGTTCAAGCTGCCAACCTCCACCTCTCAGTTTGAG	440
55		
441	AGATGTTTCAGTGTGTTGGACAAAGGTGGGATTTGATGCC	480
60		
481	GCGACTATCAATAGTCGTATAATGATTAACTAGGCTTA	520
65		
521	TTGGCAACTATACAGATCATGCTGTACGCTGGTACAATAC	560
561	GGGATTAGAGCGTGTATGGGGACCCGATTCTAGAGATTGG	600
601	ATCAGGTACAACCAGTTCAAGAGAGCTTACACTAATCTG	640
641	TATTAGATATCGTTCTCTATTTCGAACTATGATAGTAG	680
681	AACGTATCCAATTGAAACAGTTCCCAATTAACAAGAGAA	720

5	721	ATTTATACAAACCCAGTATTAGAAAAATTTGTTGGTAGTT	760
10	761	TTCGAGGCTCGGCTCAGGGCATAGAAGGAAAGTATTAGGAG	800
15	801	TCCACATTTGATGGATATACTTAATAGTATAACCATCTAT	840
20	841	ACGGATGCTCATAGAGGAGAAATACTACTGGTCCGGTCACC	880
25	881	AGATCATGGCTTCTCCTGTAGGGTTTCGGGGCCAGAATT	920
30	921	CACTTTCCGCTATATGGAACATATGGAAATGCCAGCTCCA	960
35	961	CAACAAACGTATTGTTGCTCAACTAGGTCAAGGGGTGTATA	1000
40	1001	GAACATTATCGTCCACCTTATATAGAAGACCTTTAACAT	1040
45	1041	CGGGATCAACAAACCAACAACTATCTGTTCTGACGGGACA	1080
50	1081	GAATTGCTTATGGAACCTCCTCAAATTTGCCATCCGCTG	1120

1121 TATAACGAAAAAGCGGAACGGTAGATTCGCTGGATGAAAT 1160  
 1161 ACCGCCACAGAATAACAACGTGCCACCTAGCAAGGATT 1200  
 1201 AGTCATCGATTAAGCCATGTTCAATGTTCGTTAGGCT 1240  
 1241 TTAGTAATAGTAGTGTAAAGTATAATAAGAGCTCCTATGTT 1280  
 1281 CTCTGGATACATCGTAGTGCTGAGTTCAACAAACATCATC 1320  
 1321 CCTTCATCRAAAATCACCCAAATCCCACTCACCAAGTCTA 1360  
 1361 CTAATCTGGCTCTGGAACCTCTGTCGTTAAAGGACCAGG 1400  
 1401 ATTTACAGGAGGAGATATTCTTCGAAGAACCTCACCTGGC 1440  
 1441 CAGATTCAACCTTAAGAGTAAATATTACTGCACCAATT 1480  
 1481 CACAAAGATATCGGTAAGAATTGCTACGCTTCTACCAAC 1520  
 1521 AAACCTTCAGTTCCACACATCAATTGACGGAAGACCTATT 1560  
 1561 AATCAGGGGAATTTCAGCAACTATGAGTAGTGGAGTA 1600  
 1601 ATTTACAGTCGGAAAGCTTTAGGACTGTAGGTTTACTAC 1640  
 1641 TCCGTTAACCTTCAAAATGGATCAAGTGTATTACGTTA 1680  
 1681 AGTGCTCATGTCCTCAATTCAAGGAAATGAAGTTTATATAG 1720  
 1721 ATCGAATTGAATTGTTCCGGCA 1743

55. B. Un gène de structure qui code une protéine Insecticide de *B.t.k* HD-73 comportant la séquence :

1 ATGCCATTGAAACCGTTACCTCCCATCGACATCTCCT 40  
 5  
 41 TGTCTTGACACAGTTCTGCTCAGCGAGTTCGTGCAGG 80  
 10  
 81 TGCTGGGTTCTCGTCTCGGACTAGTTGACATCATCTGGGT 120  
 15  
 121 ATCTTGGTCCATCTCAATGGGATGCATTCTGGTGCAAA 160  
 20  
 161 TTGAGCAGTTGATCAACCAGAGGATCGAAGAGTTGCCAG 200  
 25  
 201 GAACCAAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTC 240  
 30  
 241 TACCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCG 280  
 35  
 281 ATCCTACTAACCCAGCTCTCCGCAGGAAATGCGTATTCA 320  
 40  
 321 ATTCAACGACATGAACAGCGCCTTGACCACAGCTATCCCA 360  
 45  
 361 TTGTTCGCAGTCCAGAACTACCAAAGTTCCCTCTTGTCCG 400  
 50  
 401 TGTACGTTCAAGCAGCTAATCTCACCTCAGCGTGCCTCG 440

5	441	AGACGGTTAGCGTGTGGGCAAAGGTGGGGATCGATGCT	480
10	481	GCAACCATCAATAGCCGTTACAACGACCTTACTAGGCTGA	520
15	521	TTGGAAAACACACCGACCACCGTGTGGTACAACAC	560
20	561	TGGCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGG	600
25	601	ATTAGATAACAACCAGTTCAAGGAGAGAATTGACCCCTCACAG	640
30	641	TTTGGACATTGTGTCCTCTTCCCGAACATATGACTCCAG	680
35	681	AACCTACCCCTATCCGTACAGTGTCCTAACCTACAGAGAA	720
40	721	ATCTATACTAACCCAGTTCTGAGAACATTGACGGTAGCT	760
45	761	TCCGGGTTCTGCCCAAGGTATCGAAGGCTCCATCAGGAG	800
50	801	CCCACACTTGATGGACATCTGAACAGCATAACTATCTAC	840
55	841	ACCGATGCTCACAGAGGGAGACTTACTGGCTGGACACC	880
60	881	AGATCATGGCTCTCCAGTTGGATTCAAGCGGGCCCGAGTT	920
65	921	TACCTTCCCTCTATGGAACATAGGGAAACGCCCTCCA	960
70	961	CAACACGTATCGTTGCTCAACTAGGTCAAGGGTGTCTACA	1000
75	1001	GAACCTTGTCTTCCACCTTGTACAGAAGACCCCTCAATAT	1040

5	1041	CGGTATCAACAACCGCAACTTCCGTTCTGACGGAAACA	1080
10	1081	GAGTTGCCATGGAACCTCTTCAACTTGCATCCGCTG	1120
15	1121	TTTACGAAAAGAGCGGAACCGTTGATTCCCTGGACGAAAT	1160
20	1161	CCCACACAGAACAAACAATGTGCCACCCAGGCAAGGATTC	1200
25	1201	TCCACAGGTGAGGCCACGTGTCCATGTTCCGTTCCGGAT	1240
30	1241	TCAGCRAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTT	1280
35	1281	CTCTTGGATACACCGTAGTGGTAGGTTCAACAAACATCAAC	1320
40	1321	GCATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAA	1360
45	1361	ACTTCTCTCAACGGTTCTGTCATTCAAGGACCAAGGATT	1400
50	1401	CACTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAAT	1440
55	1441	AACTTCAGAAATAGAGGGTATATTGAAGTTCAATTCACT	1480
60	1481	TCCCATCCACATCTACCAAGATATAGACTTCGTGTGAGGTA	1520
65	1521	TGCTTCTGTGACCCCTATTCAACCTCAACGTTAATTGGGGT	1560
70	1561	AATTCACTCCATCTCTCCAATACAGTTCCAGCTACAGCTA	1600
75	1601	CCTCCTGGATAATCTCCAATCCAGCGATTTCGGTTACTT	1640

5  
 1641 TGAAAGTGCCAATGCTTTACATCTTCACTCGGTAAACATC 1680  
 1681 GTGGGTGTTAGAAACTTTAGTGGACTGCAGGAGTGATTA 1720  
 10 1721 TCGACAGATTGAGTTCATTCAGTTACTGCAACACTCGA 1760  
 15 1761 GGCTGAG 1767.

20 C. Un gène de structure codant une protéine insecticide de *B.t.k*. HD-1 comportant la séquence :

25 1 ATGGACAACAAACCCAAACATCAACGAATGCATTCCATACA 40  
 41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80  
 50 81 ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG 120  
 121 TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG 160  
 161 CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT 200  
 55 201 CTTTGGTCCATCTCAATGGGATGCATTCCCTGGTGCAAATT 240  
 241 GAGCAGTTGATCAACCGAGGGATCGAAGAGTTGCCAGGA 280  
 40 281 ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA 320  
 45 321 CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT 360

5	361	CCTACTAACCCAGCTCTCCGCAGGAAATGGGTATTCAAT	400
10	401	TCAACGACATGAACAGCGCTTGACCACAGCTATCCCATT	440
15	441	GTTCCAGTCCAGAACTACCAAGTTCTCTTGTCCGTG	480
20	481	TACGTTCAAGCAGCTAACCTTCACCTCAGCGTGCTTCGAG	520
25	521	ACGTTAGCGTGTGGCRAAGGTGGGATTGATGCTGC	560
30	561	AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600
35	601	GGAAAATCACCGGACCGCTGTTGTTGGTACAACACTG	640
40	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680
45	681	TAGATACACCAAGTTCAAGGAGAGAATTGACCCCTCACAGTT	720
50	721	TTGGACATTGTGTCTCTTCCCAGACTATGACTCCAGAA	760
55	761	CCTACCCCTATCCGTACAGTGTCCCAACTTACCAAGAGAAAT	800
	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
	841	CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
	881	CACACTTGATGGACATCTTGAACAGCATAACTATCACAC	920
	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAAG	960

5	961	ATCATGGCCTCTCCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
10	1001	CCTTTCCCTCTATGGAACATATGGAAACCCCGCTCCACA	1040
15	1041	ACAACGTATCGTGTCAACTAGGTCAAGGGTGTACAGA	1080
20	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCCTCAATATCG	1120
25	1121	GTATCAACACCAGCAACTTCCGTTCTGACGGAACAGA	1160
30	1161	GTTCGCCTATGGAACCTCTCTAACCTGCCATCCGCTGTT	1200
35	1201	TACAGAAAAGCGGAACCGTTGATTCTTGGACGAAATCC	1240
40	1241	CACCACAGAACAAACAATGTGCCACCCAGGCAAGGATTCTC	1280
45	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTG	1320
50	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCTATGTTCT	1360
55	1361	CATGGATTCACTCGTAGTGTGAGTTCAACAATATCATTCC	1400
	1401	TTCCCTCTCAAATCACCCAAATCCCATTGACCAAGTCTACT	1440
	1441	AAACCTGGATCTGGAACCTCTGTCGTGAAAGGACCGAGGCT	1480
	1481	TCACAGGAGGTGATATCTTAGAAGAACCTCTCCTGGCCA	1520
	1521	GATTAGCACCCCTCAGAGTTAACATCACTGCACCACTTCT	1560

1561	CAAAGATACTGTCAGGATTGTTACGCATCTACCACTA	1600
1601	ACTTGCAATTCCACACCTCCATCGACGGAAGGCCTATCAA	1640
1641	TCTGGGTAACCTCTCCGCAACCATGTCAGCGGCAGCAAC	1680
1681	TTGCAATCCGGCAGCTTCAGAACCGTCGGTTCACTACTC	1720
1721	CTTTCAACTCTCTAACGGATCAAGCGTTTCACCCCTAG	1760
1761	CGCTCATGTGTTCAATTCTGGCAATGAAGTGTACATTGAC	1800
1801	CGTATTGAGTTTGCCCTGCCGAAGTTACCTTCGAGGCTG	1840
1841	AGTAC 1845.	

D. Un gène de structure codant une protéine insecticide dérivée de *B.t.k.* HD-73 comportant la séquence :

1	ATGGACAAACAACCCAAACATCAACGAATGCATTCCATACA	40
41	ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA	80
81	ACCGCATGAAACCGGGTACACTCCCATCGACATCTCCTTG	120
121	TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG	160
161	CTGGGTTCTGTCCTCGGACTAGTTGACATCATCTGGGGTAT	200

5	201	CTTTGGTCCATCTCAATGGGATGCAATTCTGGTGC	AAATT	240
10	241	GAGCAGTTGATCAACCAGAGGGATCGAAGAGTT	GCCAGGA	280
15	281	ACCAGGCCATCTCTAGTTGGAAGGATTGAGCA	ATCTCTA	320
20	321	CCRAATCTATGCAGAGAGCTTCAGAGAGT	GGGAAGCCGAT	360
25	361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGT	ATTCAAT	400
30	401	TCAACCGACATGAACAGCGCCTTGACCACAGC	TATCCATT	440
35	441	GTTCGCAGTCCAGAACTACCA	AGTTCCCTCTCTTGTCCGTG	480
40	481	TACGTTCAAGCAGCTAATCTTCACCTCAGCGT	GCTTCGAG	520
45	521	ACGTTAGCGTGTGTTGGGCAAAAGGTGGGATT	CGATGCTGC	560
50	561	AACCATCAATAGCGTTACAACGACCTTACTAGG	CTGATT	600
55	601	GGAAACTACACCGACCACGCTGTTCTGGTACA	ACACTG	640
60	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGAT	GGAT	680
65	681	TAGATACAACCAAGTTCAAGGAGAGAATTGAC	CCCTCACAGTT	720
70	721	TTGGACATTGTGTCTCTTCCCGAACTATGACT	CCAGAA	760
75	761	CCTACCCCTATCCGTACAGTGTCCC	AACTTACCA	800

5	801	CTATACTAACCCAGTTCTTGAGAACCTTCGACGGTAGCTTC	840
10	841	CGTGGTTCTGCCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
15	881	CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC	920
20	921	CGATGCTCACAGAGGGAGAGTATTACTGGTCTGGACACCGAG	960
25	961	ATCATGGCCTCTCCAGTTGGATTCAAGCGGGCCCGAGTTTA	1000
30	1001	CCTTTCCTCTATGGAACATATGGGAAACGCCGCTCCACA	1040
35	1041	ACAACGTATCGTTGCTCAACTAGGTCAAGGTGTCTACAGA	1080
40	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCCTCAATATCG	1120
45	1121	GTATCAACAACCAGCAACTTTCCGTTCTGACCGAACAGA	1160
50	1161	GTTGCCCTATGGAACCTCTCTAACTTGCCATCCGCTGTT	1200
55	1201	TACAGAAAGAGCGGAACCGTTGATTCTGGACCGAAATCC	1240
60	1241	CACCAACAGAACAAATGTGCCACCCAGGCAAGGATTCTC	1280
65	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTTC	1320
70	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCTATGTTCT	1360
75	1361	CTTGGATAACACCGTAGTGCTGAGTTCAACAAACATCATCGC	1400

5                    1401 ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC    1440  
 .  
 .  
 .  
 10                1441 TTTCTCTCAACGGTCTGTCAATTCAAGGACCAAGGATTCA    1480  
 .  
 .  
 .  
 15                1481 CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA    1520  
 .  
 .  
 .  
 20                1521 CATTCAAAATAGAGGGTATATTGAAGTTCCAATTCACTTC    1560  
 .  
 .  
 .  
 25                1561 CCATCCACATCTACCAAGATATAGAGTTCTGTTGAGGTATG    1600  
 .  
 .  
 .  
 30                1601 CTTCTGTGACCCCTATTCAACCTCAACGTTAATTGGGTAA    1640  
 .  
 .  
 .  
 35                1641 TTCACTCCATCTTCTCCAATAACAGTTCCAGCTACAGCTACC    1680  
 .  
 .  
 .  
 40                1681 TCCTTGGATAATCTCCAATCCAGGGATTTGGTTACTTTG    1720  
 .  
 .  
 .  
 45                1721 AAAGTCCAATGCTTTACATCTTCACTCGTAACATCGT    1760  
 .  
 .  
 .  
 50                1761 GGGTGTAGAAACTTTAGTGGGACTGCAGGAGTGTATTAC    1800  
 .  
 .  
 .  
 55                1801 GACAGATTGAGTTCAATTCCAGTTACTGCACACACTCGAGG    1840  
 .  
 .  
 .  
 60                1841 CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTAAATGCG    1880  
 .  
 .  
 .  
 65                1881 CTGTTTACGTCTACAAACCAGCTTGGACTCAAGACAAATG    1920  
 .  
 .  
 .  
 70                1921 G 1921

E. Un gène de structure codant la protéine insecticide en pleine longueur de *B.t.k.* HD-73 comportant la séquence :

5	1	ATGGACAAACRACCCAAACATCAACGATGCATTCCATACA	40
10	41	ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA	80
15	81	ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCTTG	120
20	121	TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCCAGGTG	160
25	161	CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT	200
30	201	CTTTGGTCCATCTCAATGGGATGCATTCTGGTGCAAATT	240
35	241	GAGCAGTTGATCAGGAGGGATCGAAGAGTTCGCCAGGA	280
40	281	ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAAATCTCTA	320
45	321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
50	361	CCTACTAACCCAGCTCTCCCGAGGAAATGCGTATTCAAT	400
55	401	TCAACCGACATGAACAGCGCTTGTACCCACAGCTATCCCATT	440
60	441	GTTCGCAGTCCAGAACTACCAAGTTCTCTCTTGTCCGTG	480
65	481	TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG	520
70	521	ACGTTAGCGTGTGGCAAAGGTGGGATTGATGCTGC	560
75	561	AACCATCAATAGCCGTACAACGACCTTACTAGGCTGATT	600

5	601	GGAAACTACACCGACCACCGCTGTCGGTGGTACAACACTG	640
10	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680
15	681	TAGATACAACCAGTTCAAGGAGAGAATTGACCCCTCACAGTT	720
20	721	TTGGACATTGTGTCTCTCTTCCCGAACATATGACTCCAGAA	760
25	761	CCTACCCCTATCCGTACAGTGTCCCAACTTACCAAGAGAAAT	800
30	801	CTATACTAACCCAGTTCTTGAGAACCTTCGACGGTAGCTTC	840
35	841	CGTGGTTCTGCCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
40	881	CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC	920
45	921	CGATGCTCACAGAGGAGAGTATTACTGGCTGGACACCAG	960
50	961	ATCATGGCCTCTCCAGTTGGATTCAAGCGGGCCCGAGTTA	1000
55	1001	CCTTTCCCTCTATGAACTATGGAAACCGCCGCTCCACA	1040
60	1041	ACAACGTATCGTTGCTCAACTAGGTCAAGGGTGTACAGA	1080
65	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCCTCAATATCG	1120
70	1121	GTATCAACACCGACGCAACTTCCGTTCTGACGGAACAGA	1160
75	1161	GTTCGCCTATGGAACCTCTTCTAACTTGGCATCCGCTGTT	1200

1201	TACAGAAAGAGCGGAACCGTTGATTCCCTGGACGAAATCC	1240
1241	CACCAAGAACAAACATGTGCCACCCAGGCAAGGATTCTC	1280
1281	CCACAGGTTGAGCCACGTGTCCATGTTCGGTCCGGATTG	1320
1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCTATGTTCT	1360
1361	CTTGGATAACCGTAGTGCTGAGTTAACAAACATCATCGC	1400
1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
1441	TTTCTTCAACGGTTCTGTCAATTCAAGGACCAGGATTCA	1480
1481	CTGGTGGAGACCTCGITAGACTCAACAGCAGTGGAAATAA	1520
1521	CATTAGAAATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
1561	CCATCCACATCTACCAAGATATAGAGTTCTGTGAGGTATG	1600
1601	CTTCTGTGACCCCTATTCAACGTTAATTGGGGTAA	1640
1641	TTCATCCATCTTCTCAATACAGTTCCAGCTACAGCTACC	1680
1681	TCCTTGGATAATCTCCAATCCAGCGATTTCGGTTACTTTG	1720
1721	AAAGTGCCAATGCTTTACATCTCACTCGGTAAACATCGT	1760
1761	GGGTGTTAGAAACTTTAGTGGGACTGCAGGAGTGATTATC	1800

5	1801	GACAGATTGGAGTTCATTCAGTTACTGCAACACTCGAGG	1840
10	1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
15	1881	GCTGTTACGTCTACAAACCAGCTCGGCCTCAAGACCAAT	1920
20	1921	GTGACGGATTATCATATTGATCAAGTGTCCAACTTGGTGA	1960
25	1961	CCTACCTCAGCGATGAGTTCTGTCGGATGAAAAGCGAGA	2000
30	2001	ATTGTCCGAGAAAGTCAAACATGCGAAGCGACTCAGTGAT	2040
35	2041	GAACGCAATTACTCCAAGATTCAAATTCAAAGACATTA	2080
40	2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120
45	2121	TACCATCCAGGGAGGTGACCGACGTGTTCAAGGAGAACTAC	2160
50	2161	GTCACACTATCAGGTACCTTTGATGAGTGCTATCCAACAT	2200
55	2201	ACCTCTACCAGAAGATCGACGAGTCCAAGTGTGAAAGCCTT	2240
60	2241	TACCCGTTATCAATTAAAGAGGGTATATCGAAGATAGTCAA	2280
65	2281	GACCTCGAGATCTACCTCATCCGCTACAATGCAAAACATG	2320
70	2321	AAACAGTAAATGTGCCAGGTACGGGTTCCCTTATGGCCGCT	2360
75	2361	TTCAGCCCAAAGTCAAATCGGAAGTGTGGAGAGCCGAAT	2400

5	2401	CGATGCGCGCCACCTTGAATGGAATCCTGACTTAGATT	2440
10	2441	GTTCTGTAGGGATGGAGAAAAGTGTGCCCATTCGCA	2480
15	2481	TCATTTCTCCTTAGACATTGATGTAGGATGTACAGACTTA	2520
20	2521	AATGAGGACCTAGGTGTATGGGTGATCTTAAGATTAAAGA	2560
25	2561	CGCAAGATGGCACCGCAAGACTAGGGAAATCTAGAGTTCT	2600
30	2601	CGAAGAGAAACCATTAGTAGGAGAACCGCTAGCTCGTGTG	2640
35	2641	AAGAGAGCGGAGAAAAATGGAGAGAACAGTGAGAAGT	2680
40	2681	TGGAATGGGAGACCAACATCGTCTACAAAGAGGCAGAAGA	2720
45	2721	ATCTGTAGATGCTTTATTTGTAACCTCTCAATATGATCAA	2760
50	2761	TTACAAGCGGATACGAATATTGCCATGATTCATGCGGCAG	2800
55	2801	ATAAACGTGTTCATAGCATTGGAGAACGTTATCTGCCTGA	2840
	2841	GCTGTCGTGATTCCGGGTGTCAATCGGGCTATTTTGAA	2880
	2881	GAATTAGAAGGGCGTATTTCACTGCATTCTCCCTCTACG	2920
	2921	ATGCCAGAACGTCATCAAGAACGGTGACTTCACAAATGG	2960
	2961	CTTATCCTGCTGAAACGTGAAAGGGCATGTAGATGTAGAA	3000

5	3001	GAACAAAACAACCAACGTTCGGTCTTGTGTCGGAAAT	3040
10	3041	GGGAAGCAGAAGTGTACACAAGAAGTTCGTGTCGTCCGGG	3080
15	3081	TCGTGGCTATATCCTCGTGTACAGCGTACAAGGAGGG	3120
20	3121	TATGGAGAAGGTTGCGTAACCATTGAGATCGAGAAC	3160
25	3161	ATACAGACGAACTGAAGTTAGCAACTGCGTAGAAGAGGA	3200
30	3201	AATCTATCCAAAATAACACGGTAACGTGTAATGATTATACT	3240
35	3241	GTAATCAAGAAGAATACGGAGGTGCGTACACTCTCGTA	3280
40	3281	ATCGAGGATATAACGAAGCTCCTCCGTACCGCTGATTA	3320
45	3321	TGGCTCAGTCTATGAAGAAAAATCGTATAACAGATGGACGA	3360
50	3361	AGAGAGAATCCTTGTGAATTAAACAGAGGGTATAGGGATT	3400
55	3401	ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA	3440
60	3441	ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
65	3481	GAAACCGAAGGAACATTATCGTGGACAGCGTGGATTAC	3520
70	3521	TCCTTATGGAGGAA 3534	

F. Un gène de structure codant une protéine Insecticide en pleine longueur de *B.t.k* HD-73 comportant la séquence :

5	1	ATGGACAAACAAACCAACATCAACGAATGCATTCCATACA	40
10	41	ACTGCTTGAAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA	80
15	81	ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCCTG	120
20	121	TCCTTGACACAGTTCTGCTCAGCGAGTTCTGCCAGGTG	160
25	161	CTGGGTTCTCGACTAGTTGACATCATCTGGGTAT	200
30	201	CTTGGTCCATCTCAATGGGATGCATTCTGGTGCAAATT	240
35	241	GAGCAGTTGATCAACCAGAGGATCGAAGAGTTGCCAGGA	280
40	281	ACCAAGGCCATCTCTAGGTTGGAAAGGATTGAGCAATCTCTA	320
45	321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
50	361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT	400
55	401	TCAACGGACATGAAACAGCGCCCTGACCACAGCTATCCCATT	440
	441	GTTCGCAGTCCAGAACTACCAAGTTCTCTCTGTCCGTG	480
	481	TACGGTCAAGCAGCTAATCTTCACCTCAGCGTGCTTCGAG	520
	521	ACGTTAGCGTGTGGCAAAAGGTGGGATTCGATGCTGC	560

5	561	AACCATCAATAGCCGTACAAACGACCTACTAGGCTGATT	600
10	601	GGAAACTACACCGACCAACGCTGTTGGTACAAACAGTG	640
15	641	GCTTGGAGCGTGTCTGGGTCTGATTCAGAGATTGGAT	680
20	681	TAGATACAAACCAAGTTCAAGGAGAGAAATTGACCCCTACAGTT	720
25	721	TTGGACATTGTGTCTCTCTTCCCGAACTATGACTCCAGAA	760
30	761	CCTACCCCTATCCGTACAGTGTCCAACTTACCAAGAGAAAT	800
35	801	CTATACTAACCCAGTTCTTGAGAACTTCCACGGTAGCTTC	840
40	841	CGTGGTTCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
45	881	CACACTTGATGGACATCTTGAACAGCATAACTATCTACAC	920
50	921	CGATGCTCACAGAGGAGAGTATTACTGGCTGGACACCAG	960
55	961	ATCATGGCCTCTCCAGTTGGATTCAAGGGGCCGAGTTTA	1000
60	1001	CCTTCCTCTATGAACTATGGAAACGCCGCTCCACAA	1040
65	1041	ACRACGTATCGTGTCAACTAGGTCAAGGGTGTCTACAGA	1080
70	1081	ACCTTGTCTCCACCTTGTACAGAAGACCCCTCAATATCG	1120
75	1121	GTATCAACAAACCAAGCAACTTCCGTTCTGACGGAACAGA	1160
80	1161	GTTGCCATGGAAACCTCTCTAACTTGCCTCCGCTGT	1200
85	1201	TACAGAAAGACCCGAACCGTTGATTCTTGGACGAAATCC	1240
90	1241	CACCAACAGAACAAATGTGCCACCCAGGCAAGGATTCTC	1280
95	1281	CCACAGGTTGAGCCACGTGTCCATGTTCCGTTCCGGATTC	1320

5	1321	AGCAACAGTCCGTGAGCATCATCAGAGCTCTATGTTCT	1360
10	1361	CTTGGATACACCGTAGTGTGAGTTCAACAACATCATCGC	1400
15	1401	ATCCGATAGTATTACTCAAATCCCTGCRGTGAAGGGAAAC	1440
20	1441	TTTCTCTTCAACGGTTCTGTCAATTTCAGGACCGAGGATTCA	1480
25	1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
30	1521	CATTCAAGATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
35	1561	CCATCCACATCTACCAAGATATAGAGTTCGTGTGAGGTATG	1600
40	1601	CTTCTGTGACCCCTAATCACCTCAACGTTAATTGGGTAA	1640
45	1641	TTCATCCATCTCTCCAATACAGTTCCAGCTACAGCTACC	1680
50	1681	TCCCTGGATAATCTCCAATCCAGCGATTCCGGTTACTTTG	1720
55	1721	AAAGTGCCAATGCTTTACATCTTCACTCGGTAAACATCGT	1760
60	1761	GGGTGTTAGAAACTTTAGTGGGACTGCAGGAGTGATTATC	1800
65	1801	GACAGATTGAGTTCAATTCCAGTTACTGCAACACTCGAGG	1840
70	1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
75	1881	GCTGTTACGTCTACAAACCAACTAGGGCTAAAAACAAAT	1920
80	1921	GTAACGGATTATCATATTGATCAAGTGTCCAATTAGTTA	1960
85	1961	CGTATTTATCGGATGAATTGTCTGGATGAAAGCGAGA	2000
90	2001	ATTGTCCGAGAAAGTCACACATGCGAAGCGACTCAGTGAT	2040
95	2041	GAACGCAATTACTCCAAGATTCAAATTCAAGACATTA	2080

2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120
2121	TACCATCCAAGGAGGGATGACGTATTTAAAGRAAAATTAC	2160
2161	GTCACACTATCAGGTACCTTGATGAGTGCTATCCAACAT	2200
2201	ATTTGTATCAAAAATCGATGAATCAAAATTAAAGCCCTT	2240
2241	TACCCGTTATCAATTAGAGGGTATATCGAAGATAGTCAA	2280
2281	GACTTAGAAAATCTATTTAATTCGCTACATGCAAAACATG	2320
2321	AAACAGTAAATGTGCCAGGTACGGGTTCCCTATGGCCGCT	2360
2361	TTCAAGCCAAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400
2401	CGATGCGGCCACACCTTGAATGAAATCCTGACTTAGATT	2440
2441	GTTCGTGTAGGGATGGAGAAAAGTGTGCCCATTCGCA	2480
2481	TCATTTCTCCTTAGACATTGATGTAGGAATGTACAGACTTA	2520
2521	AATGAGGACCTAGGTGTAGGGTATGGTATCTTAAGATTAAGA	2560
2561	CCCAAGATGGGCACCCAAGACTAGGGATCTAGAGTTCT	2600
2601	CGAAGAGAAAACCATTAGTAGGGAGAAGCGCTAGCTCGTG	2640
2641	AAAAGAGCGGAGAAAAAATGGAGAGACAAACGTGAAAAAT	2680
2681	TGGAATGGGAAACAAATATCGTTATAAAGAGGGCAAAAGA	2720
2721	ATCTGTAGATGCTTTATTTGTAACCTCTCAATATGATCRA	2760
2761	TTACAAAGCGGATACGAATATTGCCATGATTCAATGCGGCAG	2800
2801	ATAAACGTGTTCATAGCATTGAGAAGCTTATCTGCCTGA	2840

5	2841	GCTGCTGTGATTCCGGGTGTCAATGCGGCTATTTTGA	2880
10	2881	GAATTAGAAGGGCTATTTCACTGCATTCTCCCTATATG	2920
15	2921	ATGCGAGAAAATGTCATTAATGGTGAATTAATAATGG	2960
20	2961	CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA	3000
25	3001	GAACAAAACAACCAACGTTGGTCCITGTTGTTCCGGAAAT	3040
30	3041	GGGAAGCAGAAGTGTCAACAAGAAGTTCTGTCTGTCCGGG	3080
35	3081	TCGTGGCTATATCCTTCGTGTACAGCGTACAAGGAGGGA	3120
40	3121	TATGGAGAAGGTTGGTAACCATTCAATGAGATCGAGAACAA	3160
45	3161	ATACAGACGAACCTGAAGTTTAGCAACTGCGTAGAAGAGGA	3200
50	3201	AATCTATCCRAATAACACGGTAACGGTAACTGTAATGATTAACT	3240
55	3241	GTAATCAAGAAGAATACGGAGGTGCGTACACTTCTCGTA	3280
	3281	ATCGAGGATATAACCAAGGCTCCTCCGTACCGCTGATTAA	3320
	3321	TGGGTCACTCTATGAAGAAAATCGTATACAGATGGACCA	3360
	3361	AGAGAGAATCCTTGTGAATTAAACAGAGGGTATAGGGATT	3400
	3401	ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA	3440
	3441	ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
	3481	GAAACGGAAGGAACATTATCGGGACAGCGTGGAAATTAC	3520
	3521	TCCTTATGGAGGAA	3534

G. Un gène de structure codant une protéine insecticide en pleine longueur de *B.t.k. HD-73* comportant la séquence :

5	1	ATGGACAACAAACCCAAACATCAACGAAATGCATTCCATAACA	40
10	41	ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA	80
15	81	ACGCATTGAAACCGGTTACACTCCCACATCGACATCTCCCTTG	120
20	121	TCCTTGACACAGTTCTGCTCAGGGAGTTCGTGCCAGGTG	160
25	161	CTGGGTTCGTTCTCGGACTAGTTGACATCATCTGGGTAT	200
30	201	CTTTGGTCCATCTCAATGGGATGCATTCTGGTGCATT	240
35	241	GAGCAGTTGATCAACCCAGAGGGATCGAAGAGTTGCCAGGA	280
40	281	ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA	320
45	321	CCAAATCTATGCAGAGAGCTTCAGAGAGTGGGAAGCCGAT	360
50	361	CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT	400
55	401	TCAACGACATGAACAGGCCTTGACCACAGCTATCCCATT	440
60	441	GTTCGCAGTCCAGAACTACCAAGTCCCTCTCTTGTCCGTG	480
65	481	TACGTTCAAGCAGCTAACTTCACCTCAGCGTGCTTCGAG	520
70	521	ACGTTAGCGTGTGGGCAAAGGTGGGGATTGATGCTGC	560
75	561	AACCATCAATAGCCGTTACAACGACCTTACTAGGCTGATT	600
80	601	GGAAACTACACCGACACGCTGTTCTGGTACAAACACTG	640
85	641	GCTTGGAGCGTGTCTGGGTCTGATTCTAGAGATTGGAT	680

5	681	TAGATACAACCAGTTCAAGGAGAGAATTGACCCCTCACAGTT	720
10	721	TTGGACATGGTCTCTCTTCCCGAACTATGACTCCAGAA	760
15	761	CCTACCCCTATCCGTACAGTGTCCCCACTTACCCAGAGAAAT	800
20	801	CTATACTAACCCAGTCTTGAGAACTTCGACGGTAGCTTC	840
25	841	CGTGGTTCTGCCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
30	881	CACACTTGATGGACATCTTGAAACGACATAACTATCTACAC	920
35	921	CGATGCTCACAGAGGAGAGTATTACTGGTCTGGACACCAAG	960
40	961	ATCATGGCCTCTCCAGTTGGATTCAAGCGGGCCCGAGTTTA	1000
45	1001	CCTTTCTCTCTATGGAACATATGGAAACGCCGCTCCACA	1040
50	1041	ACAACGTATCGTTGCTCAACTAGGTCAAGGTGTCACAGA	1080
55	1081	ACCTTGCTTCCACCTTGACAGAAGACCCCTCAATATCG	1120
60	1121	GTATCAACAAACCGCAACTTCCGTTCTGACGGAACAGA	1160
65	1161	GTTCCGCTATGGAACCTCTTCTAACTTGCCATCCGCTGTT	1200
70	1201	TACAGAAAGAGCGGAACCGTTGATTCTTGGACGAATCC	1240
75	1241	CACCAACGAAACACATGTGCCACCCAGGCAAGGATTCTC	1280
80	1281	CCACAGGTTGAGCCACGGTCCATGTTCCGTTCCGGATTC	1320
85	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
90	1361	CTTGGATAACCCGTAGTGTGAGTTCAACAAACATCATCGC	1400
95	1401	ATCCGATAGTATTACTCAAATCCCTGCAGTGAGGGAAAC	1440

1441	TTTCTCTCAACGGTTCTGTCATTCAGGACCAAGGATTCA	1480
1481	CTGGTGGAGACCTCGTTAGACTCAACAGCAGTGGAAATAA	1520
1521	CAATTAGAAATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
1561	CCATCCACATCTACCAGATATAGAGTTCGTGTGAGGTATG	1600
1601	CTTCTGTGACCCCTATTCACCTCAACGTTAATTGGGTAA	1640
1641	TTCATCCATCTCTCCAATACAGTTCCAGCTACAGCTACC	1680
1681	TCCTTGGATAATCTCCAATCCAGCGATTTCGGTTACTTTG	1720
1721	AAAGTCCCAATGCTTTACATCTTCACTCGGTAAACATCGT	1760
1761	GGGTGTTAGAAACTTTAGTGGGACTGCAGGAGTGATTATC	1800
1801	GACAGATTGAGTTCACTCCAGTTACTGCAACACTCGAGG	1840
1841	CTGAGTACAACCTTGAGAGAGGCCAGAAGGCTGTGAACCC	1880
1881	CCTCTTACCTCCACCAATCAGCTTGGCTTGAAAACATAC	1920
1921	GTTACTGACTATCACATTGACCAAGTGTCCAACTTGGTCA	1960
1961	CCTACCTTAGCGATGAGTTCTGCCTCGACGAGAACCGTGA	2000
2001	ACTCTCCGAGAAAGTTAACACCGCCAAAGCGTCTCAGCGAC	2040
2041	GAGAGGAATCTCTTGCAGACTCCAACATTCAAAGACATCA	2080
2081	ACAGGCAGCCAGAACGTTGGGGTGGAAAGCACCGGGAT	2120
2121	CACCATCCAAGGAGGCGACGATGTGTTCAAGGAGAACTAC	2160
2161	GTCACCCCTCTCCGGAACCTTGCACGAGTGCTACCCCTACCT	2200

5	2201	ACTTGTACCAAGAGATCGATGAGTCCAAGACTCAAAGCCTT	2240
10	2241	CACCAAGGTATCAACTTGTAGAGGCTACATCGAAGACAGCCAA	2280
15	2281	GACCTTGAAATCTACTCGATCAGGTACAATGCCAAGCAGCAGG	2320
20	2321	AGACCGTGAATGTCCCAGGTACTGGGTCCTCTGGCCACT	2360
25	2361	TTCTGCCCAATCTCCCATTGGGAAGTGTGGAGAGCCTAAC	2400
30	2401	AGATCGCGCTCCACCCCTTGAGTGGAAATCCTGACTTGGACT	2440
35	2441	GCTCCCTGCAGGGATGGCCAGAACTGTGCCACCATTCTCA	2480
40	2481	TCACTTCTCCCTGGACATCGATGTGGGATGTACTGACCTG	2520
45	2521	AATGAGGACCTCGGAGTCGGGTCACTTCAGATCAAGA	2560
50	2561	CCCAAGACCGACRCGCRAGACTTGGCAACCTTGAGTTCT	2600
55	2601	CGAAGAGAAAACCATTGGTCGGTGAAGCTCTCGCTCGTGTG	2640
60	2641	AAGAGAGCAGAGAAGAAGTGGAGGGACAAACGTGAGAAC	2680
65	2681	TCGAATGGGAAACTAACATCGTTTACAAGGAGGCCAAAGA	2720
70	2721	GTCCGTGGATGCTTGTTCGTGAACCTCCAAATATGATCAG	2760
75	2761	TTGCAAGCCGACACCAACATCGCCATGATCCACGCCAG	2800
80	2801	ACAAACGTGTGCACAGCATTCTGTGAGGGCTACTTGCTGA	2840
85	2841	GTTGTCCGTGATCCCTGGTGTGAACCGCTGCCATCTCGAG	2880
90	2881	GAACCTGAGGGACGTATCTTACCCATTCTCCTGTACGG	2920
95	2921	ATGCCAGAAACGTCAAGAACCGGTGACTTCACAATGG	2960

5 2961 CCTCAGCTGCTGGAATGTGAAAGGTCACTGGACGTGGAG 3000  
 3001 GAACAGAACAAATCAGCGTCCGTCCTGGTTGTGCCTGAGT 3040  
 10 3041 GGGAAAGCTGAAGTGTCCCAAGAGGTTAGAGTCTGTCCAGG 3080  
 3081 TAGAGGCTACATTCTCCGTGTGACCGCTTACAAAGGAGGGA 3120  
 15 3121 TACGGTGAGGGTTGCGTGACCATCCACGAGATCGAGAAC 3160  
 20 3161 ACACCGACGAGCTTAAGTTCTCCAACACTGCGTCGAGGAAGA 3200  
 3201 AATCTATCCAACACACCCGTTACTTGCAACGACTACACT 3240  
 25 3241 GTGAATCAGGAAAGAGTACGGAGGTGCGCTACACTAGCCGTA 3280  
 3281 ACAGAGGTTACACGAAAGCTCCTTCCGTTCCGTGACTA 3320  
 3321 TGCCTCCGTGTACGGAGGAGAAATCCTACACAGATGGCAGA 3360  
 3361 CGTGAGAACCCCTTGCAGTTAACACGAGGTACAGGGACT 3400  
 3401 ACACACCACTTCCAGTTGGCTATGTTACCAAGGAGCTTGA 3440  
 40 3441 GTACTTTCCGTGAGACCGACAAAGTGTGGATCGAGATCGGT 3480  
 3481 GAAACCGAGGGACCTTCATCGTGGACAGCGTGGAGCTTC 3520  
 45 3521 TCTTGATGGAGGAA 3534

50 H. Un gène de structure qui code une protéine insecticide de *B.t.t.* Comportant La séquence :

1 ATGACTGCAGACAACAACACCGAAGCCCTCGACAGTTCTA 40  
 5  
 41 CCACTAAGGATGTTATCCAGAAGGGTATCTCCGGTGTGGG 80  
 10  
 81 AGACCTCTGGCGTGGTGGATTTCCTCGGTGGAGCC 120  
 15  
 121 CTCGTGAGCTTCTATACAAACTTCTCAACACCAATTGGC 160  
 161 CAAGCGAGGACCCCTTGGAAAGCATTCAATGGAGCAAGTTGA 200  
 20  
 201 AGCTCTTATGGATCAGAAGATTGCAGATTATGCCAAGAAC 240  
 25  
 241 AAGGCTTGGCAGAACTCCAGGGCCTTCAGAACAAATGTGG 280  
 281 AGGACTACGTGAGTCATTGTCAGCTGGCAGAAAGAACCC 320  
 30  
 321 TGTAGCTCCAGAAATCCTCACACCCAGGTAGGATCAGA 360  
 361 GAGTTGTTCTCTCAAGCGGAATCCCACCTTCAGAAATTCCA 400  
 35  
 401 TGCCTAGCTTIGCTATCTCCGGTTACGAGGTTCTTTCT 440  
 441 CACTACCTATGCTCAAGCTGCCAACACCCACTTGTTCCTC 480  
 40  
 481 CTTAAGGACGCTCAAATCTATGGAGAAGAGTGGGGATACG 520  
 45  
 521 AGAAAGAGGACATTGCTGAGTTCTACACCGTCACCTTAA 560  
 561 GCTCACCCAAAGAGTACACTGACCATTGCGTGAAATGGTAT 600  
 50  
 601 AACGTTGGTCTCGATAAGCTCAGAGGCTCTTCCTACGGAT 640  
 55  
 641 CTTGGGTGAACCTCAACAGATAACAGGAGAGATGACCTT 680

5	681	GACTGTGCTCGATCTTATGGCACTCTTCCCTTGTACGGAT	720
10	721	GTGAGACTCTACCCAAAGGAAGTGAAGAACTGAGCTTACCA	760
15	761	GAGACGTGCTCACTGACCCATTGTCGGAGTCACAAACCT	800
20	801	TAGGGGTTATGGAACTACCTTCAGCAATATCGAAAACCTAC	840
25	841	ATTAGGAACCACATCTTCGACTATCTCACAGAACATT	880
30	881	AATTCCACACAGGTTCAACCAGGATACTATGGTAACGA	920
35	921	CTCCTTCACACTATTGGTCGGTAACATATGTTCCACCAAGA	960
40	961	CCAAGCATTGGATCTAATGACATCATCACATCTCCCTCT	1000
45	1001	ATGGTAACAGTCAGTGAACCTGTGCAGAACCTTGAGTT	1040
50	1041	CAACGGCGAGAAGTCTATAGAGCCGTGCAACACCAAT	1080
55	1081	CTCGCTGTGGCCATCCGCAGTTACTCAGGGTCACAA	1120
60	1121	AGGTGGAGTTAGTCAGTATAACGATCAGACCGATGAGGC	1160
65	1161	CAGCACCCAGACTTACGACTCCAAACGTAACGGTGGCGCA	1200
70	1201	GTCTCTGGGATTCATGACCAATTGCCCTCAGAAACCA	1240
75	1241	CAGACGAACCATGGAGAAGGGCTACAGCCACCACTTAA	1280
80	1281	CTATGTGATGTGCTTCTGATGCAAGGTTCCAGAGGGACC	1320
85	1321	ATTCCAGTGTGACCTGGACACACAAGTCCGTGGACTTCT	1360
90	1361	TCAACATGATCGATAGCAAGAAGATCACTCAACTTCCCT	1400
95	1401	GGTGAAGCCTACAGCTGCAATCTGGTGCTTCCGGTGT	1440

5 1441 GCGGGTCCCAGATTCACTGGAGGTGACATCATCCAGTGCA 1480  
 10 1481 CAGAGAACGGCAGCCAGCTACTATCTACGTGACACCTGA 1520  
 15 1521 TGTGTCTTACTCTCAGAAGTACAGGGCACGTATTCAATTAC 1560  
 1561 GCATCTACCAGCCAGATCACCTTCACACTCAGCTTGGATG 1600  
 20 1601 GAGCACCCCTCAACCACTATTACTTTGACAAGACCATCAA 1640  
 25 1641 CAAAGGTGACACTCTCACATACAATAGCTTCACCTTGGCA 1680  
 1681 AGTTTCAGCACACCCATTGAACTCTCAGGCCACAATCTTC 1720  
 30 1721 AGATCGGGGTACCCGGTCTCAGGCCGGAGACAAAGTCTA 1760  
 1761 CATGACAAGATTGAGTTCATCCCAGTGAAC 1791

35 I. Un gène de structure qui code une protéine insecticide de *B.t. entomocidus* comportant la séquence :

40

45

50

55

1	ATGGAGGAGAACAAACCAAAAACCAATGCATTCCATACAACT	40
5		
41	GCTTGAGTAACCCAGAAGAGGTATTGCTTGATGGAGAACG	80
10		
81	CATTCAACCGGTAACCTCTTCCATCGACATCTCCTTGTCC	120
15		
121	TTGGTCCAGTTCTGGTCAGCAACTTCGTGCCAGGTGGTG	160
20		
161	GGTTCCCTTGTCCGGACTAATTGACTTCGTCTGGGTATCGT	200
25		
201	TGGTCCATCTCAATGGGATGCATTCTGGTGCAATTGAG	240
30		
241	CAGTTGATCAACGAGAGGATCGCTGAGTCGCCAGGAACG	280
35		
281	CTGCCATCGCTAACCTTGGAGGGATTGGCAATAACTTCAA	320
40		
321	CATCTATGTGGAGGCCCTTCAAAGAGTGGAAGAGGGACCT	360
45		
361	AAACAACCCAGAGACCCGCACTAGGGTGTGACAGATTCA	400
50		
401	GAATCTTGGACGGCCTCTTGGAGAGAGATATCCCATCCTT	440
55		
441	CAGAAATCTCTGGCTTCGAAGTTCCCTCTTGTCCGTGTAC	480

481	GCTCAAGCAGCTAATCTTCACCTCGCTATCCTCGAGACA	520
521	GTGTCACTTTGGGAAAGGTGGGATGACCACTATCAA	560
561	CGTCAATGAGAATTACAACAGACTTATCAGGCACATTGAC	600
601	GAGTACGCCGACCACTGTGCTAACACCTACAAACCGTGGCT	640
641	TGAAACAATCTCCCTAAGTCTACTTATCAAGATTGGATTAC	680
681	CTACAAACAGGTTGAGGAGAGACTTGACCCCTCACAGTTTG	720
721	GACATTGCACTTCTCCCAACTATGACAACAGGAGAT	760
761	ACCCATCCAACCACTGGGTCAACTTACCAAGAGACTCTA	800
801	TACTGACCCACTTATCAACTTCAACCCCTCAGTTGCAAAGT	840
841	GTCGCCCAACTTCCACATTCAACGTCAATGGAGTCCAGCC	880
881	GTATCAGGAACCCACACTTGGTTGACATCTTGAACAACCT	920
921	TACTATCTTCACCGATTGGTCAGCGTIGGGCGTAACCTTC	960
961	TATTGGGGTGGACACAGGGTCATCTCTCTCTTATTGGAG	1000
1001	GTGGGAACATTACCTCTCCTATCTATGGACGTGAGGCCAA	1040
1041	CCAGGAGCCACCACCGTAGTTCACCTCAACGGTCCAGTC	1080
1081	TTCAAGAACCTTGTCTAACCCCTACCTTGAGATTGCTCCAGC	1120
1121	AACCTTGGCCAGCTCCACCTTCAACCTTAGAGGTGTTGA	1160
1161	GGGCCTTGAGTTCTCTACTCCTACCAACTCCCTTCACTTAC	1200
1201	AGAGGTAGAGGAACCGTTGATTCTTGACCGAACCTCCAC	1240

1241	CAGAGGACAAATAGGGTGCCACCCAGGGAAAGGGCTACTCCCA	1280
1281	CAGGGTTGTGCCACCGAACCTTCGTGCAGCGTTCCCGAACT	1320
1321	CCATTCCCTCACTACAGGGAGTTGTGTTCTCATGGACTGATC	1360
1361	GTAGTGCTACTCTCACTAAATACCAATTGATCCCGAGAGGAT	1400
1401	CAATCAAATCCCAATTGGTCAAGGGTTCCGTGTGIGGGGA	1440
1441	GGAACCTCTGTCAATCACAGGACCAGGCTTCACAGGAGGTG	1480
1481	ATATTCTTAGAAGAACACTTTGGCAGCTTGTGAGCCT	1520
1521	CCAAAGTTAACATCAACTCTCCAATTACTCAAAGATATCGT	1560
1561	CTCAGGGTTCGTTACGCATCTCCCGTGACGCTAGAGTCA	1600
1601	TCGTGCTCAGCGGAGCAGCTTCTACCGGTGTCGGTGGACA	1640
1641	AGTCTCCGTGAACATGCCACTCCAGAAAGACTATGGAGATC	1680
1681	GGCGAGAACTTGACATCCAGGACCTTCAGATAACACCGACT	1720
1721	TCTCTAACCTTTCACTTCCGTGCCAACCTGACATCAT	1760
1761	TGGCATTAGCGAACAAACCTCTCTTGGAGCTGGTAGCATC	1800
1801	TCATCTGGCGAATTGTACATTGACAAGAATTGAGATCATTC	1840
1841	TTGCCGACGCTACCTTCGAGGCTGAGTCTGACCTTGAGAG	1880
1881	AGCCCAGAAGGCTGTGAACGCCCTCTTACCTCCCTAAAT	1920
1921	CAGATTGGCTTGAAACTGACGTTACTGACTATCACATTG	1960
1961	ACCAAGTGTCCAACCTGGTCACTGACGCTAGCGATGAGTT	2000

2001	CTGCCTCGACGAGARGCGTGAACCTCTCCGAGAAAGTTAAA	2040
2041	CACGCCAAGCGTCTCAGCGACGGAGAGGAATCTCTTGCAAG	2080
2081	ACCCCAACTTCAGAGGCATCAACAGGCAGCCAGACCGTGG	2120
2121	TTGGAGAGGAAGCACCACATCACCACATCCAAGGAGGGCGAC	2160
2161	GATGTGTTCAAGGAGAACTACGTACCCCTCCCAGGAACGTG	2200
2201	TGGACGAGTGCTACCCCTACCTACTGTACCAAGAGATCGA	2240
2241	TGAGTCCAAACTCAAAGCCTACACCAGGTATGAACCTTAA	2280
2281	GGCTACATCGAAGACAGCCAAGACCTTGAAATCTACCTCA	2320
2321	TCAGGTACAATGCCAAGCACGAGATCGTAATGTCCCAGG	2360
2361	TACTGGTCCCTCTGGCCACTTCTGCCAAATGCCATT	2400
2401	GGGAAAGTGTGGAGAGCCTAACAGATGCCCTCACACCTTG	2440
2441	AGTGGAAATCCTGACTTGGACTGCTCCTGCAGGGATGGCGA	2480
2481	GAAGTGTGCCACCATTCTCATCACTTCACCTTGACATC	2520
2521	GATGTGGGATGTACTGACCTGAATGAGGACCTCGGAGTCT	2560
2561	GGGTCACTTCAAGATCAAGACCCAAGACGGACACGCAAG	2600
2601	ACTTGGCAACCTTGAGTTCTCGAAGAGAAACCATTGCTC	2640
2641	GGTGAAGCTCTCGCTCGTGTGAAGAGAGCAGAGAAGT	2680
2681	GGAGGGACAAACGTGAGAACTCCAACCTCGAGACTAACAT	2720
2721	CGTTTACAGGAGGCCAAGAGTCCGTGGATGCTTGTTC	2760

5 2761 GTGAACCTCCAATATGATAGGTTGCAAGTGGACCCAACA 2800  
 10 2801 TCGCCATGATCCACGGCTGCAGACAAACGTGTGACAGGAT 2840  
 15 2841 TCGTGAGGCTTACTTGCTGAGTTGTCCGTGATCCCTGGT 2880  
 20 2881 GTGAACGCTGCCATCTCGAGGAACCTGAGGGACGTTATCT 2920  
 25 2921 TTACCGCATACTCCTGTACCGATGCCAGAAACGTCAAA 2960  
 30 2961 GAAACGGTGAACCTCAACAATGGCCTCTTGTGCTGGAATGTG 3000  
 3001 AAAGGTCAATGTCGGACGGTGGAGGAACAGAACAAATCACCGTT 3040  
 3041 CCGTCCTGGTTATCCCTGAGTGGGARGCTGAAGTGTCCCCA 3080  
 3081 AGAGGTTAGAGTCTGTCCAGGTAGAGGCTACATTCTCCGT 3120  
 3121 GTGACCGCTTACAAGGAGGGATACGGTGGAGGGTTGCGTGA 3160  
 3161 CCATCCACGAGATCGAGGAACAACACCGACGGAGCTTAAGTT 3200  
 3201 CTCCAACACTGCGTCCAGGAAGAAGTCTATCCCAACACACC 3240  
 3241 GTTACTTGCAACAACACTACACTGGGACCCAGGAAGAGTACG 3280  
 3281 AAGGTACCTACACTAGCCGTAACCAAGGTTACGACGAAAGC 3320  
 3321 TTACGGAAACAATCCTCCGTTCTCTGACTATGCCCTCC 3360  
 3361 GTGTACGGAGGAGAAATCCTACACAGATGGCAGACGTGAGA 3400  
 3401 ACCCTTGCGAGTCCAACAGAGGTTACGGTACTACACACC 3440  
 3441 ACTTCCAGCAGGCTATGTTACCAAGGACCTTGAGTACTTT 3480  
 3481 CCTGAGACCGACAAAGTGTGGATCGAGATCGGTGAAACCG 3520

3521 AGGGAACCTTCATCGTGGACAGCGTGGAGCTCTCTTGAT 3560

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3561 GGAGGAA 3567.

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J. Un gène de structure qui code - une protéine insecticide P2 comportant la séquence :

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1 ATGGACAACAAACGTCTTGAACCTCTGGTAGAACAAACCATCT 40  
 5  
 41 GCGACGCAATACAACGTCGTGGCTCACGATCCATTCAAGCTT 80  
 10  
 81 CGAACACAAGAGCCTCGACACTATTCAAGGAGTGGATG 120  
 15  
 121 GAATGGAAACGTACTGACCACTCTCTACGTGCCACCTG 160  
 20  
 161 TGGTTGGAACAGTGTCCAGCTTCTCTCAAGAAGGTGG 200  
 25  
 201 CTCTCTCATCGAAAACGTATCTGTCCGAACTCTGGGGT 240  
 30  
 241 ATCATCTTCCATCTGGGTCCACTAAATCTCATGCAAGACA 280  
 35  
 281 TCTTGAGGGAGACCGAACAGTTCTCAACCAGCGTCTCAA 320  
 40  
 321 CACTGAAACCTGGCTAGAGTCACCGCTGAGTTGATCGGT 360  
 45  
 361 CTCCAAGCRAAACATTGTGAGTTCAACCCAGCAAGTGGACA 400  
 50  
 401 ACTTCTGAACTCAACACTAGAAATCTGTGCCCTTTCCAT 440  
 55  
 441 CACTTCTCCGTAAACACTATGCAAGCAACTCTTCTCAAC 480  
 481 AGATTGCCCTCAGTTCAAGATTCAAGGCTACCGATTGCTCC 520  
 521 TTCTTCCACTCTTGCTCAGGCTGCCAACATGCACTTGTC 560  
 561 CTTCATACGTGACGTGATCCTCAACGCTGACGAAATGGGGA 600

601	ATCTCTGCAGCCACTCTTAGGACATACAGAGACTACTTGA	640
641	GGAACATACACTCGTGATTACTCCAACTATTCATCAACAC	680
681	TTATCAGACTGCCTTCGTGGACTCAATACTAGGTTCAC	720
721	GACATGCTTGAGTTAGGACCTACATGTTCTTAACGTGT	760
761	TTGAGTACGTCAGCAATTGGAGTCTCTTCAGTACAGCAGAG	800
801	CTTGATGGTGTCCCTGGAGCCAAATCTACGCGCTCTGGC	840
841	AGTGGACCACAGCAAACCTCAGAGCTTACAGCTCAGAACT	880
881	GGCCATTCTGTATAGCTTGTCCAAAGTCAACTCCAACTA	920
921	CATTCTCAGTGGTATCTCTGGGACCAAGACTCTCCATAACC	960
961	TTTCCCAACATTGGTGGACTTCCAGGCTCCACTACAAACCC	1000
1001	ATAGCCTTAACCTGCCAGAGTGAACACTACAGTGGAGGTGT	1040
1041	CAGCTCTGGATTGATTGGTGCACACTAACTTGAACCCACAAAC	1080
1081	TTCAATTGCTCCACCGCTTGCACCTCTGAGCACACCGT	1120
1121	TTGTGAGGTCTGGCTTGACAGCGGTACTGATCGCGAAGG	1160
1161	AGTTGCTACCTCTACAAACTGGCAAACCGAGTCCTTCAA	1200
1201	ACCACTCTTAGCCTCGGTGTGGAGCTTCTCTGCACGTG	1240
1241	GGAATTCAAACACTTCCAGACTACTTCATTAGGAACAT	1280
1281	CTCTGGTGTCCCTCTCGTCATCAGGAATGAAGACCTCACC	1320
1321	CGTCCACTTCATTACAAACAGATTAGGAACATCGAGTC	1360

1361	CATCCGGTACTCCAGGAGGTGCAAGAGCTTACCTCGTGTC	1400
1401	TGTCCATAACAGGAAGAACAAACATCTACGCTGCCAACGAG	1440
1441	AATGGCACCCTGATTCACCTTGCACCAGAAGATTACACTG	1480
1481	GATTCACCACTCTCCAATCCATGCTACCCAAGTGAACAA	1520
1521	TCAGACACGGCACCTTCATCTCCGAAAAGTTCGGAAATCAA	1560
1561	GGTGACTCCTTGAGGTTCGAGCAATCCACACTACCGCTA	1600
1601	GGTACACTTGTAGAGGGCAATGGAAACAGCTACAACTTTA	1640
1641	CTTGAGAGTTAGCTCATTGGTAACCTCCACCATCCGTGTT	1680
1681	ACCATCAACGGACGTGTTACACAGTCTAAATGTGAACA	1720
1721	CTACAACGARAAATGATGGCGTTAACGACAACGGAGCCAG	1760
1761	ATTCAAGGACATCAACATTGGCAACATCGTGGCTCTGAC	1800
1801	AAACACTAACGTTACTTTGGACATCAATGTGACCCCTCAATT	1840
1841	CTGGAACTCCATTGATCTCATGAACATCATGTTGTGCC	1880
1881	AACTAACCTCCCTCCATTGTAC	1902

K. Une séquence de gène de structure codant une protéine de fusion comprenant les acides aminés 610 N-terminaux de *B.t.k.* HD-1 et les acides aminés 587 C-terminaux de *B.t.k.* HD-73, ledit gène comportant la séquence :

1 ATGGACAACAACCCAAACATCAACGAATGCATTCCATACA 40

5

41 ACTGCTTGAGTAACCCAGAAGTTGAAGTACTTGGTGGAGA 80

10 81 ACGCATTGAAACCGGTTACACTCCCATCGACATCTCCCTG 120

15 121 TCCTTGACACAGTTCTGCTCAGCGAGTTCGTGCAGGTG 160

161 CTGGGTTCTCGGACTAGTTGACATCATCTGGGTAT 200

20 201 CTTTGGTCCATCTCAATGGGATGCATTCCCTGGTGCAAATT 240

241 GAGCAGTTGATCAACCAGAGGATCGAAGAGTTGCCAGGA 280

25 281 ACCAGGCCATCTCTAGGTTGGAAGGATTGAGCAATCTCTA 320

321 CCAAATCTATGCAGAGAGCTTCAGAGAGTGAAAAGCCGAT 360

30 361 CCTACTAACCCAGCTCTCCGCGAGGAAATGCGTATTCAAT 400

35 401 TCAACGACATGAACAGCGCCATTGACCACAGCTATCCCATT 440

40

45

50

55

441 GTTCCAGTCCAGAACTACCAAGTTCCCTCTCTTGTCCGTG 480

5 481 TACGTTCAAGCAGCTAATCTTCACCTCAGCGTGGCTTCGAG 520

10 521 ACGTTAGCGTGTGGCAAAGGTGGGATTGATGCTGC 560

15 561 AACCATCAATAGCCGTTACAACGACCTTACTAAGGCTGATT 600

20 601 GGAAACTACACCGACCACGCTGTTGGTACAACACTG 640

25 641 GCTTGGACCGGTGCTGGGGCTGATTCTAGAGATTGGAT 680

30 681 TAGATACAACCAGTTCAAGGAGAGAATTGACCCCTCACAGTT 720

35 721 TTGGACATTGTGTCTCTCTCCGAACTATGACTCCAGAA 760

40 761 CCTACCCCTATCCGTACAGTGTCCCAACTTACCRAGAGAATT 800

45

50

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5	801	CTATACTAACCCAGTTCTTGAGAACTTCGACGGTAGCTTC	840
10	841	CGTGGITCTGCCAAGGTATCGAAGGCTCCATCAGGAGCC	880
15	881	CACACTTGATGGACATCTGAAACAGCATAACTATCTACAC	920
20	921	CGATGCTCACAGAGGAGAGTATTACTGGCTGGACRCCAG	960
25	961	ATCATGGCCTCTCCAGTTGGATTAGCGGGCCCGAGTTA	1000
30	1001	CCTTCCCTCATGGAACATATGGAAACGCCGCTCCACA	1040
35	1041	ACAACGTATGTTGCTCAACTAGGTCAAGGTGTACAGA	1080
40	1081	ACCTTGTCTTCCACCTTGTACAGAAGACCCCTCAATATCG	1120
45	1121	GTATCAACAAACCAAGCAACTTTCGGTCTTGACGGAAACAGA	1160
50	1161	GTTCGCCTATGGAACCTCTTCAACTTGCCATCCGTGTT	1200
55	1201	TACAGAAAGAGCGGAACCGTTGATTCCTGGACGAATCC	1240
60	1241	CRCCACAGAACACAAATGTGCCACCCAGGCAAGGATTCTC	1280
65	1281	CCACAGTTGAGCCACGTGTCCATGTTCCGTTCCGATTTC	1320
70	1321	AGCAACAGTTCCGTGAGCATCATCAGAGCTCCTATGTTCT	1360
75	1361	CATGGATTCACTGTAAGTGTGAGTTCAACAAATATCATTCC	1400
80	1401	TTCCCTCTCAAATCACCCAAATCCATTGACCAAGTCTACT	1440
85	1441	AACCTTGGATCTGAAACTTCTGCGTGAAAGGACCGAGGT	1480
90	1481	TCACAGGAGGTGATATTCTTAAAGAAACTTCTCCTGGCCA	1520

1521	GATTAGCACCCCTCAGAGTTAACATCACTGCACCACTTTCT	1560
1561	CAAAAGATATCGTGTCAAGGATTCTGTTACGCATCTACCACTA	1600
1601	ACTTGCAATTCCACACCTCCATCGACGGAAAGGCCTATCAA	1640
1641	TCAGGGTAACTTCTCCGAACCAATGTCAAGCGGCAGCAAC	1680
1681	TTGCAATCCGGCAGCTTCAGAACCGTCGGTTCACTACTC	1720
1721	CTTTCAACTTCTCTAACGGATCAAGCGTTTCACCCCTTAG	1760
1761	CGCTCATGTTCAATTCTGCAATGAAGTGTACATTGAC	1800
1801	CGTATTGAGTTTGCCCTGCCGAAGTTACCCCTGAGGCTG	1840
1841	AGTACAACCTTGAGAGAGGCCAGAAGGCTGTGACGCCCT	1880
1881	CTTACCTCCACCAATCAGCTGGCTTGAAAACTAACGTT	1920
1921	ACTGACTATCACATTGACCAAGTGTCCAACTTGGTCACCT	1960
1961	ACCTTAGCGATGAGTTCTGCCCTCGACGAGAAGCGTGAACI	2000
2001	CTCCGAGAAAGTTAACACGCCAACGGCTCTCGCGACCGAG	2040
2041	AGGAATCTTGCAGACTCCAACCTCAAAAGACATCAACA	2080
2081	GGCACCCAGAACGTGTTGGGTGGAAGCACCGGATCAC	2120
2121	CATCCAAGGAGGCCACGATGTGTCAAGGAGAACATACGTC	2160
2161	ACCCCTCCGGAACCTTCGACCGAGTGTACCCCTACCTACT	2200
2201	TGTACCAAGAGATCGATGAGTCCAACCTCAAGCCTTCAC	2240
2241	CAGGTATCAACTTAGAGGCTACATCGAAGACAGCCAAGAC	2280

2281	CTTGAAATCTACTCGATCAGGTACAATGCCAAGCAGGAGA	2320
2321	CCGTGAATGTCCCAGGTACTGGTTCCCTCTGGCCACTTTC	2360
2361	TGCCCAATCTCCCATTGGGAAGTGTGGAGAGCCTAACAGA	2400
2401	TGCCGCTCCACACCTTGAAGTGGAACTCCTGACTTGGACTGCT	2440
2441	CCTGCAGGGATGGCGAGAAGTGTGCCACCATTCTCATCA	2480
2481	CTTCTCTTGGACATCGATGTGGATGTACTGACCTGAAT	2520
2521	GAGGACCTCGGAGTCTGGTCATCTCAAGATCAAGAACCC	2560
2561	AAGACGGACACGCAAGACTTGGCAACCTTGAAGTTCTCGA	2600
2601	AGAGAAACCATTGGTCGGTGAAGCTCTCGCTCGTGTGAAG	2640
2641	AGACCGAGAGAAGAAGTGGAGGGACRAACGTGAGAAACTCG	2680
2681	AATGGGAAACTAACATCGTTACAAGGAGGCCAAGAGTC	2720
2721	CGTGGATCTTCTTCGTGAACCTCCAAATATGATCAGTTG	2760
2761	CAAGCCGACACCAACATGCCATGATCCACGCCAGACA	2800
2801	AACGTGTCCACACGATTCTGTGAGGCTTACTTGCTGAGTT	2840
2841	GTCCGTGATCCCTGGTGTGAACGCTGCCATCTCGAGGAA	2880
2881	CTTGAGGGACGTATCTTACCGCATCTCCTTGTACGATG	2920
2921	CCAGAAACGTCAAGAACGGTGACTTCAACAAATGGCCT	2960
2961	CAGCTGCTGGAATGTGAAAGGTATGTGGACGTGGAGGAA	3000
3001	CAGAACAAATCAGCGTTCCGTCTGGTTGTGCCTGAGTGGG	3040

5 3041 AAGCTGAAGTGTCCCAGAGGTTAGAGTCTGTCAGGTAG 3080  
 . . . . .  
 10 3081 AGGCTACATTCTCCGTGTGACCGCTTACAAGGAGGATAC 3120  
 . . . . .  
 15 3121 GGTGAGGGTTCCGTGACCATCCACCGAGATCGAGAACACA 3160  
 . . . . .  
 20 3161 CCGACGAGCTTAAGTCTCCAACITCGTCGAGGAAGAAAT 3200  
 . . . . .  
 25 3201 CTATCCAAACACACCGTTACTTGCACCGACTACACTGTG 3240  
 . . . . .  
 30 3241 AATCAGGAAGAGTACGGAGGTGCCTACACTAGCCGTAAAC 3280  
 . . . . .  
 3281 GAGGTTACAAACGAAGCTCCTCCGTTCTGCTGACTATGC 3320  
 . . . . .  
 3321 CTCCGTGTACGGAGGAATCCTACACAGATGGCAGACGT 3360  
 . . . . .  
 3361 GAGAACCCCTTGCAGTTCAACAGAGGTTACAGGGACTACA 3400  
 . . . . .  
 3401 CACCACTTCCAGTTGGCTATGTTACCAAGGAGGTTGAGTA 3440  
 . . . . .  
 3441 CTTTCTGAGACCGACAAGTGTGGATCGAGATCGGTGAA 3480  
 . . . . .  
 3481 ACCGAGGGAACCTTCATCGTGGACAGCGTGGAGCTTCTCT 3520  
 . . . . .  
 40 3521 TGATGGAGGAA 3531.  
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 50 . . . . .  
 55 . . . . .

Determination of DNA regions in genes to modify by  
site-directed mutagenesis for increased expression in plants

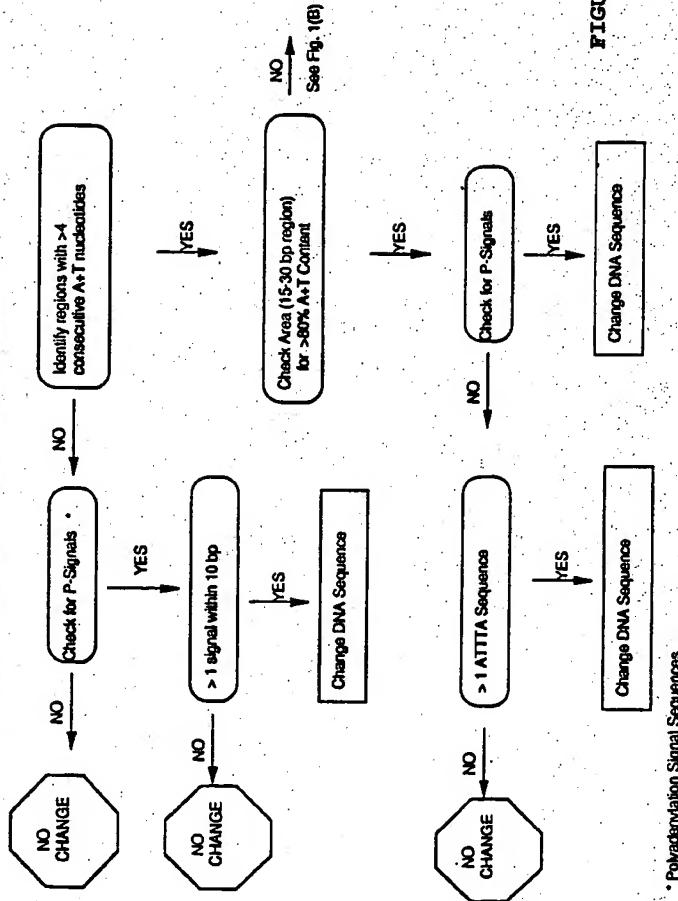
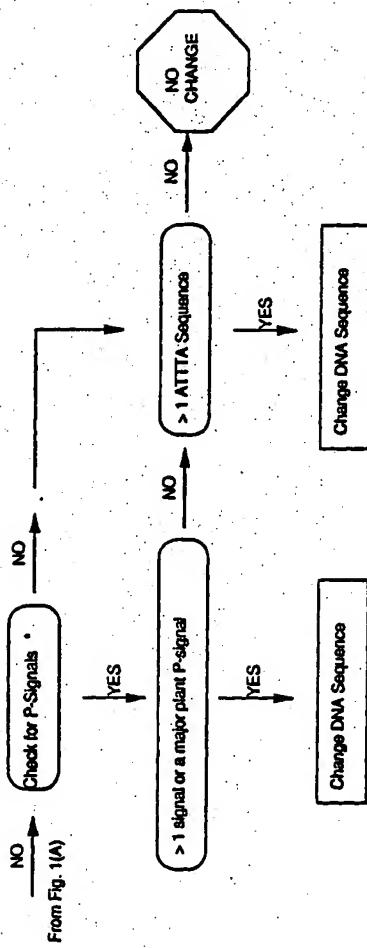


FIGURE 1 (A)

Determination of DNA regions in genes to modify by  
site-directed mutagenesis for increased expression in plants



• Polyadenylation Signal Sequences

FIGURE 1 (B)

1	ATGGCTATAGAAACTGGTTACACCCCAATCGATATTCCT	40
41	TGTCGCTAACGCAATTCTTTGAGTGAATTGTTCCGG	80
81	TGCTGGATTGTGTTAGGACTAGTTGATATAATATGGGA T C	120
121	ATTTTGGTCCCTCTCAATGGGACGCATTCCTGTACAAA	160
161	TTGAACAGTTAATTACCAAAGAATAGAAGAATTGGCTAG C C C G C G	200
201	GAACCAAGCCATTCTAGATTAGAAGGACTAACGAACTTT T	240
241	TATCAAATTACGCAGAACTTTAGAGAGTGGGAAGCAG	280
281	ATCCTACTAATCCAGCATTAGAGAAGAGATGCGTATTCA	320
321	ATTCAATGACATGAACAGTGCCTTACAACCGCTATTCT	360
361	CTTTTGCAAGTCAAATTATCAAGTTCCCTTTTATCAG CC C C	400
401	TATATGTTCAAGCTGCAAATTACATTATCAGTTTGAG G C C C C C C	440
441	AGATGTTCAAGCTGTTGGACAAAGGTGGGATTTGATGCC	480
481	GCGACTATCAATAGTCGTTATAATGATTTAACTAGGCTTA	520
521	TTGGCAACTATACAGATCATGCTGTACGCTGGTACAATAC	560
561	GGGATTAGAGCGTGTATGGGGACCGGATTCTAGAGATTGG	600
601	ATAAGATATAATCAATTAGAAGAGAATTAACACTAACTG C G C C G C G C T	640
641	TATTAAGATATCGTTCTATTTCCGAACTATGATAGTAG	680
681	AACGTATCCAATTGAAACAGTTCCCAATTAAACAAGAGAA	720

FIGURE 2A

721	ATTTATACAAACCCAGTATTAGAAAATTTGATGGTAGTT	760
761	TTCGAGGCTCGGCTCAGGGCATAGAAGGAAGTATTAGGAG	800
801	TCCACATTTGATGGATATACTTAATAGTATAACCATCTAT	840
841	ACGGATGCTCATAGAGGGAGAATATTATGGTCAGGGCATH C C C T C	880
881	AAATAATGGCTTCTCCGTAGGGTTTCGGGCCAGAATT G C	920
921	CACTTTCCGCTATATGGAACTATGGAAATGCAGCTCCA	960
961	CAACAAACGTATTGTTGCTCAACTAGGTCAAGGGCGTGTATA	1000
1001	GAACATTATCGTCCACCTTATATAGAAGACCTTTAATAT C	1040
1041	AGGGATAATAATCAACAACTATCTGTTCTGACGGGACA C C C C	1080
1081	GAATTTGCTTATGGAACCTCCTCAAAATTGCCATCCGCTG	1120
1121	TATACAGAAAAAGCGGAACGGTAGATTGCTGGATGAAAT	1160
1161	ACCGCCACAGAATAACAAACGTGCCACCTAGGCAAGGATTT	1200
1201	AGTCATCGATTAAGCCATGTTCAATGTTCTGAGGCT	1240
1241	TTAGTAATAGTAGTGTAAAGTATAATAAGAGCTCCTATGTT	1280
1281	CTCTTGATAACATCGTAGTGCTGAATTAAATAATATAATT G C C C C C	1320
1321	CCTTCATCACAAATTACACAAATACCTTTAACAAAATCTA C C C AC C C G	1360
1361	CTAATCTGGCTCTGGAACCTCTGTCGTTAARGGACCAGG	1400

FIGURE 2B

1401 ATTTACAGGAGGAGATATTCTCGAAGAACCTCACCTGGC 1440  
 1441 CAGATTCACCTTAAGAGTAAATATTACTGCACCATAT 1480  
 1481 CACAAAGATATCGGGTAAGAATTCGCTACGCTTCTACCAC 1520  
 1521 AAATTTACAATTCCATACATCAATTGACGGAAGACCTATT 1560  
 CC T G C  
 1561 AATCAGGGATTTCAGCAACTATGAGTAGTGGAGTA 1600  
 1601 ATTTACAGTCCGGAAAGCTTTAGGACTGTAGGTTTACTAC 1640  
 1641 TCCGTTAACCTTCAAATGGATCAAGTGATTTACGTTA 1680  
 1681 AGTGCTCATGTCTCAATTCAAGGCAATGAAGTTATAG 1720  
 1721 ATCGAATTGAATTGTTCCGGCA 1743

FIGURE 2C

1	ATGGATAACAATCCGAAACATCAATGAATGCATTCTTATA	40
	C C A C A C	
41	ATTGTTAACGTAACCCCTGAAGTAGAAGTATTAGGGAGA	80
	C C G A T C T	
81	AAGAATAGAAACTGGTTACACCCCCAATCGATATTCCTTG	120
	C C T C T C C C	
121	TCGCTAACGCAATTCTTTGAGTGAAATTGTTCCGGTG	160
	CT G A G G C C C G C G A	
161	CTGGATTGTGTTAGGACTAGTTGATATAATATGGGAAT	200
	G C T C C C C C C T	
201	TTTGCTCCCTCTCAATGGGACGCATTCTTGATCAAATT	240
	C A T C G G G	
241	GAACAGTTAACCAACAAAGAATAGAAGAATTGGCTAGGA	280
	G G C G G G C G C	
281	ACCAAGCCATTCTAGATTAGAAGGACTAAGCAATTTA	320
	G C G G T G C	
321	TCAAATTACGCGAGAATCTTTAGAGAGTGGGAAGCAGAT	360
	C C T GAGC C C	
361	CCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCAAT	400
	C T C C C G A	
401	TCAATGACATGAACAGTGCCTTACAACCGCTATTCTCT	440
	C C T G C A C AT	
441	TTTGCGAGTCAAAATTATCAAGTTCTCTTTATCAGTA	480
	G C C G C C C G G C G	
481	TATGTTCAAGCTGAAATTACATTATCAGTTTGAGAG	520
	C A T C T C C C A G C G C	
521	ATGTTTCAGTGTGGACAAAGGTGGGATTGATGCCGC	560
	C A G C G C T	
561	GACTATCAATAGTCGTATAATGATTAACTAGGCTTATT	600
	A C C C C C C T G	
601	GGCAACTATACAGATCATGCTGTaCGCTGGTACAATACGG	640
	A C C C C T T C T	
641	GATTAGAGCGTGTATGGGGACCGGATTCTAGAGATTGGAT	680
	C G C T T	

FIGURE 3A

681	AAGATATAATCAATTAGAAGAGAATTAACACTAACTGTA	720
	T C C G C G G C C A T	
721	TTAGATATCGTTCTCTATTTCGGAACTATGATAGTAGAA	760
	G C T G C C C T C C	
761	CGTATCCAATTGAAACAGTTCCCAATTAAACAGAGAAAT	800
	C C T C T G C T C	
801	TTATACAAACCCAGTATTAGAAAATTGATGGTAGTTT	840
	C T T C T G C C C C	
841	CGAGGCTCGGCTCAGGGCATAGAAGGAAGTATTAGGAGTC	880
	T T C A T C C T C C	
881	CACATTGATGGATATACTTAATAGTATAACCATCTATAC	920
	C C C T G C C T C	
921	GGATGCTCATAGAGGGAGAATATTATTGGTCAGGGCATCAA	960
	C C G C T A C G	
961	ATAATGGCTTCTCCTGTTAGGGTTTCGGGCCAGAATTCA	1000
	C C A T A C A G C C G T	
1001	CTTTCCGCTATATGAACTATGGAAATGCAGCTCCACA	1040
	C T C C C	
1041	ACAACGTATTGTTGCTCAACTAGGTCAAGGGCGTGTATAGA	1080
	C T C C	
1081	ACATTATCGTCCACCTTATATAGAAGACCTTTAAATATAG	1120
	C G T G C C C C	
1121	GGATAAAATAATCAACAACCTATCTGTTCTTGACGGGACAGA	1160
	T C C C G T C A	
1161	ATTTGCTTATGGAACCTCTCAATTGCCATCCGCTGTA	1200
	G C C T T C T T	
1201	TACAGAAAAAGCGGAACGGTAGATTCCCTGGATGAAATAC	1240
	G C T C T C C	
1241	CGCCACAGAATAACAAACGTGCCACCTAGGCAGGGATTAG	1280
	A C T C C T C	
1281	TCATCGATTAAGCCATGTTCAATGTTGCTCAGGCTT	1320
	C C A G G C G C C C A C	
1321	AGTAATAGTAGTGTAACTATAATAAGAGCTCTATGTTCT	1360
	C C T C C G C C C	
1361	CTTGGGATACATCGTAGTGCTGAATTAAATAATATAATTCC	1400
	A T G C C C	

FIGURE 3B

1401	TTCATCACAAATTACACAAATACCTTTAACAAAATCTACT	1440
	C T C C C A G C G	
1441	AATCTGGCTCTGGAACCTCTGCGTTAAGGACCGAGGAT	1480
	C A G G C	
1481	TTACAGGAGGAGATATTCTCGAAGAACCTCACCTGGCCA	1520
	C T A T	
1521	GATTTCAACCTTAAGAGTAAATATTACTGCGACCTTATCA	1560
	AGC C C T C C C T T	
1561	CAAAGATATCGGGTAAGAATTCTCGTACGCTTACACAA	1600
	T C G T A A A	
1601	ATTTACAATTCCATACATCAATTGACGGAAGACCTATTAA	1640
	C G C C C G C	
1641	TCAGGGAAATTTTCAGCAACTATGAGTAGTGGGAGTAAT	1680
	T C C C C T C A C C C C	
1681	TTACAGTCCGGAAAGCTTCTGGACTGTAGGTTTACTACTC	1720
	G A C C A C C C	
1721	CGTTTAACTTTCAATGGATCAAGTGTATTTACGTAAG	1760
	T C C T C C T C C T	
1761	TGCTCATGTCTTCATTCAAGGCAATGAACCTTATATAGAT	1800
	C G T G C T C	
1801	CGAATTGAATTGTTCCGGCAGAAGTAACCTTGAGGCAG	1840
	T G G T C T C T	
1841	AATAT 1845	
	G C	

FIGURE 3C

1	ATGGATAACAATCCGAAACATCAATGAATGCATTCCTTATA C C A C A C	40
41	ATTGTTAACGTAACCCCTGAAGTAGAAGTATTAGGGAGA C C G A T C T	80
81	AAGAATAGAAACTGGTTACACCCCCATCGATATTCCTTG C C T C T C C C	120
121	TCGCTAACCCAATTCTTTGAGTGAAATTGTTCCCGTG C T G A G G C C G C G A	160
161	CTGGATTTGTGTTAGGACTAGTTGATATAATATGGGAAT G C T C C C C C T	200
201	TTTGGTCCCTCTAACGGACGCATTCTTGATCAAATT C A T C G G G	240
241	GAACAGTTAACCAACAAAGAATAGAAGAATTGCGTAGGA G G C G G C G C	280
281	ACCAAGCCATTCTAGATTAGAAGGACTAACGAACTTTA G C G G T G C	320
321	TCAAATTACGCGAGAATCTTTAGAGAGTGGGAAGCAGAT C C T G A G C C	360
361	CCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCAAT C T C C C G A	400
401	TCAATGACATGAAACAGTGCCTTACACCGCTATTCTCT C C T G C A C A T	440
441	TTTGCGAGTTCAAAATTATCAAGTCCCTTTTATCAGTA G C C G C C C G C G G	480
481	TATGTTCAAGCTGCAATTACATTATCAGTTTGAGAG C A T C T C C C A G C G C	520
521	ATGTTCAAGTGTGGACAAAGGTGGGATTGATGCCGC C A G C G C T	560
561	GACTATCAATAGTCGTTATAATGATTAACTAGGCTTATT A C C C C C C T G	600
601	GGCAACTATACAGATTATGCTGTACGCTGGTACAATACGG A C C C C T T C T	640
641	GATTAGAACGTGTATGGGGACCGGATTCTAGAGATTGGGT C G G C T T A	680

FIGURE 4A

681	AAGGTATAATCAATTAGAAGAGAATTAACACTAAGTGA T A C C G C G G C C A T	720
721	TTAGATATCGTTGCTCTGTTCCCGAATTATGATAGTAGAA G C T GT C C C CTCC	760
761	GATATCCAATTCGAACAGTTCCCAATTAAACAAGAGAAAT CC C T C T G C T C	800
801	TTATACAAACCCAGTATAGAAAATTGGATGGTAGTTT C T T C T G C C C C C	840
841	CGAGGCTCGGCTCAGGGCATAGAAAGAGTATTAGGAGTC T T C A T C G C T C C	880
881	CACATTTGATGGATATACTAACAGTATAACCATCTATAC C C C CTG C T C	920
921	GGATGCTCATAGGGTTATTATTATTGGTCAGGGCATCAA C C A AG G C T A C G	960
961	ATAATGGCTTCTCTGTAGGGTTTCCGGGCCAGAATTCA C C A T A C A G C C G T	1000
1001	CTTTCCGCTATATGAACTATGGAAATGCAGCTCCACA C T C C	1040
1041	ACAACGTATTGGTGCCTCAACTAGGTCAAGGGCGTGTATAGA C T C C	1080
1081	ACATTATCGTCCACTTTATATAGAAGACCTTTAATATAG C G T C G C C C C	1120
1121	GGATAAAATAATCAACAACATCTGTTCTGACGGGACAGA T C C C G T C A	1160
1161	ATTTGCTTATGGAACCTCTCAATTGGCATCCGCTGTA G C C T T C T	1200
1201	TACAGAAAAGCGGAACGGTAGATTCCGCTGGATGAAATAC G C T C T C C	1240
1241	CGCCACAGAATAACAAACGTGCCACCTAGGCAAGGATTAG A C T C C T C	1280
1281	TCATCGATTAAAGCCATGTTCAATGTTCTGTTCAAGGCTT C C A G G C G C C A C	1320
1321	AGTAATAGTAGTGTAAAGTATAATAAGAGCTCTATGTTCT C C T C G C C C	1360
1361	CTTGGATACATCGTAGTGCTGAATTAAATAATTGCG C G C C C C C	1400

FIGURE 4B

1401	ATCGGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC	1440
	C	
1441	TTTCTTTTAATGGTTCTGTAATTCAGGACCAGGATTAA	1480
	C C C C	C
1481	CTGGTGGGGACTTAGTTAGATTAATAGTAGTGGAAATAA	1520
	A C C C	C C C C
1521	CATTCAAGAATAGAGGGTATATTGAAGTTCAATTCACTTC	1560
1561	CCATCGACATCTACCAAGATATCGAGTTCGTGTACGGTATG	1600
	C A	GA
1601	CTTCTGTAACCCCGATTCAACCTCAACGTTAATTGGGTAA	1640
	G T	
1641	TTCATCCATTTTCCAAATACAGTACCAAGCTACAGCTACG	1680
	C C T	C
1681	TCATTAGATAATCTACAAATCAAGTGTATTTGGTTATTTG	1720
	C G C C C C C C	C C
1721	AAAGTGCCAATGCTTTACATCTTCAATTAGGTAATATAGT	1760
	C C C C	
1761	AGGTGTTAGAAATTAGTGGACTCCAGGAGTGATAATA	1800
	G C T C	T C
1801	GACAGATTGAAATTATTCAGTTACTGCAACACTCGAGG	1840
	C G C	
1841	CTGAATATACTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
	A TGCG	
1881	GCTGTTACGTCTACAAACCAACTAGGGCTAAAAACAAAT	1920
	CTGT ACGTCTACA C AGCT G ACTC G CA TG	
1921	G 1921	

FIGURE 4C

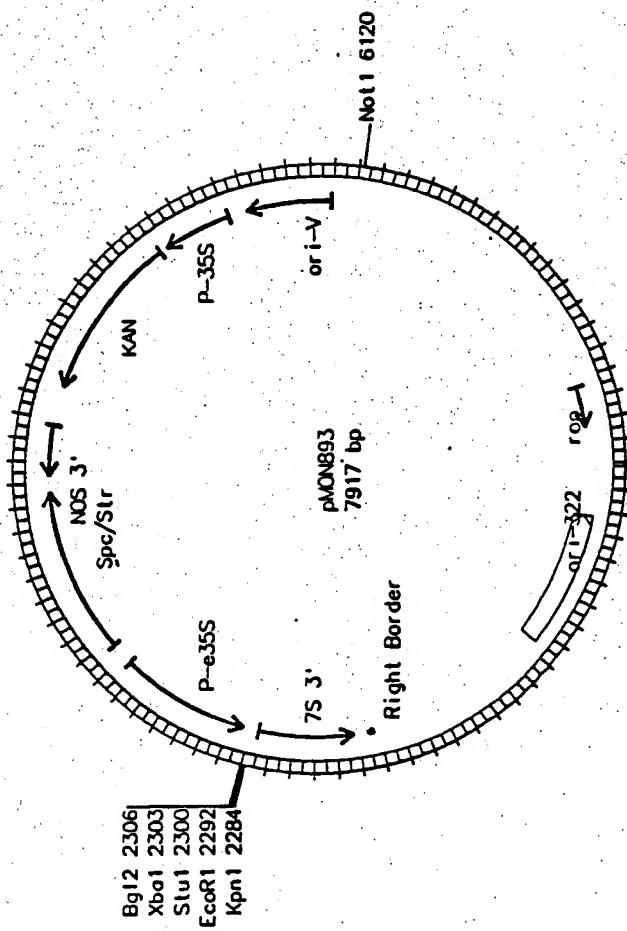


FIGURE 5

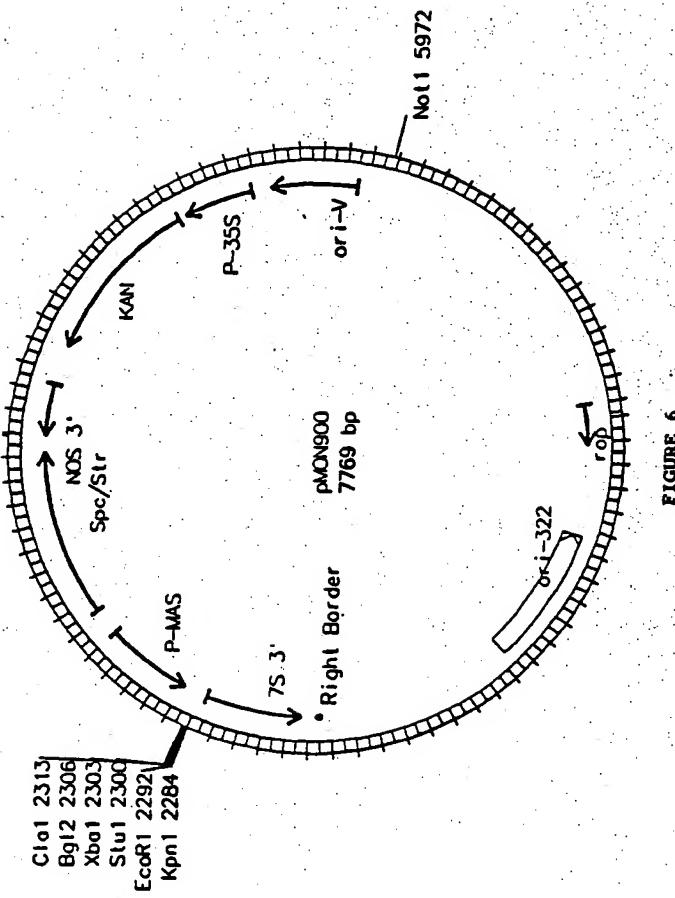


FIGURE 6

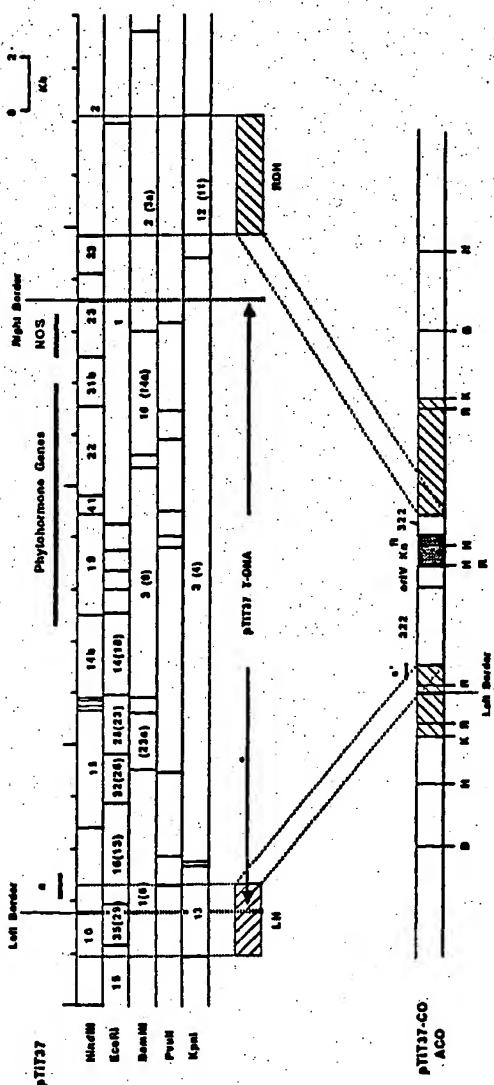


FIGURE 7

1	GAAAGAATAGAAACTGGTTACACCCCAATCGATATTCCT	40
	ATGGCC T C T C C C	
41	TGTCGCTAACGCAATTCTTTGAGTGAATTGTTCCGG	80
	CT G A G GC C C G C G A	
81	TGCTGGATTGTGTTAGGACTAGTTGATATAATATGGGA	120
	G C TC C C C C T	
121	ATTTTGGTCCCTCTCAATGGGACGCATTTCTTGTACAAA	160
	C A T C G G	
161	TTGAACAGTTAATTAACCAAGAATAGAAGAATTGCTAG	200
	G G C G G C G C	
201	GAACCAAGCCATTCTAGATTAGAAGGACTAAGCAATCTT	240
	G C G G C T G C	
241	TATCAAATTACGCAGAATTTAGAGAGTGGGAAGCAG	280
	C T GAGC C G	
281	ATCCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCA	320
	C TC CC C G A	
321	ATTCAATGACATGAAAGCTGCCCTTACAACCGCTATTCTT	360
	C C T G C A C A	
361	CTTTTGCAAGTCAAAATTATCAAGTCTCTTTATCAG	400
	T G C C G C C C G C	
401	TATATGTTCAAGCTGCAATTACATTATCAGTTTGAG	440
	G C A T C T CC CAGC GC TC	
441	AGATGTTCAAGTGTGGACAAAGGTGGGATTGATGCC	480
	C AGC G C T	
481	GCGACTATCAATAGTCGTTATAATGATTTAAGCTAGGCTTA	520
	A C C C C C T G	
521	TTGGCAACTATACAGATTATGCTGTACGCTGGTACAATAC	560
	A C C C C T T C	
561	GGGATTAGAACGTGTGGGACCGGATTCTAGAGATTGG	600
	T C G G C T T	
601	GTAAGGTATAATCAATTAGAAGAGAATTAACACTAAG	640
	A T A C C G C G G C C A	
641	TATTAGATATCGTGTGCTGTGTCGGAAATTATGATAGTAG	680
	T G C T G T C C	

FIGURE 8A

681	AAGATATCCAATTCGAACAGTTCCCAATTAAACAAGAGAA	720
	CC C T C T G C T C	
721	ATTTATACAAACCCAGTATTAGAAAATTGATGGTACTT	760
	C T T C T G C C C	
761	TTCGAGGCTCGGCTCAGGGCATAGAAAGAAGTATTAGGAG	800
	C T T T C A T C G C T C C C	
801	TCCACATTTGATGGATATACTTAACAGTATAACCATCTAT	840
	C C C C T G C T C	
841	ACGGATGCTCATAGGGTTATTATTATGGTCAGGGCATC	880
	C C A A G G C T A C	
881	AAATAATGGCTTCTCTGTAGGGTTTCGGGGCCAGAATT	920
	G C C A T A C A G C C G	
921	CACTTTCCGCTATATGGAACATGGAAATGCGAGCTCCA	960
	T C T C C C	
961	CAACAACGTATTGTTGTCAACTAGGTCAAGGGCGTGTATA	1000
	C T C C	
1001	GAACATTATCGTCCACTTTATATAGAAGACCTTTAATAT	1040
	C G T C G C C C	
1041	AGGGATAAAATAATCAACAACTATCTGTTCTGACGGGACA	1080
	C T C C C G T C A	
1081	GAATTTGCTTATGAAACCTCTCAAAATTGCCATCCGCTG	1120
	G C C T T C	
1121	TATACAGAAAAAGCGGAACGGTAGATTCGCTGGATGAAAT	1160
	T G C T C T C	
1161	ACCGCCACAGAATAACAAACGTGCCACCTAGGCAAGGATT	1200
	C A C T C C	
1201	AGTCATCGATTAAGCCATTTCAATGTTCTGTTCAAGGCT	1240
	TCC C A G G C G C C C A	
1241	TTAGTAATAGTAGTGTAAAGTATAATAAGAGCTCCATGTT	1280
	C C C T C G C C C	
1281	CTCTGGATAACATCGTAGTGCTGAATTAAATAATATAATT	1320
	C G C C C C C	
1321	GCATCGGATAGTATTACTCAAATCCCTGCAGTGAAGGGAA	1360
	C	
1361	ACTTTCTTTAAATGGTTCTGTAATTTCAGGACCAGGATT	1400
	C C C C	

FIGURE 8B

1401 TACTGGTGGGGACTTAGTTAGATTAATAGTAGTGGAAAT 1440  
 C A C C C C C C  
 1441 AACATTCAAGAATAGAGGGTATATTGAAGTTCCAATTCACT 1480  
 1481 TCCCACATCTACCAAGATATCGAGTTCGTGTACGGTA 1520  
 C A G A  
 1521 TGCTTCTGTAACCCGATTCAACCTCAACGTTAATTGGGGT 1560  
 G T  
 1561 AATTCACTCCATTTCACAGTACAGCTACAGCTA 1600  
 C C T  
 1601 CGTCATTAGATAATCTACAATCAAGTGATTTGGTTATTT 1640  
 C C G C C C C C  
 1641 TGAAAGTCCAATGCTTTACATCTTCATTAGGTAAATA 1680  
 C C C C  
 1681 GTAGGTGTTAGAAATTAGTGGGACTGCAGGAGTGATAA 1720  
 G C T  
 1721 TAGACAGATTGAATTATTCCAGTTACTGCAACACTCGA 1760  
 C C G C  
 1761 GGCTGAA 1767  
 G

FIGURE 8C

1	ATGGATAACAATCCGAACATCAATGAATGCATTCCCTATA	40
	C C A C A C	
41	ATTGTTAACGTAACCCCTGAAGTAGAAGTATTAGTGGAGA	80
	C C G A T C T	
81	AAGAATAGAAACTGGTACACCCCCAATCGATATTCCTTG	120
	C C T C T C C C	
121	TCGCTAACGCAATTCTTTGAGTGAAATTGTTCCGGTG	160
	C T G A G G C C C G G C G A	
161	CTGGATTGTGTTAGGACTAGTTGATATAATATGGGAAT	200
	G C T C G C C C T	
201	TTTGTTCCCTCTCAATGGGACGCATTCTTGATCAAATT	240
	C A T C G G G	
241	GAACAGTTAACCAAAGAATAGAAGAATTCTCGTAGGA	280
	G G C G G C G	
281	ACCAAGCCATTCTAGATTAGAAGGACTAAGCAATCTTA	320
	G C G G T G C	
321	TCAAATTACGCGAGAATCTTTAGAGAGTGGGAAGCAGAT	360
	C T GAGC C C	
361	CCTACTAATCCAGCATTAAGAGAAGAGATGCGTATTCAAT	400
	C T C C C G A	
401	TCAATGACATGAAACAGTGGCCCTTACAACCGCTATTCTCT	440
	C T G C A C AT	
441	TTTGCGAGTTCAAAATTATCAAGTTCCCTTTTATCAGTA	480
	G C C G C C C G G	
481	TATGTTCAAGCTGCAAATTACATTATCAGTTTGAGAG	520
	C A T C T C C C A G C G C T C	
521	ATGTTTCAGTGTGTTGGACAAAGGTGGGGATTGATGCCGC	560
	C A G C G C T	
561	GACTATCAATAGTCGTATAATGATTTAACCTAGGCTTATT	600
	A C C C C C T G	
601	GGCAACTATAACAGATTATGCTGTACCGCTGGTACAATACGG	640
	A C C C C C T T C T	
641	GATTAGAACGTGTATGGGGACCGGATTCTAGAGATTGGGT	680
	C G G C T T A	

FIGURE 9A

681	AAGGTATAATCAATTAGAAGAGAATTAACACTAACTGTA T A C C G C G G C C A T	720
721	TTAGATATCGTGTGCTGTTCCCGAATTATGATAGTAGAA G C T G T C C C T C C C T C C	760
761	GATATCCAATTGAAACAGTTTCCCAATTAAACAAGAGAAAT CC C T C T G C T C C	800
801	TTATACAAACCCAGTATTAGAAAATTGATGGTAGTTT C T T C T G C C C C C	840
841	CGAGGCCTGGCTCAGGGCATAGAAAGAAGTATTAGGAGTC T T C A T C G C T C C	880
881	CACATTGATGGATATACTTAAACAGTATAACCATCTATAAC C C C T G C T C	920
921	GGATGCTCATAGGGGTATTATTATTGGTCAGGGCATCAA C C A A G G C T A C G	960
961	ATAATGGCTCTCCTGTAGGGTTTCGGGCCAGAAATTCAC C C A T A C A G C C G T	1000
1001	CTTTCCGCTATATGAACTATGGAAATGCAAGCTCCACA C T C C C	1040
1041	ACAACGTATTGTTGCTCAACTAGGTCAAGGGCTGTATAGA C T C C	1080
1081	ACATTATCGTCCACTTTATATAGAAGACCTTTAATATAG C G T C G C C C C	1120
1121	GGATAAATAATCAACAACATCTGTTCTGACGGGACAGA T C C C G T C A	1160
1161	ATTTGCCATTGGAACCTCCCTCAAATTGCCATCCGCTGTA G C C T T C T	1200
1201	TACAGAAAAGCGGAACGGTAGATTGCTGGATGAAATAC G C T C T C C	1240
1241	CGCCACAGAATAACACCGTCCACCTAGGCAAGGATTAG A C T C C T C C	1280
1281	TCATCGATTAAGCCATGTTCAATGTTGCTGTCAGGCTT C C A G G C G C C C A C	1320
1321	AGTAATAGTAGTGTAAAGTATAATAAGAGCTCCTATGTTCT C C T C G C C C	1360
1361	CTTGGATACATCGTAGTGCTGAATTAAATAATATAATTGC C G C C C C C	1400

FIGURE 9B

1401	ATCGGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC C	1440
1441	TTCTTTTAATGGTTCTGAAATTCAAGGACCAGGATT C C C C C	1480
1481	CTGGTGGGACTTAGTTAGATTAAATAGTAGTGGAAATAA A CC C C C C	1520
1521	CATTCAGAAATAGAGGGTATATTGAAGGTTCAATTCACTTC	1560
1561	CCATCGACATCTACCAAGATATCGAGTCGTGTACGGTATG C A GA	1600
1601	CTTCTGTAACCCCGATTCACTCAACGTTAATTGGGGTAA G T	1640
1641	TTCATCCATTTTTCCAATACAGTACCACTACAGCTACG C C T C	1680
1681	TCATTAGATAATCTACAACTCAAGTGAATTGGTTATTTG C G C C C C C	1720
1721	AAAGTGCCAAATGCTTTACATCTTCATTAGGTAAATAGT C C C C	1760
1761	AGGTGTTAGAAATTAGTGGGACTGCAGGAGTGATAATA G C T C	1800
1801	GACAGATTGAAATTATTCAGTTACTGCAACACTCGAGG C G C	1840
1841	CTGAATATAATCTGGAAAGAGCGCAGAAGGCGGTGAATGC	1880
1881	GCTGTTACGTCTACAAACCAACTAGGGCTAAAACAAAT	1920
1921	GTAACGGATTATCATATTGATCAAGTGTCCAATTAGTTA	1960
1961	CGTATTATCGGATGAATTGTCTGGATGAAAAGCGAGA	2000
2001	ATTGTCCGAGAAAGTCAAACATGCGAAGCGACTCAGTGAT	2040
2041	GAACGCAATTACTCCAAGATTCAATTCAAAGACATTA	2080
2081	ATAGGCAACCAGAACGTGGGTGGGGCGGAAGTACAGGGAT	2120

FIGURE 9C

2121	TACCATCCAAGGGAGGGATGACGTATTTAAGAAAATTAC	2160
2161	GTCACACTATCAGGTACCTTGATGAGTGCTATCCAACAT	2200
2201	ATTTGTATCAAAAATCGATGAATCAAAATTAAAAGCCTT	2240
2241	TACCCGTTATCAATTAAGAGGGTATATCGAAGATAGTCAA	2280
2281	GA CTTAGAAATCTATTTAATTG C TACAAATG C AAAACATG	2320
2321	AAACAGTAAATGTGCCAGGTACGGGTTCTTATGGCCGCT	2360
2361	TTCAGCCCAAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400
2401	CGATGCGGCCACACCTGAAATGGAATCCTGACTTAGATT	2440
2441	GTTCGTGTAGGGATGGAGAAAAGTGTGCCCATTCGCA	2480
2481	TCATTTCTCTTAGACATTGATGTAGGATGTACAGACTTA	2520
2521	AATGAGGACCTAGGTGTATGGGTGATCTTAAGATTAAGA	2560
2561	CGCAAGATGGCACGCAAGACTAGGGAACTAGAGTTCT	2600
2601	CGAAGAGAAACCATTAGTAGGAGAAGCGCTAGCTCGTGTG	2640
2641	AAAAGAGCGGAGAAAAATGGAGAGACAAACGTAAAAAT	2680
2681	TGGAATGGGAAACAAATATCGTTATAAGAGGCAAAAGA	2720
2721	ATCTGTAGATGCTTATTTGTAACACTCTCAATATGATCAA	2760
2761	TTACAAGCGGATACGAATATTGCCATGATTCTGCGGCAG	2800
2801	ATAAACGTGTTCATAGCATTGAGAAGCTTATCTGCCCTGA	2840

FIGURE 9D

2841	GCTGTCTGTGATTCCGGGTGTCATGCGCTATTTGAA	2880
2881	GAATTAGAAGGGCGTATTTCACTGCATTCTCCCTATATG	2920
2921	ATGCGAGAAATGTCATTAATAATGGTGAATTTAATATGG	2960
2961	CTTATCCTGCTGGAACCTGAAAGGGCATGTAGATGTAGAA	3000
3001	GAACAAAACAACCAACGTTCGGTCTTGTGTTCCGGAAAT	3040
3041	GGGAAGCAGAAGTGTACAAGAAGTTCGTGTCTGTCCGGG	3080
3081	TCGTGGCTATATCCTCGTGTACAGCGTACAAGGAGGGAA	3120
3121	TATGGAGAAGGTTGCGTAACCATTCATGAGATCGAGAACAA	3160
3161	ATACAGACGAACGTAAAGTTAGCAACTGCGTAGAAGAGGA	3200
3201	AATCTATCCAATAACACGGTAACGTGTAATGATTATACT	3240
3241	GTAAATCAAGAAGAACACGGAGGTGCGTACACTTCTCGTA	3280
3281	ATCGAGGATATAACGAAGCTCCTCCGTACCGCTGATTA	3320
3321	TGCGTCAGTCTATGAAGAAAATCGTATACAGATGGACGA	3360
3361	AGAGAGAAATCCTTGTGAATTAAACAGAGGGTATAGGGATT	3400
3401	ACACGCCACTACCAAGTTGGTTATGTGACAAAAGAATTAGA	3440
3441	ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
3481	GAAACGGAAGGAACATTATCGTGGACAGCGTGGAAATTAC	3520
3521	TCCTTATGGAGGAA 3534	

FIGURE 9E

1	ATGGATAACAATCCGAACATCAATGAATGCATTCCCTATA	40
	C C A C A C A C	
41	ATTGTTAACCTGTTAACCTGAAAGTAGAAGTATTAGTGGAGA	80
	C C G A T C T	
81	AGAAATAGAAACTGGTTACACCCCCATCGATATTCCTTG	120
	C C T C T C C C	
121	TCGCTAACGCAATTCTTTGAGTGAATTGTTCCCGTG	160
	CT G A G G C C C G C G A	
161	CTGGATTGTGTTAGGACTAGTTGATATAATATGGGAAT	200
	G C T C C C C T	
201	TTTGGTCCCTCTCAATGGGACCCATTCTGTACAAATT	240
	C A T C G G G	
241	GAACAGTTAATTAACCAAAGAATAGAAGAATTGCTAGGA	280
	G G C G G C G C	
281	ACCAAGCCATTCTAGATTAGAAGGACTAACGCAATCTTA	320
	G C G G T G C	
321	TCAAATTACCGAGAACATCTTTAGAGAGTGGAAGCAGAT	360
	C C T GAGC C C	
361	CCTACTAATCCAGCATTAAAGAGAAGAGATGCGTATTCAAT	400
	C T C C C G A	
401	TCAATGACATGAACAGTGCCTTACACCCGCTATTCCCT	440
	C C T G C A C A T	
441	TTTGCAGTCAAAATTATCAAGTCTCTTTATCAGTA	480
	G C C G C C C G G C G G	
481	TATGTTCAAGCTGCAAAATTACATTATCAGTTGAGAG	520
	C A T C T C C C A G C G T C	
521	ATGTTTCAGTGGGACAAAGGTGGGATTTGATGCCGC	560
	C A G C G C T	
561	GACTATCAATAGTCGTTATAATGATTAACTAGGCTTATT	600
	A C C C C C T G	
601	GGCAACTATACAGATTATGCTGTACGCTGGTACAATACGG	640
	A C C C C T T C T	

FIGURE 10A

641	GATTAGAACGTGTATGGGGACCGGATTCTAGAGATTGGT	680
	C G G C T T A	
681	AAGGTATAATCAATTAGAAGAGAATTAAACACTRACTGTA	720
	T A C C G C G G C C A T	
721	TTAGATATCGTTGCTCTGTTCCCGAATTATGATAGTAGAA	760
	G C T G T C C T C C	
761	GATATCCAATTGAAACAGTTCCCAATTAAACAAGAGAAAT	800
	CC C T C T G C T C	
801	TTATACAAACCCAGTATTAGAAAAATTGATGGTAGTTT	840
	C T T C T G C C C C	
841	CGAGGCCGCTCAGGGCATAGAAAGAAGTATTAGGAGTC	880
	T T C A T C G C T C C C	
881	CACATTGATGGATATACTTAACAGTATAACCATCTATAC	920
	C C C T G C T C	
921	GGATGCTCATAGGGGTATTATTATTGGTCAGGGCATCAA	960
	C C A A G G C T A C G	
961	ATAATGGCTTCTCCCTGAGGGTTTCGGGCCAGAAATTCA	1000
	C C A T A C A G C G G T	
1001	CTTTTCCGCTATATGAACTATGGGAAATGCAGCTCCACA	1040
	C T C C C	
1041	ACAACGTATGTTGCTCACTAGGTCAAGGGCGTGTATAGA	1080
	C T C C	
1081	ACATTATCGTCCACTTTATATAGAAGAGACCTTTAATATAG	1120
	C G T C G C C C C	
1121	GGATAAAATAATCAACAACTATCTGTTCTGACGGGACAGA	1160
	T C C C G T C A	
1161	ATTTGCTTATGGAACCTCCCTCAAATTGCCATCCGCTGTA	1200
	G C C T T C T C T	
1201	TACAGAAAAACGGGAACGGTAGATTCGCTGGATGAAATAC	1240
	G C T C T C C	
1241	CGCCACAGAATAACACAGTCCACCTAGGCAAGGATTAG	1280
	A C T C C T C	
1281	TCATCGATTAAAGCCATGTTCAATGTTGTTAGGCTT	1320
	C C A G G C C C A C	
1321	AGTAATAGTAGTGTAAAGTATAAAAGAGCTCCTATGTTCT	1360
	C C T C C G C C C	

FIGURE 10B

1361	CTTGGATACATCGTAGTGCTGAATTAAATAATATAATTGC	1400
	C G C C C C	
1401	ATCGGATAGTATTACTCAAATCCCTGCAGTGAAAGGGAAAC	1440
	C	
1441	TTTCTTTAAATGGTTCTGTAATTCAAGGACCAGGTTA	1480
	C C C C C C	
1481	CTGGTGGGACTTAGTTAGATTAAATAGTAGTAGTGGAAATAA	1520
	A C C C C C	
1521	CATTCAAGAATAGAGGGTATATTGAAGTTCCAATTCACTTC	1560
1561	CCATCGACATCTACCAAGATATCGAGTCGTCGTACCGTATG	1600
	C A G A	
1601	CTTCTGTAACCCCGATTCACCTCAACGTTAATTGGGGTAA	1640
	G T	
1641	TTCATCCATTTTCCAAATACAGTACCCAGCTACAGCTACG	1680
	C C T C	
1681	TCATTAGATAATCTACAATCAAGTGAATTGGTTATTTG	1720
	C G C C C C	
1721	AAAGTGCCAATGCTTTACATCTTCATTAGGTAATATAGT	1760
	C C C C	
1761	AGGTGTTAGAAATTTAGTGGACTGCAGGAGTGATAATA	1800
	G C T C	
1801	GACAGATTTGAATTATTCCAGTTACTGCAACACTCGAGG	1840
	C G C	
1841	CTGAATATAATCTGGAAAAGAGCGCAGAAGGCGGTGAATGC	1880
1881	GCTGTTACGTCTACAAACCAACTAGGGCTAAAAACAAAT	1920
	G C C C G C	
1921	GTAACGGATTATCATATTGATCAAGTGTCCAATTAGTTA	1960
	G C G G	
1961	CGTATTATCGGATGAATTGTCTGGATGAAAAGCGAGA	2000
	C CC CAGC G C	
2001	ATTGTCCGAGAAAAGTCAAACATGCGAAGCGACTCAGTGAT	2040
2041	GAACGCAATTACTCCAGATTCAAATTCAAAGACATTA	2080

FIGURE 10C

2081	ATAGGCAACCAGAACGTGGGTGGGCGGAAGTACAGGGAT	2120
2121	TACCATCAGGAGGGGATGACGTATTTAAGAAAAATTAC G T C G C G G C	2160
2161	GTCACACTATCAGGTACCTTGATGAGTGCTATCCAACAT	2200
2201	ATTTGTATCAAAAAATCGATGAATCAAAATAAAAGCCTT CC C C G G C G C G G C	2240
2241	TACCCGTTATCAATTAAGAGGGTATATCGAAGATAGTCAA	2280
2281	GACTTAGAAATCTATTAAATCGCTACAATGCAAAACATG C C G C C C	2320
2321	AAACAGTAAATGTGCCAGGTACGGTTCCTTATGGCGCT	2360
2361	TTCAGCCCAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT	2400
2401	CGATGGCGCCACACCTTGAATGGAATCCTGACTTAGATT	2440
2441	GTTCGTGTAGGGATGGAGAAAAGTGTGCCCATCGCA	2480
2481	TCATTTCTCCTTAGACATTGATGTAGGATGTACAGACTTA	2520
2521	AATGAGGACCTAGGTATGGGTATCTTAAGATTAAGA	2560
2561	CGCAAGATGGCACCGCAAGACTAGGGATCTAGAGTTCT	2600
2601	CGAAGAGAAACCATAGTAGGAGAACCGCTAGCTCGTGTG	2640
2641	AAAAGAGCGGAGAAAAATGGAGAGACAAACGTAAAAAT G G	2680
2681	TGGAATGGAAACAATATCGTTATAAAGAGGGCAAAAGA G C C C C	2720
2721	ATCTGTAGATGCTTTATTTGTAACACTCTCAATATGATCAA	2760
2761	TTACAAGCGGATACGAATATTGCCATGATTGATGCCAG	2800

FIGURE 10D

2801	ATAAACGTGTTCATAGCATTGGAGAAGCTTATCTGGCTGA	2840
2841	GCTGTCGTGATTCCGGGTGTCATGGCTATTTTGAA	2880
2881	GAATTAGAAGGGCGTATTTCACTGCATTCTCCCTATATG C C	2920
2921	ATGCGAGAAATGTCATTAATGGTATTTAATAATGG C C C G C C C C	2960
2961	CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA	3000
3001	GAACAAAACAAACCAACGTTCGGTCTGTGTTCCGGAAT	3040
3041	GGGAAGCAGAAGTGTACAGAAGAGTCGTGTCGTGTCGGG	3080
3081	TCGTGGCTATATCCTCGTGTACACCGTACAAGGAGGGA	3120
3121	TATGGAGAAGGTTGCGTAACCATTCAATGAGATCGAGAACAA	3160
3161	ATACAGACGAACGTGAAAGTTAGCAACTGCGTAGAAGAGGA	3200
3201	AATCTATCCAAATAACACGGTAACGTGTAATGATTACT	3240
3241	GTAAATCAAGAAGAATACGGAGGTGCGTACACTTCGTA	3280
3281	ATCGAGGATATAACGAAGCTCCTCCGTACCGCTGATTA	3320
3321	TGGCTCAGTCTATGAAGAAAATCGTATAACAGATGGACGA	3360
3361	AGAGAGAATCCTGTGAAATTAAACAGAGGTATAGGGATT	3400
3401	ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA	3440
3441	ATACTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
3481	GAAACGGAAGGAACATTATCGTGGACAGCGTGGATTAC	3520
3521	TCCTTATGGAGGAA 3534	

FIGURE 10E

1	ATGGATAACATCCGAAACATCAATGAATGCATTCCCTATA	40
	C C A C C A C	
41	ATTGTTAACGAAACCTGAAAGTAGAAGTATTAGGGAGA	80
	C C G A T C T	
81	AAGAATAGAAAATGGTTACACCCCCAATCGATATTCCTTG	120
	C C T C T C C C	
121	TCGCTAACCGAATTCTTTGAGTGAATTGTTCCCGTG	160
	CT G A G G C C C G C G A	
161	CTGGATTTGTTAGGACTAGTTGATATAATATGGGAAT	200
	G C T C C C C T	
201	TTTGGTCCCTCTCAATGGGACGCATTCTGTACAAATT	240
	C A T C G G G	
241	GAACAGTTAACCAACAAAGAATAGAAGAATTGCGTAGGA	280
	G G C G G C G C	
281	ACCAAGCCATTCTAGATTAGAAGGACTAAGCAATTCTTA	320
	G C G G T G C	
321	TCAAATTTCAGCAGAACATTTAGAGAGTGGGAGCAGAT	360
	C T G A G G C C	
361	CCTACTAACCCAGCTTAAGAGAAGAGATGCGTATTCAAT	400
	C T C C C G A	
401	TCAATGACATGAACAGTGCCCTTACAACCGCTATTCCCT	440
	C C T G C A C A T	
441	TTTGCAAGTCAAAATTATCAAGTTCTTTATCAGTA	480
	G C C G C C C G G C G G	
481	TATGTTCAAGTCGCAATTTCATTTACAGTTTGAGAG	520
	C A T C T C C C A G C G C T C	
521	ATGTTTCAGTGTGGACAAAGGTGGGATTTGATGCCGC	560
	C A G C G C T	
561	GACTATCAATAGTCGTTATAATGATTTAACCTAGGCTTATT	600
	A C C C C C T G	
601	GGCAACTATACAGATTATGCTGTACGCTGGTACAATACGG	640
	A C C C C T T C T	
641	GATTAGAACGTGTATGGGACCGGATTCTAGAGATTGGGT	680
	C G G C T T A	

FIGURE 11A

681	AAGGTATAATCAATTAGAAGAGAATTAACACTAAGTGTAT	720
721	T A C C G G G C C C A T	
721	TTAGATATCGTTGCTCTGTTCCCGAATTATGATAGTAGAAAGCCTGTCTCC	760
761	GATATCCAATTGCAACAGTTTCCCAATTAAACAAGAGAAATCCCTCTGCTC	800
801	TTATACAAACCCAGTATTAGAAAATTGATGGTAGTTTCTCTGCTGCCCC	840
841	CGAGGCTCGGCTCAGGGCATAGAAAGAAGTATTAGGAGTCCTTCATCCTCC	880
881	CACATTGATGGATACATTAACAGTATAACCATCTATACCTGCTGCTC	920
921	GGATGCTCATAGGGTTATTATTATTGGTCAGGGCATCAACAGGAGGAGG	960
961	ATAATGGCTCTCCTGTAGGGTTTCGGGCCAGAATTCACTAAGGAGGAGG	1000
1001	CTTTCCGCTATATGAACTATGGAAATGCAAGCTCCACACAGGAGGAGG	1040
1041	ACAACTGTTGCTCAACTAGGTCAAGGGCGTGTATAGACAGGAGGAGG	1080
1081	ACATTATCGTCCACTTTATATAGAAGACCTTTAATATAGCTGCTGCTG	1120
1121	GGATAAAATAATCAACAACTATCTGTTCTGACGGGACAGATCCTGCTG	1160
1161	ATTTGCTTATGGAACCTCCTCAAATTGCCATCCGCTGTAACAGGAGGAGG	1200
1201	TACAGAAAAAGCGGAACGGTAGATTGCTGGATGAAATACCTGCTGCTG	1240
1241	CGCCACAGAATAACAACGTGCCACCTAGGCAAGGATTAGCTCTGCTG	1280
1281	TCATCGATTAAGCCATGTTCAATGTTGCTTCAGGCTTCAGGCTTCAGGCTT	1320
1321	AGTAATAGTAGTGTAAAGTATAATAAGAGCTCTATGTTCTGCTGCTG	1360
1361	CTTGGATACATCGTAGTGCTGAATTAAATAATATAATTGCGCTGCTG	1400

FIGURE 11B

1401 ATCGGATAGTATTACTCAAATCCCTGCAGTGAAGGGAAAC 1440  
           C  
 1441 TTTCTTTTAATGGTCTGTAATTTCAGGACCAGGATTTC 1480  
           C C C    C                    C  
 1481 CTGGGGGGACTTAGTTAGATTAATAGTAGTGGAAATAA 1520  
           A    C    C C C C  
 1521 CATTAGAATAGAGGGTATATTGAAGTTCAAATTCACTTC 1560  
 1561 CCATCGACATCTACCAAGATATCGAGTTCGTGTACGGTATG 1600  
           C            A            GA  
 1601 CTTCTGTAACCCCCATTCAACCTCAACGTTAATTGGGGTAA 1640  
           G            T  
 1641 TTCACTCCATTTCACAACTACAGTACCCAGCTACAGCTACG 1680  
           C C            T            C  
 1681 TCATTAGATAATCTACAATCAAGTGATTTGGTTATTTG 1720  
           C G            C    C    C    C  
 1721 AAAGTGCCAATGCTTTACATCTCATTAGGTAATATAGT 1760  
           C C            C C C  
 1761 AGGTGGTAGAAATTTAGTGGGACTGCAGGAGTGTATAA 1800  
           G            C            T C  
 1801 GACAGATTTGAATTTATTCCAGTTACTGCAACACTCGAGG 1840  
           C G C  
 1841 CTGAATATAATCTGGAAAAGAGCGCAGAAGGCGGTGAATGC 1880  
           G C C T G    C            T C  
 1881 GCTGTTACGTCTACAAACCAACTAGGGCTAAAACAAAT 1920  
           C C            C C T G T    CT G    T C  
 1921 GTAACGGATTATCATATTGATCAAGTGTCCAATTAGTTA 1960  
           T T C    C    C            C G C  
 1961 CGTATTATCGGATGAATTGGTCTGGATGAAAAGCGAGA 2000  
           C C C TAGC    G C C C C G    T  
 2001 ATTGTCGGAGAAAGTCAAACATGCGAAGCGACTCAGTGAT 2040  
           C C            T    C C    T    C C  
 2041 GAACGCAATTACTCCAAGATTCAAATTCAAAGACATTA 2080  
           G A G    C C T G    C C C    C  
 2081 ATAGGCAACCAGAACGGTGGGTGGGGCGGAAGTACAGGGAT 2120  
           C G            T    T    C C

FIGURE 11C

2121	TACCATCCAAGGAGGGATGACGTATTTAAAGAAAATTAC C C T G C G G C	2160
2161	GTCACACTATCAGGTACCTTGTATGAGGTGCTATCCAACAT C C C A T C C C T C	2200
2201	ATTTGTATCAAAAAATCGATGAATCAAAATTAAAGCCTT C C G G G C C C	2240
2241	TACCCGTTATCAATTAAAGAGGTATATCGAAGATAGTCAA C A G C T C C C C	2280
2281	GACTTAGAAATCTATTAAATCGCTACAATGCAAACATG C T C C G C A G C G C	2320
2321	AAACAGTAAATGTGCCAGGTACGGGTCCCTATGGCGCT G C G C T C C A	2360
2361	TTCAGCCCCAAGTCCAATCGGAAAGTGTGGAGAGCCGAAT T T C C T G T C	2400
2401	CGATGCGGCCACACCTTGAATGGAATCCTGACTTAGATT A T G G C	2440
2441	GTTCGTGTAGGGATGGAGAAAAGTGTGCCATCATTGCA C C C C G C T	2480
2481	TCATTTCTCCTTAGACATTGATGTAGGATGTACAGACTTA C G C G T C G	2520
2521	AATGAGGACCTAGGTATGGGTATTTAAGATTAAGA C A C C C C	2560
2561	CGCAAGATGGCACGCAAGACTAGGGAACTAGAGTTCT C C A T C C T	2600
2601	CGAAGAGAAACCATTAAGTAGGAGAAGCGCTAGCTCGTGTG G C T T C	2640
2641	AAAAGAGCGGGAGAAAAATGGAGAGACAAACGTAAAAAT G A G G G G C	2680
2681	TGGAATGGAAACAATATGTTATAAAGAGGCAAAAGA C T C C G C	2720
2721	ATCTGTAGATGCTTTATTTGATCAAATCTCAATATGATCAA G C G G C G C G	2760
2761	TTACAAGCGGATACGAATATTGCCATGATTCAATGCCAG G C C C C C C C C	2800
2801	ATAAACGTGTTCATAGCATTGAGAAGCTTATCTGCCCTGA C G C T G C T	2840

FIGURE 11D

2841	GCTGTCGTGATTCCGGTGTCAATGCGGCTATTTTGAA	2880
	T C C T G C T C C C G	
2881	GAATTAGAAGGGCGTATTTCACTGCATTCTCCCTATATG	2920
	C T G A C T C T T G C	
2921	ATGGAGAAAATGTCATTAATGGTGAATTAAATAATGG	2960
	C C C G C C C C	
2961	CTTATCCTGCTGGAACGTGAAAGGGCATGTAGATGTAGAA	3000
	C CAG T T G C G G	
3001	GAACAAAACAACCAACGTTCGGTCTTGTGTCGGAAAT	3040
	G T G C G G T G	
3041	GGGAAGCAGAAGTGTACAAGAAGTTCGTGTCGCCCC	3080
	T C G A A A	
3081	TCGGGCTATATCCTCGTGTACAGCGTACAAGGAGGA	3120
	A A C T C T G C T	
3121	TATGGAGAAGGTTGCGTAACCATTGAGATCGAGAACAA	3160
	C T G G C C	
3161	ATACAGACGAACGTGAAAGTTAGCAACTGCGTAGAAGAGGA	3200
	C C G T C T C G A	
3201	AATCTATCCAAAATAACACGGTAACGTGTAATGATTAACT	3240
	C C C T T C C C C	
3241	GTAAATCAAGAAGATAACGGAGGTGCGTACACTTCGTA	3280
	G G G C AGC	
3281	ATCGAGGATATAACGAAAGCTCCTCCGTACCGAGCTGATTA	3320
	CA T C T T C	
3321	TGGTCAGTCTATGAGAAAATCGTATAACAGATGGACGA	3360
	C C G C G G C C A	
3361	AGAGAGAATCCTGTGAATTAAACAGAGGGTATAGGGATT	3400
	C T C C G C T C C	
3401	ACACGCCACTACCAGTTGGTTATGTGACAAAAGAATTAGA	3440
	A T C T C G G C T	
3441	ATACTTCCCAGAAACCGATAAGGTATGGATTGAGATTGGA	3480
	G T T G C A G C C T	
3481	GAAACGGAGAACATTATCGTGGACAGCGTGGAAATTAC	3520
	C G C C G C T	
3521	TCCTTATGGAGGAA	3534
	T G	

FIGURE 11E

1	ATGACTGCAGATAATAATACGGAAGCACTAGATAGCTCTA C C C C C C C C T	40
41	CAACAAAAGATGTCATTCAAAAAGGCATTCCGTAGTAGG C T G T C G G T C T G	80
81	TGATCTCTAGGGTAGTAGGTTCCCGTTGGTGGAGCG A C T G G T A T C C C C	120
121	CTTGGTTCTGTTTATACAAAACTTTAAATACTATTTGGC C GAGC C C C C C	160
161	CAAGTGAAGACCCGTGGAGGGCTTTATGGAACAAGTAGA C G T A A C G T	200
201	AGCATTGATGGATCAGAAAATAGCTGATTATGCAAAAAT T G T A C G G C	240
241	AAAGCTCTGCGAGGTTACAGGGCCTCAAAATAATGTCG G T G A C C G C G	280
281	AAGATTATGTCAGTGATTGAGTTCATGCCAAAAAATCC G C C TCCAGC G G C	320
321	TGTGAGTTACGAAATCCACATAGCCAGGGCGGATAAGA T C C A T C A T A T C	360
361	GAGCTGTTTCTCAAGCAGAAAGTCATTTCTGTAATTCAA T C C T C C A A C	400
401	TGCCCTCGTTGCAATTCTGATACGAGGTTCTATTCT A G C T C C T T C	440
441	AACAAACATATGCACAAAGCTGCCAACACACATTTC C T C T C C G C C	480
481	CTAAAAGACGCTCAAAATTATGGAGAAGAAATGGGATACG T G C C G	520
521	AAAAAGAAGATATTGCTGAATTTATAAAAGACAACTAAA G G C G C G C T T	560
561	ACTTACGCAAGAATATACTGACCATTGTCATGGTAT G C C G C G	600
601	AATGTTGGATTAGATAAAATTAAAGAGGTTCATCTTATGAAT C T C C G C C T C C G	640
641	CTTGGGTAACCTTAAACCGTTATCGCAGAGAGATGACATT G C A A C A G C	680

FIGURE 12A

681	AACAGTATTAGATTAATTGCACTATTCCATTGTATGAT	720
	G T G C C C T C C C C C	
721	GTTCCGGCTATACCCAAAAGAAGTTAAACCGAATTAACAA	760
	GA A C G G T G C T C	
761	GAGACGTTTAACAGATCCAATTGTCGGAGTCACAAACCT	800
	GC C T C T	
801	TAGGGGCTATGGAACAAACCTCTCTAATATAGAAAATTAT	840
	T T AGC C C C	
841	ATTCGAAAACCACATCTATTGACTATCTGCATAGAACATT	880
	A G C C T C	
881	AATTTCACACGGGTTCCAACCCAGGATATTATGGAAATGA	920
	C AA T C T C T C	
921	CTCTTCAATTATTGGTCCGGTAATTATGTTCAACTAGA	960
	C C C C C C	
961	CCAAGCATAGGATCAAATGATATAATCACATCTCCATTCT	1000
	T T C C C	
1001	ATGGAAATAAATCCAGTGAAACCTGTACAAAATTAGAATT	1040
	T C G G G C C T G	
1041	TAATGGAGAAAAGTCTATAGAGCCGTAGCAAATACAAAT	1080
	C C C G C C C	
1081	CTTGGGGCTGGCCGTCGGCTGTATATTCAAGGTGTTACAA	1120
	C T G A A T C C C	
1121	AAGTGGATTAGCCAATATAATGATCAAACAGATAGAAC	1160
	G G T G C G C G	
1161	AAGTACACAAACGTACCGACTCAAAAAGAAAATGTTGGCGCG	1200
	C C C G T C C T C A	
1201	GTCAGCTGGGATTCTATCGATCAATTGCCCTCCAGAAACAA	1240
	TCT C C	
1241	CAGATGAACCTCTAGAAAAGGGATATGCCATCAACTCAA	1280
	C AT G G C C C T	
1281	TTATGTAATGTCCTTTAATGCAAGGGTAGTAGAGGAAACA	1320
	C G C G A T C C G C	
1321	ATCCCAGTGTAACTGGACACATAAAAGTGTAGACTTTT	1360
	T G C C C G T C G C	
1361	TTAACATGATTGATTCGAAAAAAATTACACAACTTCCGTT	1400
	C C AGC G G C T C	

FIGURE 12B

1401 AGTAAAGGCATATAAGTTACAATCTGGTGCTTCGGTTGTC 1440  
 G G A C C C G  
 1441 GCAGGTCTAGGTTACAGGAGGAGATCATTCAATGCA 1480  
 C A C T T C C G  
 1481 CAGAAAATGGAAGTGC GGCAACTATTACGTTACACCGGA 1520  
 G C C C A T C G T  
 1521 TGTGTCGTA C T C A A A A T A T C G A G C T A T C A T T A T 1560  
 T G G C A G A C T C  
 1561 GCTTCTACATCTCAGATAACATTTACACTCAGTTAGACG 1600  
 A C A G C C C C C G T  
 1601 GGGCACCATTTAATCAACTA T T C G A T A A A C G A T A A A 1640  
 A C C C G T C T C G C C  
 1641 TAAAGGAGACACATTAACCTATAATTCAATTCAATTAGCA 1680  
 C T T C C A C A G C C C G  
 1681 AGTTTCAGCACACCATTCAATTATCAGGGATAACTTAC 1720  
 T C C C C C T C T  
 1721 AAATAGGCGTCACAGGATTAAGTGCTGGAGATAAGTTA 1760  
 G C C T C C C C C C  
 1761 TATAGACAAAATTGAATTATTCCACTGAAT 1791  
 C C G G C C C C

FIGURE 12C

1	ATG	AATAATGTATTGAATAGTGGAAAGAACAACTATTT	40
	GAC	C C C	CTC T C C C
41	GTGATGCGTATAATGTAGTAGGCCATGATCCATTAGTTT	80	
	C C A	C C C G T C	C C
81	TGAACATTAATCATTAGATACCATCCAAAAGAACATGGATG	120	
	C C GAGCC	C C T T G G G	
121	GAGTGGAAAAGAACAGATCATAGTTTATATGTAGCTCCTG	160	
	A C T T C	CTC C C C C A	
161	TAGTCGGAACGTGTCTAGTTTTGCTAAAGAACATGGG	200	
	G T A C C	C C T C G C	
201	GAGTCTTATTGGAAAAAGGATATTGAGTGAATTATGGGG	240	
	CTC C C	C T C T C C C T	
241	ATAATATTCCTAGTGGTAGTACAATCTAACAGATA	280	
	C C ATC	GTCC T C C	
281	TTTTAAGGGAGACAGAACAACTTCTAAATCAAAGACTAA	320	
	C G C	G T C C G C T C	
321	TACAGATACCCTGCTCGTAAATCGAGAATTGATAGGG	360	
	C T T G A A C C T G C T		
361	CTCCAAGCGAATATAAGGGAGTTAACAAACAGTAGATA	400	
	A C T C T	C C G G C	
401	ATTTTTAAACCCCTACTCAAAACCCCTGTCCTTATCAAT	440	
	C C G T A G T	G C T C	
441	AACTTCTCGGTTAACAAATGCAGCAATTATTCCTAAAT	480	
	C C G C T	C C C C C	
481	AGATTACCCCAAGTCCAGATAACAGGATAACCAAGTTTAT	520	
	G T T T C	C C C	
521	TATTACCTTATTGACAGGCAGCCAAATATGCATCTTC	560	
	T C T A C C T T C	C T G	
561	TTTTTATTAGAGATGTTATCTTAATGCAGATGAATGGGT	600	
	C C A C T C G C C C T C	A	
601	ATTTCAAGCAACATTACGTACGTACGATTACCTGA	640	
	C T C T A G A C A	C T	
641	GAAATTATACAAAGAGATTATTCTAATTATTGTATAAAATAC	680	
	G C C T C T	C C C C C	

FIGURE 13A

681	GTATCAA T	ACTCGGTTAGAGGGTTAACACCCGTTACAC G C C T A C C T T A G C T	720
721	GATATGTTAGA C C T	TTAGAACATATATGTTTAAATGTTAT G C G C C C C T C G	760
761	TTGAATATG G C	TATCCATTGGTCATTGGTTAAATATCAGAG CAG AGTC C C G C	800
801	TCTTATGGT CT G	TATCTCTGGCGCTAATTATATGCTAGCGGT G C A C C C C T C T C	840
841	AGTGGACC A T	ACACAGACACAATCATTACAGCACAAA G C G G C C T G	880
881	GGCCATT C G	TTTATATTCTCTTTCCAAGTTAATTG AGCT G C C C C C	920
921	TATATTATCTGGT C TC	ATTAGTGGTACTAGGGCTTCTATTAC CAG CTC G C A C C A	960
961	TTCCCTA T C C	ATATTGGGTTTACCGGGTAGTACTACA ACT A C T A C T C C C	1000
1001	ATTCA AGCC	TGGAAATAGTGC T C T C G G A G G A G T AGCC T CTC A G C C T T	1040
1041	TTCATCTGGT CAGC	CTCATAGGGCG AT G T T A C T G C	1080
1081	TTTAATTG C TC	CAGCACGGT C T C T G A C G A G C	1120
1121	TTGTTAGA G GTCC	AGTTGGCTGGATT T CAGC T C A	1160
1161	CGTTGCT A	ACCTCTACGA A C A C G C	1200
1201	ACAAC C C T	TTAAGGTTAGG CC TC G C T A	1240
1241	GAAATTCAA G C T	ACTATTTCC C C C C T A G C	1280
1281	TTCTGGG C T	TTCTTACTGTTATTAG C C C C G T C C C	1320
1321	AGACCGT C T	ACTATAACCA ACT T T C G T G C C G T C	1360
1361	CTTCGG A C T	AAACACCTGGGAG T T A A T C C C G	1400

FIGURE 13B

1401	TGTGCATAACAGAAAAATAATATCTATGCCGCTAATGAA	1440
	C G G C C C T C C G	
1441	AATGGTACTATGATCCATTGGCGCAGAAGATTATACAG	1480
	C C T C C T A C T	
1481	GATTTACTATATGCCAATACATGCCACTCAAGTGAATAA	1520
	C C C T C T C C	
1521	TCAAACCTGAACATTTATTCTGAAAAATTGGAAATCAA	1560
	G A C C C C C G C	
1561	GGTGATTCTTAAGATTGAACAAAGCAACACGACAGCTC	1600
	C G G C G T C T C A	
1601	GTTATACCGTTAGAGGGAATGAAATAGTTACAATCTTA	1640
	G C T T G C C C C	
1641	TTTAAGAGTATCTTCAATAGGAAATTCAACTATTGGAGTT	1680
	C G TAGC C T T C C C C T	
1681	ACTATAAACGGTAGAGTTTACTGTTCAAATGTTAATA	1720
	C C A C T C A C T G C	
1721	CCACTACAAATAACGATGGAGTTAATGATAATGGAGCTG	1760
	T A G C T C C C C C A	
1761	TTTTCAAGATATTAATATCGGTAATATAGTAGCAAGTGT	1800
	A C A G C C C T C C C G C T C	
1801	AATACTAATGTAACCGTAGATATAAATGTGACATTAACT	1840
	C C T T G C C C C G T	
1841	CCGGTACTCCATTGATCTCATGAATATTATGTTGTGCC	1880
	T A C C C	
1881	AACTAATCTTCCACCACTTTAT	1902
	C C T T G C	

FIGURE 13C

1	ATGGAGGAAAATAATCAAATCAATGCATAACCTTACAATT	40
	G C C   C   T A C	
41	GTTTAAGTAATCTGAAGAAGTACTTTGGATGGAGAACG	80
	C G   C A   G   T G C T	
81	GATATCAACTGGTAATTCTCAATTGATATTCTCTGTCA	120
	C T   C   C T C C C C C T C	
121	CTTGTTCAGTTCTGGTATCTAACCTTGATACCAGGGGAG	160
	T G C   CAGC   C G T T	
161	GATTTTAGTTGGATTAATAGATTTGTATGGGGATAGT	200
	G C C T C C T C C C C T C	
201	TGGCCCTCTCAATGGATGCATTCTAGTACAAATTGAA	240
	T A           C G G   G	
241	CAATTAATTAAATGAAAAGATAGCTGAATTGCTAGGAATG	280
	G G C C G G C   G C C C C	
281	CTGCTATTGCTAATTAGAAGGATTAGGAAACAATTCAA	320
	C C   C G   G C T C	
321	TATATATGTGGAAGCATTAAAGAATGGGAAGAAGATCCT	360
	C C   G C C   G   G C	
361	AATAATCCAGAAAACCAGGACCAGAGTAATTGATCGCTTC	400
	C G C C T G G C C A A C A	
401	GTATACTTGATGGCTACTTGAAAGGGACATTCTTCGTT	440
	A C T G C C C T G G A T C A C	
441	TCGAATTCTGGATTGAGTACCCCTTTATCCGTTTAT	480
	C A   C C   T T C G   G C	
481	GCTCAAGGCCAATCTGCATCTAGCTATATTAAAGAGATT	520
	A T   T C C   C C T C C A	
521	CTGTAATTGGAGAAAAGATGGGGATTGACAACGATAAA	560
	G C C   G   G           C T C	
561	TGTCAATGAAAACATAATAGACTAATTAGGCATATTGAT	600
	C   G T C C   T C C C C	
601	GAATATGCTGATCACTGTGCAAATACGTATAATCGGGAT	640
	G C C C   T C C C C T C	
641	TAAATAATTACCGAAATCTACGTATAACAGATTGGATAAC	680
	G C C C T G   T           T	
681	ATATAATCGATTACGGAGAGACTTAACATTGACTGTATTA	720
	C C C A G   G   G C C C A T G	

FIGURE 14A

721	GATATGCCGCTTCTTCCAACTATGACAATAGGAGAT	760
	C T A . . . C G . . . C	
761	ATCCAATTCAAGCCAGTTGGTCAACTAACAAAGGGAAAGTTA	800
	C T C A . . . G . . . T C A . . . C	
801	TACGGACCCATTAATTAATTTAACGTTACAGTTACAGTCT	840
	T . . . C T . . . C C C T . . . G A A G	
841	GTAGCTCAATTACCTACTTTAACGTTATGGAGAGGAGCC	880
	C C C T C A C . . . C . . . T C	
881	GAATTAGAAAATCCTCATTATTTGATATATTGAATAATCT	920
	T C G C A C G . . . C C . . . C C	
921	TACAATCTTACGGATTGGTTAGTGGTGGACGCAATTCT	960
	T . . . C C . . . C C . . . G T C C C	
961	TATTGGGGAGGACATCGAGTAATATCTAGCCTTATAGGAG	1000
	T . . . C A G . . . C C T C T . . . T	
1001	GTGGTAACATAACATCTCTATATATGGAAAGAGAGGGCAA	1040
	G . . . T C . . . C . . . C T A	
1041	CCAGGAGGCCCTCCAAGATCCTTACTTTAACGGACCGTA	1080
	A . . . C T A G T . . . G C C C C T A C	
1081	TTTAGGACTTTATCAAATCCTACTTTACGATTATTACAGC	1120
	C A C G T C . . . C G A . . . G C C .	
1121	AACCTTGGCCAGGCCACCATTTAACGGTGGTGTGA	1160
	T . . . T C C C T A	
1161	AGGAGTAGAATTCTCACACCTACAAATAGCTTACGTAT	1200
	G C T G C . . . T C C T C C T C	
1201	CGAGGAAGGGTACGGTGATTCTTAACTGAATTACCGC	1240
	A . . . T A C . . . C G C C C A	
1241	CTGAGGATAATAGTGTGCCACCTCGCGAAGGATATAGTC	1280
	A C . . . C A G . . . C C T C C	
1281	TCGTTTATGTCACTGGTAGTAAAGATCTGGAAACA	1320
	C A G G C C . . . C C G G C T C . . . T	
1321	CCTTTTTAACAACTGGTAGTATTTCTGGACCGATC	1360
	A C C C T A A T G C A . . . T	
1361	GTAGTGCAACTCTTACAAATACAATTGATCCAGAGAGAAT	1400
	T C T . . . C . . . C G	

FIGURE 14B

1401	TAATCAAATACCTTACTGAAAGGATTAGAGTTGGGG C C A G C G T C C T G A	1440
1441	GGCACCTCTGTCAATTACAGGACCAGGATTACAGGAGGG A T C C C T	1480
1481	ATATCCTCGAAGAAATACCTTGGTATTGTATCT T A C T C C GAGC	1520
1521	ACAACTCAATTAAATTACCAATTACCAAAGATACCGT C T C C C T T T	1560
1561	TTAAGATTTCGTTACGCTTCCAGTAGGGATGCACGAGTTA C C G A TTCC C T C TA C	1600
1601	TAGTATTAAACAGGAGCGGCATCCACAGGAGTGGGAGGCCA C GC C C C A T T C T C T A	1640
1641	AGTTAGTGTAAATATGCCCTTCAGAAAATATGAAATA CTCC G C A C G G C	1680
1681	GGGGAGAACTTAACATCTAGAACATTAGATATACCGATT C G C G C C C	1720
1721	TTAGTAATCCTTTTCAATTAGAGCTAATCCAGATAATAAT CTC C CAGT CC T C C T C C	1760
1761	TGGGATAAGTGAACAACTCTATTGGTGAGGTTCTATT C T C C A T AGC C	1800
1801	AGTAGGGTGAACTTTATATAGATAAAATTGAAATTATTC TCATCT C T G C T C G G C	1840
1841	TACCAAGATGCAACATTGAAGCAGAACCTGATTTAGAAAG T C C T C C G T G A C A C C T G	1880
1881	AGCACAAAAGGCGGTGAATGCCCTGTTACTCTTCCAAT C G T C C C C C A	1920
1921	CAAATCGGGTAAAAACCGATGTGACGGATTATCATATTG G C T C G T A C T T C C	1960
1961	ATCAAGTATCCAATTAGTGGATTGTTATCAGATGAATT C G C G C A C C A C C T A G C	2000
2001	TTGTCTGGATGAAAAGCCAGAATTGTCGGAGAAAGTCAAA C C C C G T C C T	2040
2041	CATGCGAAGCGACTCAGTGTAGAGCGGAATTACTTCAG C C T C C A C C T G	2080
2081	ATCCAAACTCAGAGGATCAATAGACAAACCAGACCGTGG CT C A A C C G G A	2120

FIGURE 14C

2121	CTGGAGAGGAAGTACAGATATTACCATCCAAGGAGGAGAT	2160
	T G T C C G G C C C	
2161	GACGTATTCAAAGAGAATTACGTCACACTACCGGGTACCG	2200
	T G G C C C T C A T T	
2201	TTGATGAGTGTATCCAACCTATTATATACGAAAATAGA	2240
	C C C T C C G C G C	
2241	TGAGTCGAAATTAAAGCTTACCCGTTATGAATTAGA	2280
	C C C C T C A G C C T	
2281	GGGTATATCGAAGATAGTCAGACTTAGAAATCTATTGAG	2320
	C C C C T C T C C	
2321	TCCGTTACATGCAAACACGGAAATAGTAAATGTGCCAGG	2360
	A G C G G C G C	
2361	CACGGGTTCCCTATGGCCGCTTCAGGCCAATGCCAAC	2400
	T T C C A T T C T C T C T	
2401	GGAAAGTGTGGAGAACCGAATCGATGCGCCACACCTTG	2440
	G G T C A T	
2441	AATGGAATCTGATCTAGATTTCTGAGAGACGGGGAA	2480
	G C T G C C G T C	
2481	AAAATGTGCACATCATCCCATCATTCACTTGGATATT	2520
	G G C C T C T C C	
2521	GATGTTGGATGTACAGACTTAAATGAGGACTTAGGTGAT	2560
	G T C G C C A C	
2561	GGGTGATATTCAAGATTAAGACGCAAGATGGCATGCAAG	2600
	C C C C C A C	
2601	ACTAGGGAATCTAGAGTTCTGAAAGAGAACCATTTATT	2640
	T C C T G G C	
2641	GGGGAAAGCACTAGCTCGTGTGAAAGAGCGGAGAAGAAGT	2680
	T T C G A A	
2681	GGAGAGACAAACGAGAGAAACTGCAGTTGAAACAAATAT	2720
	G T C G A G T C	
2721	TGTTTATAAGAGGCAAAAGAAATCTGATGCTTTATT	2760
	C C G C C G G G C	
2761	GTAAACTCTCAATATGATAGATTACAAGTGAGATACCAACA	2800
	G C C A G G C C C	
2801	TCGCCATGATTGATCGCGGAGATAAACCGGTTCATAGAAT	2840
	C C C C T G C C	

FIGURE 14D

2841	CCGGGAAGCGTATCGCCAGAGTTGTCGTGATTCCAGGT	2880
	T T G T C T T C C T	
2881	GTCAATGCCGCATTTGAGAATTAGAGGGACGTATT	2920
	G C T C G G C T C	
2921	TTACAGCGTATTCCTTATATGATGCCAGAAAATGTCATTAA	2960
	C A T C G C C C C C	
2961	AAATGGCGATTCAATAATGGCTTATTATGCTGGAACGTG	3000
	G C T C C C C A G C T	
3001	AAAGGTCAATGAGATGTAGAAGAGC AAAACAACCACCGTT	3040
	G C G G A G T G	
3041	CGGTCCCTGTTATCCAGAATGGAGGGCAGAAGTGTCA	3080
	C G G G T G A T C	
3081	AGAGGTTGGTGTCTGCCAGGTGGTGTGCTATATCCTTCGT	3120
	X A A A C T C	
3121	GTCACAGCATATAAAGAGGGATATGGAGAGGGCTGGTAA	3160
	G C T C G C T T G	
3161	CGATCCATGAGATCGAAGACAATAACAGACGA	3200
	ACTGAAATT	
3201	CAGCAACTGTGAGAAGAGGA	3240
	AGTATATCCAAACACACA	
3241	GTAACGTGTAATAATTATACTGGGACTCAAGAAGAATATG	3280
	T T C C G C C T A G G C	
3281	AGGGTACGTACACTTCTCGTAATCAAGGATATGACGAAGC	3320
	G A G C A G C T C A	
3321	CTATGGTAATAACCTTCGGTACCGAGCTGATTACGCTTCA	3360
	T C C T C X	
3361	GTCTATGAAAGAAAATCGTATACAGATGGACGAAGAGAGA	3400
	G C G G C C C A C T	
3401	ATCCTTGTGAATCTAACAGAGGGCTATGGGATTACACACC	3440
	C C G T C T C A C C	
3441	ACTACCGGCTGGTTATGTAACAAAGGATTAGAGTACTTC	3480
	T A T C T C G C T T	
3481	CCAGAGACCGATAAGGTATGGATTGAGATCGGAGAACAG	3520
	T C A G C T C	
3521	AAGGAACATTCACTGTGGATAGCGTGGATTACTCCTTAT	3560
	G C C G C T T G	
3561	GGAGGAA 3567	

FIGURE 14E

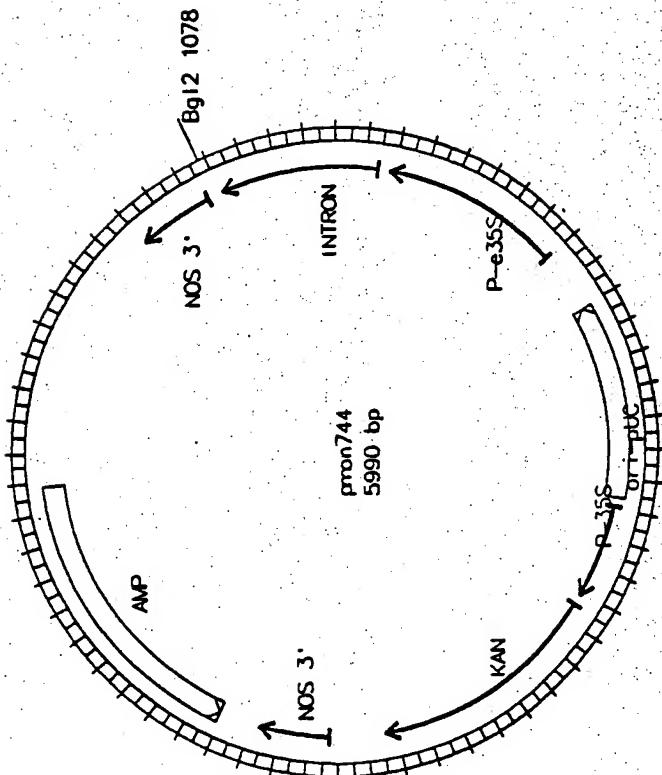


FIGURE 15

1	AGATCTAGAGGTAA	TGTTATGAGTACTGTCGTGGTTAAG	40
		GATC	
41	GGAAACGTCAACGGTGGT	TGACAACAACCTAGAAGGGAGGA	80
	G	T	A
81	GAAGGCAATCCCTTCG	CAGGAGGGCTAACAGAGTACAGCC	120
	T	A	T
121	AGTGGTTATGGTCA	CTGCTCCTGGCGAACCCAGGGAGG	160
	GC	A A	A
161	AGACCGCAGAAGAGGAGG	AGCAATCGCAGGTCAAGAAGAACTG	200
	A G	T	A
201	GAGTTCCCAGGGGAA	AGGGGCTCAAGCGAGACATTGTGTT	240
	A	A T	
241	TACAAAGGACAAC	CTCGTGGCAACTCCAAAGGAAGTTTC	280
281	ACCTTCGGACCA	AACTGTATCAGACTGTCCAGCATTCAAGG	320
		T	
321	ATGGAATACTCAAGG	CCTACCATGAGTACAAGATCACAAG	360
		T	
361	TATCCTTCTTCAG	TGGTCAAGCGAGGCCTCTCCACCTCA	400
	T G	T	
401	CCAGGATCCATCGCTT	TATGAGTTGGACCCACATTGCAAAG	440
	C	A T	
441	TATCATCCCTCCAG	TCTACGTCAACAAGTTCAAATCAC	480
	T		
481	AAAGGGAGGAGCTA	AGACCTATCAAGCTAGGATGATCAAC	520
	T T	C T	
521	GGAGTAGAATGGCACG	ATTCATCTGAGGATCAGTGCAGGA	560
	T	T	A
561	TACTTGGAAAGGAAGT	GGAAAATCTCAGACCCAGCAGG	600
	C	A G	T T
601	ATCTTCAGAGTCACCA	TCAAGACTGGCTTCAAAACCCC	640
	T	T	A
641	AAGTAATAGACTCCGG	ATCAGAGCCTGGTCCAAGCCCACA	680
	A T		

FIGURE 16A

681 ACCAACACCCACTCCAACCTCCCCAAAAGCATGAGCGATT 720  
721 ATTGCTTACGTCGGCATACTATGCTGACCATTCAAGAAT 760  
761 TC 762

FIGURE 16B